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1 **REVIEW**

2 **A review of chronic wasting disease in North America with**
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5 **implications for Europe**

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17 **Abstract** Cervids are keystone species in ecosystems and are associated with enormous
18 cultural and economic value. Chronic wasting disease (CWD) is a fatal prion disease
19 spreading in North American cervid populations. The 2016 emergence of CWD in Europe
20 makes it urgent to understand the basics of CWD and to assess the extent to which current
21 CWD knowledge is transferable to Europe. CWD is difficult to detect in the early stages due
22 to very low prevalence and slow growth rates. The negative population effect of CWD is
23 mainly due to increased female adult mortality, as infected individuals continue to reproduce.
24 It may take decades before CWD leads to population declines. The population dynamics of
25 mule deer are affected more by CWD than those of white-tailed deer, which in turn are more
26 affected than those of elk, and depending on other factors limiting the populations. Species-
27 and population-specific differences in dynamical consequences are linked to the balance
28 among the rates of transmission, incubation period (linked to the prion protein gene, *PRNP*),
29 and reproductive rates. This make it difficult to predict effects of CWD in Europe with other
30 cervids, but the dynamic impact may be marked to cervid populations over the long term. The
31 process of spillover across the species barrier is not well understood. Occasional spillover to
32 moose without an apparent epizootic suggests specific conditions can limit CWD. Frequency-
33 dependent transmission or weak density-dependent transmission makes it difficult to control
34 CWD using density reductions through harvest and/or culling. CWD is difficult to eradicate
35 once it becomes endemic, and it calls for immediate management actions. These actions
36 involve extensive culling, fencing and ceasing of wildlife feeding and are likely to cause
37 significant controversy.

38 **Key words** Frequency-dependent versus density-dependent transmission • direct and
39 environmental transmission routes • spatially targeted harvesting • extermination and
40 fallowing • salt licks and supplemental feeding • genetics and pathology • epizootiology and
41 population dynamics

42 **Introduction**

43 The first case of chronic wasting disease (CWD) in Europe was diagnosed in March 2016 in a
44 female reindeer (*Rangifer tarandus*) in the Nordfjella mountains, Norway (Benestad et al.
45 2016). Since then, several more CWD-infected reindeer from the same population were
46 detected by testing during the 2016 and 2017 hunting seasons (Viljugrein et al. 2018). Hence,
47 we have the first reported outbreak of CWD in Europe. CWD was first documented in a
48 captive mule deer (*Odocoileus hemionus*) in 1967 in Colorado, USA (Williams & Young
49 1980), and it appeared in wild mule deer in 1981 (Williams & Young 1992; Spraker et al.
50 1997; Miller et al. 2000). CWD in the wild has since spread to 25 states and, through sales of
51 farmed elk, has been introduced to two Canadian provinces and to South Korea (Uehlinger et
52 al. 2016); however, the origin of CWD in Norway remains unknown (Benestad et al. 2016).

53 It is important to realize that although CWD was first identified among wild deer in 1981, it is
54 still spreading to new areas and continuing to increase in prevalence in most, if not all,
55 endemic areas. Evidence of declining populations in endemic areas are recently reported for
56 white-tailed deer (*Odocoileus virginianus*) (Edmunds et al. 2016) and mule deer (DeVivo et
57 al. 2017). There are also increasing impacts on elk (*Cervus canadensis*) populations (Monello
58 et al. 2014; Monello et al. 2017), which is a closely related species to the European red deer
59 (*Cervus elaphus*). Due to the timing and slow rise in prevalence of CWD, it seems likely that
60 more such reports of population declines will appear in the coming years.

61 Due to the immediate risk of CWD becoming endemic in Norway and spreading
62 geographically in Europe, it is important to know what to expect in affected populations. To
63 what extent are CWD dynamics sufficiently understood in North America? Will CWD
64 prevalence always increase in a population or does it require specific conditions to do so?
65 What are the expected population impacts and how fast will they appear? Will CWD spill
66 over across cervid species, and if so, will the population impacts be species-specific? To what

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67 extent is the current CWD knowledge transferable to Europe? How will CWD change
68 surveillance and cervid management in Europe? We try to give an initial answer to these
69 difficult questions and to highlight the gaps in knowledge to guide research and management
70 efforts.

71 **What are we up against?**

72 The disease agents of CWD are proteinaceous infectious particles called prions (PrP^{CWD}), and
73 hence CWD groups with other prion diseases such as bovine spongiform encephalopathy
74 (BSE), scrapie in sheep, and Creutzfeldt–Jakob disease (CJD) in humans (Prusiner 1998).
75 Prion diseases are invariably fatal, and there are no vaccines or treatments currently available.
76 Susceptibility to CWD is linked to similarity in the structure of the prion protein (PrP), which
77 is present in all mammals. A prion causes misfolding of the normal cellular prion protein
78 (PrP^C) into a form (PrP^{res}) not degraded in the organism, which in turn causes a chain-reaction
79 of further misfoldings (Robinson et al. 2012b). Aggregates of PrP^{res} constitutes the prion. The
80 structure of PrP is determined by the prion protein gene (*PRNP*), which is highly conserved
81 and with few polymorphisms within cervids. In general, most cervids are therefore considered
82 susceptible to various degrees, while susceptibility of some species are not determined
83 (Robinson et al. 2012b). The importance of genetics is well covered elsewhere (Robinson et
84 al. 2012b), and we here only cover *PRNP* variation as it relates to population dynamic
85 impacts. Prion diseases are usually not very contagious, the exception being CWD in cervids
86 and ‘classical’ scrapie in sheep. A type of ‘non-classical’ scrapie in sheep occur as a sporadic
87 disease mainly in old animals (Benestad et al. 2003). In addition to the ‘classical’ type of
88 CWD, a new type of CWD was found in two moose in 2016 in Norway, one more in 2017,
89 and also in a moose in 2018 in Finland (Pirisinu et al. 2018). A ‘non-classical’ type of CWD
90 was also confirmed in one red deer in 2017 in Norway (Våge et al. 2018). The moose cases

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91 appear unrelated to the reindeer cases (Pirisinu et al. 2018). We will focus our review on what
92 can be termed ‘classical’ CWD, which is highly contagious to be a prion disease.

93 **Transmission routes**

94 CWD can be transmitted both directly from animal to animal (Miller & Williams 2003) and
95 indirectly through the environment (Miller et al. 2004). Though vertical transmission from
96 mother to offspring may occur (Nalls et al. 2013), it is regarded as of minor importance for
97 the epizootic characteristics (Miller & Williams 2003). A major knowledge gap preventing
98 understanding, managing and modelling the development of CWD, is that the quantitative
99 importance of various transmission routes is uncertain. It is likely that direct animal-to-animal
100 contact is the main route of infection in early stages of CWD, while environmental
101 transmission becomes more important in later stages as prions build up in the environment
102 (Almberg et al. 2011). Direct contact is typically higher within than between social groups
103 (Schauber et al. 2015), suggesting that the level of female sociality can be important for
104 transmission. Direct contact of genetically related females, which typically have overlapping
105 home ranges, is a risk factor leading to a higher prevalence of CWD (Gear et al. 2010;
106 Cullingham et al. 2011a). Direct contact rates among females were higher during the rut
107 (autumn) and lowest during summer (Kjær et al. 2008). Female-male contact is highest during
108 rut, while male-male contact is typically higher during summer and pre-rut when social rank
109 is determined. Any action that limits artificial aggregation of cervids is likely to reduce
110 transmission.

111 **Density-dependent or frequency-dependent transmission?**

112 Disease dynamics are affected by the mode of transmission, which can be either density- or
113 frequency-dependent at the population level. Density-dependent infectious diseases are easier
114 to control in wildlife populations because culling efforts can limit transmission. With
115 frequency-dependent transmission, CWD could only be eliminated by removal of the infected

116 population (Wasserberg et al. 2009), since the lack of clinical signs in the early stages impede
117 selective harvest of infected individuals. Several lines of evidence have been used to evaluate
118 whether CWD is likely to have density-dependent or frequency-dependent transmission. Most
119 evidence suggests CWD has close to frequency-dependent transmission (Table 1). Frequency-
120 dependent transmission is typical of transmission in socially regulated contact networks. The
121 proximity of animals, based on evidence from GPS-marked animals and grouping patterns of
122 deer, is related to population density. However, proximity of animals may not measure the
123 actual direct contact rates necessary for transmission, and hence it is uncertain whether such
124 kind of data can be used to infer density-dependent transmission of CWD. In any case, such a
125 weak impact of population density on transmission does not support culling to reduce density
126 as a tool to control CWD in Europe. Rather, host eradication is required for diseases with
127 frequency-dependent transmission (Wasserberg et al. 2009). The transmission mode for CWD
128 in reindeer is unknown. However, the expectation that transmission would be close to
129 frequency-dependent was an important part of the basis for the aim to remove the whole herd
130 of over 2000 reindeer infected with CWD in Norway (Hansen et al. 2016), which is now
131 completed (Mysterud & Rolandsen 2018).

Table 1 A brief overview of four lines of evidence for whether chronic wasting disease has a
density-dependent (DD) or frequency-dependent (FD) mode of transmission at the population
level.

Type and approach of study	Parameter or type of data	Mode of transmission	Reference
Mathematical modelling of transmission modes based on empirically estimated functions	Output from transmission models compared to demographic pattern of CWD infection	FD fit data better	(Wasserberg et al. 2009; Jennelle et al. 2014)
Empirical observations of contact rates	Contact rates among GPS-marked animals; group sizes across a population density range	DD or intermediate, season-specific	(Habib et al. 2011; Cross et al. 2013)

1 2 3 4	Empirical observations of CWD prevalence	Analysing spatial variation in CWD prevalence and the relationship to population density	DD at low density, FD at high density	(Storm et al. 2013)
5 6 7 8 9	Culling efforts by state/provincial wildlife agencies	Analysing variation in CWD prevalence before and after management efforts to reduce population density	FD	Reviews in (Conner et al. 2007; Uehlinger et al. 2016)

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12 136 **Epizootic characteristics of CWD**

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14 137 The time from infection to death in the case of CWD is typically 1.5-2.5 years in white-tailed

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16 138 deer and mule deer (Fox et al. 2006; Robinson et al. 2012a), but can be as long as 4 years in

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18 139 elk (Moore et al. 2018); depending on *PRNP*-genotypes. In mice models, also the PrP^{CWD}

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20 140 strain play a role for duration of infection and transmission (Raymond et al. 2007; Angers et

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22 141 al. 2010; Perrott et al. 2012). A long time from infection to death is typical of prion diseases.

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24 142 The basic reproductive number (R_0) measures how fast diseases transmit and grow in a

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26 143 population; it is the expected number of new individuals infected by an infected individual. In

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28 144 captive mule deer, the R_0 values for CWD were determined to be 1.3 and 1.5 in two different

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30 145 epizootics (Miller et al. 2006), and a substantial increase in prevalence may take decades

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32 146 (Wasserberg et al. 2009). CWD in white-tailed deer in endemic areas of Wyoming has now

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34 147 reached a prevalence of 30-40% an estimated 35 years after introduction (Edmunds et al.

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36 148 2016). However, the R_0 was much higher (in the range of 2.2 to 4.5) even in the early stages

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38 149 of CWD outbreak among mule deer in Alberta, Canada (Potapov et al. 2015). In modelling

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40 150 studies, the estimated R_0 values rarely reach above 2-3 when direct transmission is assumed,

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42 151 but the R_0 can reach considerably higher values with environmental transmission (Almberg et

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44 152 al. 2011; Sharp & Pastor 2011). Other modelling of CWD dynamics suggested that it was not

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46 153 given that R_0 will be >1 under all conditions (Miller et al. 2006), which is required to

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48 154 establish an epizootic. However, the empirical basis for many model parameters are still often

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50 155 weak or absent, and results should be interpreted with caution.

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156 **Spatial pattern of CWD in North America**

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2 157 Management actions, environmental conditions, and the properties of the affected cervid
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5 158 populations may all influence the magnitude to which CWD will affect a given population.
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7 159 Several management actions aimed at limiting CWD have been implemented in Colorado,
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10 160 Wisconsin, and Illinois and the Canadian provinces of Alberta (Uehlinger et al. 2016) and
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12 161 Saskatchewan (Cullingham et al. 2011b), whereas Wyoming has mainly implemented CWD
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14 162 surveillance with no direct action. CWD appear successfully eradicated from New York after
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17 163 detection in two captive and two wild white-tailed deer (Evans et al. 2014). However, spatial
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19 164 variation in infection rates and whether management has been successful in limiting CWD
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22 165 elsewhere in North America remains unclear. It is difficult to estimate prevalence of CWD
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24 166 empirically in the initial stages due to very low prevalence and imperfect detectability
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27 167 (Viljugrein et al. 2018), and hence large sample sizes are required for detecting temporal
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29 168 changes and spatial variation in CWD prevalences. Therefore, if CWD prevalence has
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32 169 remained low for a long period (Geremia et al. 2015), it is difficult to determine whether
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34 170 prevalence is stable, or whether sample sizes are insufficient to detect changes. Changes in the
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37 171 size of monitoring areas are also a problem affecting estimation of prevalence. A formal
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39 172 analysis could not detect spatial variation in the growth of CWD prevalence in Wisconsin
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41 173 (Heisey et al. 2010). The changes in CWD prevalence were determined primarily by the time
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44 174 point of disease introduction (Heisey et al. 2010).

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47 175 **Demographic patterns of CWD prevalence**

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49 176 The observed demographic pattern of CWD infection in a given area results from a
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52 177 combination of time available for exposure and low detectability in early infectious stages,
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54 178 while age- and sex differences in behaviour may lead to different exposure. Prevalence is very
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57 179 low in fawns or calves, and yearlings have about half the infection levels as adults (Miller &
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59 180 Conner 2005; Samuel & Storm 2016). The low infection levels in juveniles may reflect a
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181 shorter time of exposure and less exposure to environmental contamination (when they
182 suckle), in combination with the delay between exposure to PrP^{CWD} and detection using
183 standard CWD tests (Viljugrein et al. 2018). New detection methods are now under rapid
184 development and becoming more sensitive to the early stages of infection (Haley & Richt
185 2017). Nevertheless, yearlings had lower infection prevalence than adults even after
186 accounting for time of exposure (Samuel & Storm 2016). In both mule deer (Miller & Conner
187 2005) and white-tailed deer (Heisey et al. 2010), the prevalence of CWD peaked at ages 5-6
188 years in males, but this result may have been due to biases in age estimation based on tooth
189 wear (Samuel & Storm 2016). CWD prevalence in deer is generally approximately twice as
190 high in males as in females (Miller & Conner 2005; Gear et al. 2006). The higher
191 prevalences in males is likely linked to behavioural differences affecting exposure, but how is
192 not understood. The pattern was slightly reversed between the sexes for white-tailed deer in
193 Wyoming, where the prevalence was as high as 28.8% in males and 42% in females
194 (Edmunds et al. 2016). The extent to which the demographic pattern of infection changes in
195 late epizootic stages remains uncertain.

196 **CWD, mortality and reproduction**

197 Empirical evidence does not indicate markedly reduced reproductive rates in CWD-infected
198 individuals that are pre-clinical for either mule deer (Dulberger et al. 2010a) or white-tailed
199 deer (Blanchong et al. 2012; Edmunds et al. 2016). All CWD infected animals die of clinical
200 disease if they live long enough, and increased adult female mortality is the main effect of
201 CWD on population dynamics (Dulberger et al. 2010b; Edmunds et al. 2016). The effect on
202 populations will further depend on whether mortality from CWD is additive or compensatory
203 to other causes of mortality. Mortality from CWD is in part compensatory to other mortality
204 sources in areas with selective predation (Krumm et al. 2010), hunting (Conner et al. 2000) or
205 accidents (Krumm et al. 2005) of CWD-infected individuals, but a sufficiently large part of

206 mortality caused by CWD is additive leading to population limitation. Any limiting factor
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2 207 affecting adult female mortality will have the greatest impact on large herbivore populations
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4 208 (Gaillard et al. 1998), and CWD hence has the potential to modulate population dynamics at
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7 209 high prevalence (Edmunds et al. 2016; DeVivo et al. 2017).
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10 210 **Population dynamic effects of CWD**

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13 211 The effect of CWD on populations is driven mainly by the balance between the time since
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15 212 infection, the rate of transmission, the incubation period (linked to the *PRNP* gene), how
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17 213 quickly new offspring (without infection) are produced (Potapov et al. 2016), and it will
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20 214 depend on other limiting factors in a given area. Once CWD is established, prevalence among
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22 215 adult females will rise slowly, to an increasing degree limit population growth, and over
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25 216 decades cause a gradual population decline that may become substantial (DeVivo et al. 2017).
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27 217 The impacts vary between species and geographic location. Individuals or species with host
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30 218 genotypes that are associated with lower susceptibility and longer incubation periods
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32 219 (O'Rourke et al. 1999; O'Rourke et al. 2004; Jewell et al. 2005; Moore et al. 2018) can
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35 220 produce more offspring before death, slowing the rise in CWD prevalence (Table 2).
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37 221 Similarly, the population dynamical consequences will be lower for species and populations
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40 222 with higher reproductive rates, diluting the prevalence by rapidly adding new non-infected
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42 223 individuals to the population (Potapov et al. 2016).
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45 224 Population dynamic effects are larger in mule deer than in white-tailed deer, while elk
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48 225 populations are less affected. In endemic areas of Wyoming, CWD led to a 10.4% annual
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50 226 population decline in white-tailed deer (Edmunds et al. 2016) and a 21% annual decline in
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52 227 mule deer (DeVivo et al. 2017). The lower reproductive potential of mule deer may explain
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55 228 the larger population effects of CWD compared to in white-tailed deer populations. Elk
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58 229 populations have consistently lower CWD prevalence (Miller et al. 2000). In elk, a CWD
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60 230 incidence above 15% is not reported. The annual incidence of CWD was estimated at 0.08
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231 [0.05-0.12] in a high-density elk herd in the Rocky Mountains, USA, after 25 years (Monello
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2 232 et al. 2014). The elk population in Wind Cave National Park, South Dakota had a prevalence
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4 233 reaching 14% (12-15%) in adults during the winter of 2016-2017 (Glen Sargeant, pers.
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7 234 comm.). CWD develops slower, with longer incubation periods (Moore et al. 2018), in elk
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9 235 than in deer (Race et al. 2007); this was assumed due to the substitution in the *PRNP* gene at
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11 236 residue 226 (Angers et al. 2010). However, even in elk, the CWD prevalence may rise
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14 237 sufficiently to become population limiting. In Colorado, population declines were predicted at
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17 238 13% [0-35%] adult female prevalence for elk (Monello et al. 2014) and at 26% for white-
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19 239 tailed deer in Wyoming (Edmunds et al. 2016).
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22 240 Over longer time scales, less-susceptible host genotypes may become more common and
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25 241 dampen the population effects of CWD (Williams et al. 2014; DeVivo 2015; Monello et al.
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27 242 2017). The very long-term effects (century scale) are currently not known, and it is therefore
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30 243 unclear if CWD will cause local extinctions.
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33 244 Predicting the population dynamic impact of CWD on European cervids without any
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35 245 empirical evidence is uncertain even to a very coarse level (Table 2). The variation in the
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38 246 *PRNP* gene is low in moose and roe deer, while it is somewhat higher in red deer and reindeer
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41 247 leading to a potential for larger individual variability in susceptibility. Moose and roe deer are
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43 248 generally more solitary than other cervids. Moose have quite high and roe deer have very high
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45 249 reproductive rates (Table 2). Both these factors may limit the growth of CWD in a population.
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48 250 Anecdotal evidence from North America suggests occasional spillover to moose (Baeten et al.
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50 251 2007; Haley & Hoover 2015), but it is uncertain whether the absence of subsequent epizootics
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53 252 is due to the solitary behaviour of moose alone. Group sizes typically increase with the
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55 253 openness of the habitat (Pays et al. 2007), with increasing population density (Vincent et al.
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57 254 1995), and aggregation in agricultural fields or at supplementary feeding sites can increase
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Table 2. An overview of susceptible species' risk factors related to the effects of chronic wasting disease (CWD) on population dynamics. Variation in social organization and population growth rates are considerable within species. The normal incubation period is considered 1.5-2.5 years, but can be longer. There are varying levels of susceptibility based on host prion protein gene (*PRNP*) allele variation (Robinson et al. 2012b; EFSA Panel on Biological Hazards (BIOHAZ) et al. 2016). For annual population growth rates, we based this on the maximum expected growth from Gaillard et al. (2000), and pers. comm.), mainly varying depending on the number of offspring produced; such population growth rates will depend on the ecological conditions in a given area. Amino acid abbreviations: A = alanine; E = glutamic acid; F = phenylalanine; G = glycine; H = histidine; I = isoleucine; L = leucine; M = methionine; P = proline; Q = glutamine; S = serine; T = threonine; V = valine.

Continent Species	<i>PRNP</i> allele variation	Pathology, incubation	Social organization	Maximum population growth rates (λ_{max})	Population impact
North America					
Mule deer <i>Odocoileus hemionus</i>	3 <i>PRNP</i> genotypes (Jewell et al. 2005; Robinson et al. 2012b): 225SS, SF, FF; susceptibility varies by genotype	Normal (Race et al. 2007); <i>PRNP</i> genotype differences vary by incubation period ³	Large groups	Intermediate (1.40-1.45)	Moderate (Geremia et al. 2015) to large impact (Dulberger et al. 2010a)
White-tailed deer <i>Odocoileus virginianus</i>	9 <i>PRNP</i> genotypes (O'Rourke et al. 2004; Velásquez et al. 2015): 95QQ, QH, HH ; 96GG, GS, SS ; 116AA, AG, GG (O'Rourke et al. 2004); susceptibility varies by genotype	Normal (Race et al. 2007), 1.8-2.6 years; incubation period vary by <i>PRNP</i> genotype (Johnson et al. 2011)	Small familial groups; larger groups in winter in northern latitudes	Very high (> 1.6)	Intermediate impact; normal incubation but rapid population growth; may cause population declines (Edmunds et al. 2016; Foley et al. 2016).
Elk <i>Cervus canadensis</i>	3 <i>PRNP</i> genotypes (O'Rourke et al. 1999; Robinson et al. 2012b): 132MM, ML, LL	Slow (Race et al. 2007); incubation period vary by <i>PRNP</i> genotype (Moore et al. 2018)	Large groups	Slow (1.30-1.35)	Low-to-moderate impact; long incubation but slow population growth and close to population decline levels (Monello et al. 2014)

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Continent Species	<i>PRNP</i> allele variation	Pathology, incubation	Social organization	Maximum population growth rates (λ_{max})	Population impact
Moose <i>Alces alces</i>	2 <i>PRNP</i> alleles (Robinson et al. 2012b); 209M, 209I	Uncertain, likely normal	Solitary summer, small groups in open habitat	Intermediate (1.40-1.45)	Repeated spillover, no known epizootic (Baeten et al. 2007; Haley & Hoover 2015)
Caribou <i>Rangifer tarandus</i>	8 <i>PRNP</i> alleles (Robinson et al. 2012b), one Alberta population with alleles with some resistance (Cheng et al. 2017); 2V, M (Robinson et al. 2012b); 129GG, GS; 138SS, SN, NN (Cheng et al. 2017); 169V, M (Robinson et al. 2012b)	Normal (Moore et al. 2016)	Very large groups all year	Slow (1.30-1.35)	Uncertain, likely very high
Europe	In general, uncertain due to previously low interest				
Reindeer <i>Rangifer tarandus</i>	See above for North America	Normal (Viljugrein et al. 2018), terminal CWD 18.5-20 months post inoculation (Mitchell et al. 2012)	Very large groups all year	Slow (1.30-1.35)	Uncertain, likely very high; outbreak in Nordfjella, Norway
Red deer <i>Cervus elaphus</i>	8 <i>PRNP</i> alleles; 59G, S; 98T, A, 168P, S, 226E, Q (Robinson et al. 2012b)	As in elk (Schwablander et al. 2013)	Small groups in summer; larger in open habitat; larger in winter/spring	Slow (1.30-1.35)	Uncertain, likely moderate impact
Moose <i>Alces alces</i>	2 <i>PRNP</i> alleles	Uncertain, likely normal	Solitary summer, small groups occur in winter	Intermediate (1.40-1.45)	Uncertain, likely low (depending on population density & supplemental feeding?)

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Continent Species	<i>PRNP</i> allele variation	Pathology, incubation	Social organization	Maximum population growth rates (λ_{max})	Population impact
			(Bonenfant et al. 2004)		
Roe deer <i>Capreolus capreolus</i>	1 <i>PRNP</i> allele (Robinson et al. 2012b)	Uncertain	Solitary, small groups occur winter in agricultural landscapes	Very high (> 1.5)	Uncertain, likely low (depending on population density & supplemental feeding?)
<i>Introduced species</i>					
White-tailed deer	See above for North America				
Axis deer <i>Axis axis</i>	Uncertain	Uncertain	Solitary and small groups, larger groups in winter	Slow (1.30-1.35)	Uncertain
Fallow deer <i>Dama dama</i>	No <i>PRNP</i> allele variation (Robinson et al. 2012b)	Delayed; 4-5 yrs of incubation post inoculation (Hamir et al. 2011)	Large groups year-round	Slow (1.30-1.35)	Long incubation, lower impact despite being highly social; seem not to be infected via the natural route (Rhyan et al. 2011)
Sika deer <i>Cervus nippon</i>	4 <i>PRNP</i> alleles; 100S, G; 226E, Q (Robinson et al. 2012b)	Uncertain	Small groups	Slow (1.30-1.35)	Uncertain, likely intermediate
Muntjac <i>Muntiacus reevesi</i>	Uncertain	Normal (Nalls et al. 2013)	Solitary	Very high (>1.5)	Uncertain, likely low

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263 transmission of parasites in general (Milner et al. 2013). Social group size of red deer in
1
2 264 Europe differ depending on the habitat; group sizes as small as only 2-3 females occur during
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4 265 summer, but can be much larger during winter, especially when they are aggregating on
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7 266 feeding sites or agricultural pastures. Additionally, in an open habitat, such as in Scotland,
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10 267 they occur in large herds. Farmed red deer is known to contract CWD (Schwablander et al.
11
12 268 2013), but impact of CWD on red deer populations may differ from elk due different *PRNP*
13
14 269 genotypes (Table 2). Due to the gregarious nature of reindeer, we would expect higher contact
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16
17 270 rates among individuals of this species than for any other deer species and, hence, more rapid
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19 271 development towards endemic CWD. Hence, many aspects of cervid biology likely to affect
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22 272 transmission of CWD differ markedly within Europe even for the same species.
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25 273 **Effects of predation**

26
27 274 Any factor causing increased mortality of CWD-infected deer relative to non-infected deer
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29
30 275 may aid in limiting CWD, as it would decrease the period infected individuals can transmit
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32 276 and spread disease. Predators vary widely in the degree to which they target weak animals,
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34
35 277 and the effect of predators on infectious disease depends on epizootic detail. Predators can
36
37 278 keep herds healthy when the disease agent is highly virulent and aggregated in prey, prey are
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40 279 long-lived, and predators are selective for infected individuals (Packer et al. 2003). CWD
41
42 280 meets the conditions of having a strong impact on infected prey and with a clear distinction
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45 281 between infected and non-infected individuals that are long-lived, so the key question is the
46
47 282 level of selectivity. This is not a trivial issue due to the long incubation period. In the early
48
49 283 stages, the animals appear healthy but can spread disease (Tamguney et al. 2009) before they
50
51
52 284 slowly change behaviour and become more vulnerable to predation. Modelling wolf predation
53
54 285 on CWD-infected mule and/or white-tailed deer suggests that if predation is sufficiently
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56
57 286 selective for CWD-infected individuals, it could cause a marked decline in CWD prevalence
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59 287 (Wild et al. 2011). Empirical evidence for selective predation on CWD-infected individuals
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288 is, however, not consistent. CWD-infected mule deer were more likely to be depredated by
1
2 289 mountain lions (*Puma concolor*) than non-infected mule deer (Krumm et al. 2010). Predation
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5 290 can thus, to some extent, remove a higher proportion of CWD-infected individuals than is
6
7 291 present in the population. However, empirical evidence found that remarkably high CWD
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9
10 292 infection rates of mule deer were sustained even in the face of intense selective mountain lion
11
12 293 predation (Miller et al. 2008).

15 294 **Spillover among cervid species**

17
18 295 The evidence for population-level effects of CWD is derived from white-tailed deer, mule
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20 296 deer, and elk in North America. How fast CWD will grow, spread and spillover among the
21
22 297 cervid species in Europe remain uncertain. From a genetic perspective, there is a low barrier
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24
25 298 for transfer of CWD among most cervid species (Robinson et al. 2012b). However, fallow
26
27 299 deer (*Dama dama*) housed together with infected mule deer did not become infected via the
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30 300 natural route (Rhyan et al. 2011), even though fallow deer can contract CWD via intracerebral
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32 301 inoculation (Hamir et al. 2011). In North America, CWD is known to have been transferred
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34
35 302 from mule deer to white-tailed deer, mule deer to elk, and elk to mule deer and white-tailed
36
37 303 deer (Williams 2005); it is likely CWD was transferred from one or all of those three species
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39
40 304 to moose as well. A main uncertainty is whether CWD in reindeer in Norway will transmit
41
42 305 across species as has happened in North America. From experiments with mice, it is known
43
44 306 that the first transmission of a new prion strain to a new host may be difficult, but that
45
46
47 307 subsequent transmission (serial passage) becomes easier within the new species (Raymond et
48
49 308 al. 2007; Angers et al. 2010; Velásquez et al. 2015).

51
52
53 309 Hence, the process of spillover from one cervid species to another in the wild is not well
54
55 310 described or understood. It is likely that such transmission among species is indirect (i.e.,
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57 311 through environmental contamination), as direct contact between individuals of different
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59
60 312 species is rare. Even though the transmission of CWD within a species is not strongly density-

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313 dependent, it is likely that a spillover event would be linked to: 1) the population density of
314 the receiver species, 2) the spatial overlap of the two species, and 3) the density of infected
315 individuals in the donor population (Hansen et al. 2016). Contact points attracting multiple
316 species, such as common mineral licks (Plummer et al. 2018), supplemental feeding and
317 watering sites, or riparian habitats (Edmunds et al. 2018), are likely to be risk factors. Similar
318 feeding niches linked to feeding on low vegetation may be risk factors, as it is more likely to
319 transfer prions through ingestion of soil (Johnson et al. 2006; Johnson et al. 2007) and
320 vegetation (Pritzkow et al. 2015). It is suggested that the lower levels of prions in the lymph
321 nodes of elk compared with white-tailed deer and mule deer reduces the risk of elk
322 transmitting CWD to other species (Race et al. 2007). There is no direct empirical evidence to
323 support these hypotheses, so they are all inferred from general knowledge about CWD
324 transmission.

325 **Geographic spread of disease**

326 The spread of CWD in North America results from the movement of deer, which is often
327 linked to the dispersal of male yearlings (Lang & Blanchong 2012), but spread is also due to
328 the movement of infected deer by farming (Rorres et al. 2018). Male-biased dispersal is the
329 common pattern in cervids. Male yearlings typically have the longest dispersal distances for
330 all the affected North American species: white-tailed deer, mule deer and elk. In Europe, red
331 deer also have male-biased dispersal (Loe et al. 2009), but this is not the case for roe deer
332 (Wahlström & Liberg 1995; Gaillard et al. 2008) and likely not for moose. Juvenile dispersal
333 of roe deer is longer and a higher proportion takes place in low-quality than in high-quality
334 habitats (Wahlström & Liberg 1995), so expansion of CWD will likely be faster in low-
335 quality habitats with low population density and slower in areas with good habitats (Andersen
336 et al. 2004). There is also extensive long-distance migration of moose across the borders of
337 Norway and Sweden (Bunnefeld et al. 2011; Singh et al. 2012). In the case of deer movement,

338 major roads and rivers appear as semipermeable barriers (Blanchong et al. 2008; Long et al.
1
2 339 2010; Robinson et al. 2013). For both white-tailed deer (Cullingham et al. 2011a) and mule
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4 340 deer (Cullingham et al. 2011b) in western Canada, limited evidence of natural barriers for
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7 341 dispersal based on genetic structure were found, and even the Mississippi River in the USA
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10 342 had little impact on genetic differentiation (Lang & Blanchong 2012). In Scandinavia,
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12 343 highways are increasingly barriers to cervid movement and the barrier effect is often
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14 344 strengthened by wildlife fencing to avoid traffic accidents. Often these fences have wildlife
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17 345 passages, which could be closed to limit the spread of disease by deer movement. The spread
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19 346 of CWD at a broader scale is not easy to predict, as humans have played a major role in long-
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22 347 distance spread of CWD in North America, partly linked to transport of farmed deer (Rorres
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24 348 et al. 2018). Spread of CWD to Canada (Bollinger et al. 2004) and South Korea (Kim et al.
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26
27 349 2005) was through sales of farmed elk. If CWD becomes endemic in Scandinavia, human
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29 350 transport of infectious material to continental Europe will be a risk factor to consider. In a
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32 351 European setting, restrictions on the movement of farmed cervids are likely to hinder such
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34 352 spread; there is already a regulation on the export of live cervids from Norway.

353 **Surveillance for CWD in Europe**

354 Whether or not classical CWD is present in countries of Europe other than Norway remains to
355 be established. A survey during the period 2006–2010 across Europe detected no CWD in
356 either farmed or wild cervid populations (EFSA Panel on Biological Hazards (BIOHAZ) et al.
357 2016). However, the sample sizes were quite low. After the discovery of CWD in Norway,
358 the European Food Safety Authority proposed a 3-year surveillance program for Estonia,
359 Finland, Latvia, Lithuania, Norway, Poland and Sweden (EFSA Panel on Biological Hazards
360 (BIOHAZ) et al. 2016). This surveillance program for CWD will include both farmed and
361 wild cervids, and it will consist of random sampling at a population unit level and
362 convenience sampling targeting high-risk animals, typically fallen stock. The surveillance in

363 EU is aimed to detect CWD, and if present, intentions are to contain (avoid geographic
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2 364 spread) and to limit CWD transmission (actively stabilize or reduce infection rates) in an
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4 365 infected population (EFSA Panel on Biological Hazards (BIOHAZ) et al. 2016). This
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7 366 surveillance started in 2018, and the fallen stock sampling has already revealed the ‘non-
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10 367 classical’ type of CWD in a moose in Finland. The countries included in the surveillance
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12 368 program were based on the distribution of reindeer and moose, which at the time were the
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14 369 only species with CWD detected in Europe. Later (2017), ‘non-classical’ CWD was also
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17 370 discovered in red deer in Norway (Våge et al. 2018), and surveillance may become
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19 371 geographically extended to countries with red deer (EFSA Panel on Biological Hazard
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21 372 (BIOHAZ) et al. 2018). If the ‘non-classical’ CWD is a sporadic type of prion disease, which
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24 373 remains uncertain, it should be found at low prevalence in older animals with no clear
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26 374 geographic clustering of cases (Pirisinu et al. 2018). If correct, the discovery of ‘non-
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29 375 classical’ CWD will likely not require the same drastic management actions as ‘classical
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31 376 CWD’, which we describe in the following section.

35 377 **Hunting management strategies**

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37 378 Controlling CWD with hunting is difficult and has typically had limited success once
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40 379 established in the landscape (Uehlinger et al. 2016). We briefly go through the main
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42 380 principles of the different options (Table 3).

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45 381 *Depopulation or host eradication.* Complete eradication of an infected herd, following and
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48 382 subsequent restocking is the option typically used for farmed deer. Herd reduction to eradicate
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50 383 CWD was the aim of management when first detected in Wisconsin, but it was unsuccessful
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53 384 as significant herd reduction was not accomplished (Heberlein 2004). In Norway, the open
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55 385 habitat and use of professional marksmen made it possible to take out the entire reindeer
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58 386 population with detected CWD in the Nordfjella mountain range (Mysterud & Rolandsen
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60 387 2018), though the success in terms of CWD eradication is still uncertain. In forested areas, the

388 removal of all animals is difficult to achieve. This strategy is hence intended mainly for
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2 389 smaller populations, but it may be an option in some of Europe's fragmented landscapes. The
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4 390 recommended fallowing period is usually 5 years, but this limit was set without rigorous
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7 391 scientific testing. Due to the prion contamination of soil, it is uncertain whether this tactic
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10 392 works once CWD has become established, and early management action appears important.

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13 393 *Spatially targeted harvesting.* In the early stages of an epizootic, CWD is mainly transmitted
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15 394 by direct contact (Almberg et al. 2011). Therefore, non-selective harvest in a spatially
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18 395 confined region can take out infected individuals and limit the spread of CWD. The
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20 396 sharpshooter programme in Illinois is controversial, but it is the best evidence that such an
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23 397 effort may limit growth of CWD. They target deer non-selectively within blocks of 64 km²
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25 398 when an infected deer is discovered (Mateus-Pinilla et al. 2013; Manjerovic et al. 2014). For
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27 399 Europe, this appears to be a promising strategy for forest-living cervids. However, the
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30 400 distribution of CWD on the landscape is important, and the actual spatial scale of such
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32 401 targeted efforts should follow evidence about functional connectivity and migration of the
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35 402 given infected population. This is, however, not an alternative for species such as reindeer
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37 403 with no marked home range behaviour.

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40 404 *Male-targeted harvest.* CWD infection rates are strongly sex and age-specific (Jennelle et al.
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43 405 2014; Samuel & Storm 2016). Hunting (and predation) that targets specific sex and age
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45 406 groups may hence change the population prevalence of CWD due to changes in the
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47
48 407 demographic composition. Targeting males, who usually have higher CWD infection
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50 408 prevalence, are a management alternative (Jennelle et al. 2014; Uehlinger et al. 2016). The
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53 409 efficacy of such an action is not well established, and we regard it likely to slow, rather than
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55 410 stop, the growth rate of CWD.

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411 *Targeting clinical suspects.* Targeting clinical suspects when hunting may be either
412 intentional or non-intentional. The active targeting of clinical suspects appears to have limited
413 success due to prion shedding soon after infection (Hoover et al. 2017). CWD changes the
414 behaviour of animals (Edmunds et al. 2018), and this can make them more exposed to hunters
415 even with no active targeting. There was a selective harvest of CWD-positive white-tailed
416 deer in Wyoming (Edmunds et al. 2016) and for mule deer in Colorado (Conner et al. 2000).
417 For white-tailed deer in Wisconsin, the male offspring with CWD-infected mothers were
418 harvested more often than would be expected by chance (Blanchong et al. 2012). However, a
419 larger study of white-tailed deer in Wisconsin found no difference in the proportional harvest
420 of CWD-infected and non-infected deer over the hunting season (Grear et al. 2006; Heisey et
421 al. 2010). In heavily infected populations, hunters may also avoid shooting deer with unusual
422 behaviour to avoid getting infected meat (Conner et al. 2000). Relying on such measures is
423 not sufficient to limit CWD.

424 *Capture-test-and-cull.* An attempt was made in a mule deer population to capture, test and
425 mark individuals with GPS collars (Wolfe et al. 2018). Individuals later established to be
426 CWD-positive were removed from the population. These actions were only partly successful,
427 and they are highly invasive, economically costly and only likely to be an option in small
428 populations.

429 **Human dimension and consequences for wildlife management**

430 The above harvest management actions towards CWD are all rather drastic measures. In
431 addition, since aggregation of hosts is a risk factor for disease transmission, the governments
432 are likely to implement bans on both wildlife feeding and use of artificial mineral licks. This
433 is common practice in CWD-endemic areas in North America. Such bans have already been
434 implemented for the whole of Norway (Landbruks- og matdepartementet 2016), even though
435 CWD was only discovered in one location. During the severe winter of 2018 in Norway, this

436 cessation of supplemental feeding resulted in massive die-off of cervids locally; this was not
1
2 437 without controversy. Hunters in CWD-infected areas must take care of offal and several more
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4 438 minor restrictions will likely be implemented to avoid spatial spread, such as fencing
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7 439 (Mysterud & Rolandsen 2019). Therefore, the management actions of the government may
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9 440 have a far-reaching impact on wildlife management, even if CWD is discovered at a very low
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11 441 prevalence. These drastic management actions to combat CWD have been controversial and
12
13 442 politically contentious in North America (Heberlein 2004; Vaske 2010; Holsman et al. 2010;
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15 443 Vaske et al. 2018). Local resistance towards the depopulation strategy to fight CWD in the
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17 444 reindeer herd in Nordfjella was massive in Norway (Mysterud & Rolandsen 2018), and the
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19 445 public resistance towards a ban on winter feeding of wildlife appear common. In Wisconsin,
20
21 446 the number of hunting licenses sold declined initially even though the management tactic was
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23 447 for herd reduction to eradicate CWD; this decrease resulted from uncertainty among hunters
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25 448 regarding the zoonotic potential of eating CWD-infected meat (Heberlein 2004) and from
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27 449 reluctance to reduce deer density. Similarly, in Norway, the effectiveness of recreational
28
29 450 hunters was low compared to professional marksmen in the eradication process (Mysterud &
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31 451 Rolandsen 2018), but it is typically unpopular among hunters and landowners to use
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33 452 professionals. We can say with certainty that the emergence of CWD in any country will
34
35 453 cause considerable controversy and become a game changer for wildlife management. There
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37 454 is an overabundance of deer in many areas of North America and Europe. Some may argue
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39 455 CWD can be positive since it will contribute towards lowering deer densities. However, since
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41 456 CWD has a mainly frequency-dependent rather than density-dependent transmission (Table
42
43 457 1), CWD is unlikely to regulate deer numbers in a moderate way around a stable lower
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45 458 equilibrium. Also, the uncertainty regarding the zoonotic potential require testing of meat to
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47 459 avoid exposure. Both the surveillance for and combat of CWD are economically costly. The
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460 discovery of CWD may lead to a ban on the export of cervid meat and products from the
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 2 461 affected areas, which will affect livelihoods in many rural areas.
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 9 463 **Table 3** An overview of harvest management strategies aimed to eradicate or limit increases
 10
 11 464 in the prevalence of CWD.
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Management strategy	Biological basis	Rationale	Aim	Comment	Reference
Depopulation/host eradication	CWD has frequency-dependent transmission	Host eradication, fallowing and restocking	Eradication of CWD	Mainly for small and closed populations, or new outbreak with limited distribution	(Williams et al. 2002)
Spatially targeted non-selective harvest	CWD spread among related females with overlapping home ranges	Spatial clustering of positives allows lowering of overall prevalence	Limit growth in prevalence	Main option for CWD management in forested areas and open populations	(Manjerovic et al. 2014)
Male-targeted harvest	Higher infection prevalence in males, male-biased dispersal	Removing males will lower the overall prevalence and may limit spread	Limit growth in prevalence	For large populations where other options are not feasible, efficacy unknown	(Jennelle et al. 2014; Potapov et al. 2016)
Targeting clinical suspects	Late stage CWD associated with visible clinical signs of disease	Selective removal of positives lowers prevalence	Limit growth in prevalence	Most animals are asymptomatic until late stage, low efficacy	(Gross & Miller 2001)
Capture-test-and-cull	Mark animals, test for CWD, remove infected	Selective removal of positives lowers prevalence	Limit growth in prevalence	Costly and intrusive, for small populations, some effect	(Wolfe et al. 2018)

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466 **The future of cervids in Europe with CWD**

1
2 467 The future of many cervid populations in North America with CWD appears grim from a
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4
5 468 long-term perspective. The endpoint of the CWD epizootic has not been observed even in
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7 469 North America. Will endemic CWD progression lead to local extinction? We are potentially
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9
10 470 up against a disease that may have a devastating effect on cervid populations for as long as
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12 471 50-100 years or more. Indeed, a 50 year time period is regarded as the early stage of a CWD
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14 472 epizootic (Wasserberg et al. 2009; Almberg et al. 2011). The main uncertainty about the
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17 473 biological effect of CWD in Europe is linked to the following question: How transferable is
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19 474 the knowledge from different species in North America? Most populations are likely
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22 475 susceptible. It is entirely clear that the European Union (EU) will not allow free growth of
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24 476 such a serious disease without attempting management actions (EFSA Panel on Biological
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27 477 Hazards (BIOHAZ) et al. 2016), partly because of the experience with the mad cow disease
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29 478 (BSE) and the uncertain zoonotic potential of CWD (Waddell et al. 2018). Therefore, the
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32 479 impact on cervid populations through management countermeasures aiming to limit disease
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34 480 spread may have a large indirect impact on populations, even in early stages with low direct
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36 481 impact of CWD. However, even such drastic management countermeasures are not very
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39 482 effective, at least partly due to high levels of environmental contamination, if CWD becomes
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41 483 endemic (Uehlinger et al. 2016). The coming years will therefore be critical to avoid taking
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44 484 such risks. Early action require early detection and rigorous surveillance is key. We currently
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46 485 can only hope that early management actions will be successful in the quick eradication of
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49 486 CWD from Europe (Hansen et al. 2016; Stokstad 2017); the first stage of eradication of the
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51 487 whole reindeer herd infected with CWD in Norway were successful (Myserud & Rolandsen
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53 488 2018). Due to the keystone role of cervids across ecosystems in Europe and their high
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56 489 associated economic and cultural importance (Apollonio et al. 2010), the consequences of
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58 490 failure may be dramatic.

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References

Almberg, E.S., Cross, P.C., Johnson, C.J., Heisey, D.M. & Richards, B.J. 2011: Modeling routes of Chronic Wasting Disease transmission: environmental prion persistence promotes deer population decline and extinction. *Plos One* 6: e19896

Andersen, R., Herfindal, I., Sæther, B.-E., Linnell, J.D.C., Odden, J. & Liberg, O. 2004: When range expansion rate is faster in marginal habitats. *Oikos* 107: 210-214.

Angers, R.C., Kang, H.E., Napier, D., Browning, S., Seward, T., Mathiason, C., Balachandran, A., McKenzie, D., Castilla, J., Soto, C., et al. 2010: Prion strain mutation determined by prion protein conformational compatibility and primary structure. *Science* 328: 1154

Apollonio, M., Andersen, R. & Putman, R. 2010: European ungulates and their management in the 21st century. Cambridge University Press; Cambridge.

Baeten, L.A., Powers, B.E., Jewell, J.E., Spraker, T.R. & Miller, M.W. 2007: A natural case of chronic wasting disease in a free-ranging moose (*Alces alces shirasi*). *Journal of Wildlife Diseases* 43: 309-314.

- 514 Benestad, S.L., Mitchell, G., Simmons, M., Ytrehus, B. & Vikøren, T. 2016: First case of
1
2 515 chronic wasting disease in Europe in a Norwegian free-ranging reindeer. *Veterinary*
3
4
5 516 *Research* 47: 88
6
- 7 517 Benestad, S.L., Sarradin, P., Thu, B., Schonheit, J., Tranulis, M.A. & Bratberg, B. 2003:
8
9
10 518 Cases of scrapie with unusual features in Norway and designation of a new type,
11
12 519 Nor98. *Veterinary Record* 153: 2002-2008.
13
- 14 520 Blanchong, J.A., Grear, D.A., Weckworth, B.V., Keane, D.P., Scribner, K.T. & Samuel,
15
16
17 521 M.D. 2012: Effects of Chronic Wasting Disease on reproduction and fawn harvest
18
19 522 vulnerability in Wisconsin white-tailed deer. *Journal of Wildlife Diseases* 48: 361-
20
21
22 523 370.
23
- 24 524 Blanchong, J.A., Samuel, M.D., Scribner, K.T., Weckworth, B.V., Langenberg, J.A. &
25
26
27 525 Filcek, K.B. 2008: Landscape genetics and the spatial distribution of chronic wasting
28
29 526 disease. *Biology Letters* 4: 130-133.
30
- 31 527 Bollinger, T., Caley, P., Merrill, E., Messier, F., Miller, M.W., Samuel, M.D. &
32
33
34 528 Vanopdenbosch, E. 2004: Chronic Wasting Disease in Canadian wildlife: an expert
35
36
37 529 opinion on the epidemiology and risks to wild deer. *Canadian Cooperative Wildlife*
38
39 530 *Health Centre*; Saskatoon, Canada.
40
- 41 531 Bonenfant, C., Loe, L.E., Mysterud, A., Langvatn, R., Stenseth, N.C., Gaillard, J.-M. &
42
43
44 532 Klein, F. 2004: Multiple causes of sexual segregation in European red deer:
45
46 533 enlightenments from varying breeding phenology at high and low latitude.
47
48
49 534 *Proceedings of the Royal Society of London, Series B* 271: 883-892.
50
- 51 535 Bunnefeld, N., Börger, L., Van Moorter, B., Rolandsen, C.M., Dettki, H., Solberg, E.J. &
52
53
54 536 Ericsson, G. 2011: A model-driven approach to quantify migration patterns:
55
56 537 individual, regional and yearly differences. *Journal of Animal Ecology* 80: 466-476.
57
58
59
60
61
62
63
64
65

- 538 Cheng, Y.C., Musiani, M., Cavedon, M. & Gilch, S. 2017: High prevalence of prion protein
1
2 539 genotype associated with resistance to Chronic Wasting Disease in one Alberta
3
4
5 540 woodland caribou population. *Prion* 11: 136-142.
6
- 7 541 Conner, M.M., McCarthy, C.W. & Miller, M.W. 2000: Detection of bias in harvest-based
8
9
10 542 estimates of Chronic Wasting Disease prevalence in mule deer. *Journal of Wildlife*
11
12 543 *Diseases* 36: 691-699.
13
- 14 544 Conner, M.M., Miller, M.W., Ebinger, M.R. & Burnham, K.P. 2007: A meta-BACI approach
15
16
17 545 for evaluating management intervention on Chronic Wasting Disease in mule deer.
18
19 546 *Ecological Applications* 17: 140-153.
20
- 21
22 547 Cross, P.C., Creech, T.G., Ebinger, M.R., Manlove, K., Irvine, K., Henningsen, J.,
23
24 548 Rogerson, J., Scurlock, B.M. & Creel, S. 2013: Female elk contacts are neither
25
26
27 549 frequency nor density dependent. *Ecology* 94: 2076-2086.
28
- 29 550 Cullingham, C.I., Merrill, E.H., Pybus, M.J., Bollinger, T.K., Wilson, G.A. & Coltman, D.W.
30
31 551 2011a: Broad and fine-scale genetic analysis of white-tailed deer populations:
32
33
34 552 estimating the relative risk of chronic wasting disease spread. *Evolutionary*
35
36 553 *Applications* 4: 116-131.
37
- 38
39 554 Cullingham, C.I., Nakada, S.M., Merrill, E.H., Bollinger, T.K., Pybus, M.J. & Coltman,
40
41 555 D.W. 2011b: Multiscale population genetic analysis of mule deer (*Odocoileus*
42
43 556 *hemionus hemionus*) in western Canada sheds new light on the spread of chronic
44
45
46 557 wasting disease. *Canadian Journal of Zoology* 89: 134-147.
47
- 48
49 558 DeVivo, M.T. 2015: Chronic disease ecology and epidemiology of mule deer in Wyoming.
50
51 559 PhD-thesis, University of Wyoming; Laramie, WY, USA.
52
- 53 560 DeVivo, M.T., Edmunds, D.R., Kauffman, M.J., Schumaker, B.A., Binfet, J., Kreeger, T.J.,
54
55
56 561 Richards, B.J., Schätzl, H.M. & Cornish, T.E. 2017: Endemic chronic wasting disease
57
58 562 causes mule deer population decline in Wyoming. *Plos One* 12: e0186512
59
60
61
62
63
64
65

563 Dulberger, J., Hobbs, N.T., Swanson, H.M., Bishop, C.J. & Miller, M.W. 2010a: Estimating
1
2 564 Chronic Wasting Disease effects on mule deer recruitment and population growth.
3
4 565 Journal of Wildlife Diseases 46: 1086-1095.
5
6
7 566 Dulberger, J., Hobbs, N.T., Swanson, H.M., Bishop, C.J. & Miller, M.W. 2010b: Estimating
8
9 567 Chronic Wasting Disease effects on mule deer recruitment and population growth.
10
11
12 568 Journal of Wildlife Diseases 46: 1086-1095.
13
14 569 Edmunds, D.R., Albeke, S.E., Grogan, R.G., Lindzey, F.G., Legg, D.E., Cook, W.E.,
15
16 570 Schumaker, B.A., Kreeger, T.J. & Cornish, T.E. 2018: Chronic wasting disease
17
18 571 influences activity and behavior in white-tailed deer. Journal of Wildlife Management
19
20
21 572 82: 138-154.
22
23
24 573 Edmunds, D.R., Kauffman, M.J., Schumaker, B.A., Lindzey, F.G., Cook, W.E., Kreeger,
25
26 574 T.J., Grogan, R.G. & Cornish, T.E. 2016: Chronic Wasting Disease drives population
27
28 575 decline of white-tailed deer. Plos One 11: e0161127
29
30
31 576 EFSA Panel on Biological Hazard (BIOHAZ), Ricci, A., Allende, A., Bolton, D., Chemaly,
32
33 577 M., Davies, R., Escámez, P.S.F., Gironés, R., Herman, L., Koutsoumanis, K., et al.
34
35 578 2018: Scientific opinion on chronic wasting disease (II). EFSA Journal 16: e05132
36
37
38 579 EFSA Panel on Biological Hazards (BIOHAZ), Ricci, A., Allende, A., Bolton, D., Chemaly,
39
40 580 M., Davies, R., Escámez, P.S.F., Gironés, R., Herman, L., Koutsoumanis, K., et al.
41
42 581 2016: Chronic Wasting Disease (CWD) in cervids. EFSA Journal 15: 4667
43
44
45 582 Evans, T.S., Schuler, K.L. & Walter, W.D. 2014: Surveillance and monitoring of white-tailed
46
47 583 deer for Chronic Wasting Disease in the Northeastern United States. Journal of Fish
48
49 584 and Wildlife Management 5: 387-393.
50
51
52 585 Foley, A.M., Hewitt, D.G., DeYoung, C.A., DeYoung, R.W. & Schnupp, M.J. 2016:
53
54 586 Modeled impacts of Chronic Wasting Disease on white-tailed deer in a semi-arid
55
56 587 environment. PLoS ONE 11: e0163592
57
58
59
60
61
62
63
64
65

- 588 Fox, K.A., Jewell, J.E., Williams, E.S. & Miller, M.W. 2006: Patterns of PRPCWD
1
2 589 accumulation during the course of Chronic Wasting Disease infection in orally
3
4 590 inoculated mule deer (*Odocoileus hemionus*). Journal of General Virology 87: 3451-
5
6
7 591 3461.
8
9 592 Gaillard, J.-M., Festa-Bianchet, M. & Yoccoz, N.G. 1998: Population dynamics of large
10
11 593 herbivores: variable recruitment with constant adult survival. Trends in Ecology and
12
13 594 Evolution 13: 58-63.
14
15 595 Gaillard, J.-M., Festa-Bianchet, M., Yoccoz, N.G., Loison, A. & Toigo, C. 2000: Temporal
16
17 596 variation in fitness components and population dynamics of large herbivores. Annual
18
19 597 Review of Ecology and Systematics 31: 367-393.
20
21
22 598 Gaillard, J.-M., Hewison, A.J.M., Kjellander, P., Pettorelli, N., Bonenfant, C., Van Moorter,
23
24 599 B., Liberg, O., Andrén, H., Van Laere, G., Klein, F., et al. 2008: Population density
25
26 600 and sex do not influence fine-scale natal dispersal in roe deer. Proceedings of the
27
28 601 Royal Society of London, Series B 275: 2025-2030.
29
30
31 602 Geremia, C., Miller, M.W., Hoeting, J.A., Antolin, M.F. & Hobbs, N.T. 2015: Bayesian
32
33 603 modeling of prion disease dynamics in mule deer using population monitoring and
34
35 604 capture-recapture data. Plos One 10: e0140687
36
37
38 605 Grear, D.A., Samuel, M.D., Langenberg, J.A. & Keane, D. 2006: Demographic patterns and
39
40 606 harvest vulnerability of chronic wasting disease infected white-tailed deer in
41
42 607 Wisconsin. Journal of Wildlife Management 70: 546-553.
43
44
45 608 Grear, D.A., Samuel, M.D., Scribner, K.T., Weckworth, B.V. & Langenberg, J.A. 2010:
46
47 609 Influence of genetic relatedness and spatial proximity on chronic wasting disease
48
49 610 infection among female white-tailed deer. Journal of Applied Ecology 47: 532-540.
50
51
52 611 Gross, J.E. & Miller, M.W. 2001: Chronic Wasting Disease in mule deer: disease dynamics
53
54 612 and control. Journal of Wildlife Management 65: 205-215.
55
56
57
58
59
60
61
62
63
64
65

- 613 Habib, T.J., Merrill, E.H., Pybus, M.J. & Coltman, D.W. 2011: Modelling landscape effects
1
2 614 on density-contact rate relationships of deer in eastern Alberta: Implications for
3
4 615 chronic wasting disease. *Ecological Modelling* 222: 2722-2732.
5
6
7 616 Haley, N.J. & Hoover, E.A. 2015: Chronic Wasting Disease of cervids: current knowledge
8
9 617 and future perspectives. *Annual Review of Animal Biosciences* 3: 305-325.
10
11 618 Haley, N.J. & Richt, A.J. 2017: Evolution of diagnostic tests for Chronic Wasting Disease, a
12
13 619 naturally occurring prion disease of cervids. *Pathogens* 6: E35
14
15
16 620 Hamir, A.N., Greenlee, J.J., Nicholson, E.M., Kunkle, R.A., Richt, J.A., Miller, J.M. & Hall,
17
18 621 M. 2011: Experimental transmission of chronic wasting disease (CWD) from elk and
19
20 622 white-tailed deer to fallow deer by intracerebral route: Final report. *Canadian Journal*
21
22 623 of *Veterinary Research* 75: 152-156.
23
24
25
26 624 Hansen, H., Kapperud, G., Mysterud, A., Solberg, E.J., Strand, O., Tranulis, M., Ytrehus, B.,
27
28 625 Asmyhr, M.G. & Grahek-Ogden, D. 2016: CWD in Norway - a state of emergency for
29
30 626 the future of cervids (phase II). Opinion of the panel on biological hazards of the
31
32 627 Norwegian scientific committee for food safety.; Oslo.
33
34
35
36 628 Heberlein, T.A. 2004: "Fire in the Sistine Chapel": How Wisconsin responded to Chronic
37
38 629 Wasting Disease. *Human Dimensions of Wildlife* 9: 165-179.
39
40
41 630 Heisey, D.M., Osnas, E.E., Cross, P.C., Joly, D.O., Langenberg, J.A. & Miller, M.W. 2010:
42
43 631 Linking process to pattern: estimating spatiotemporal dynamics of a wildlife epidemic
44
45 632 from cross-sectional data. *Ecological Monographs* 80: 221-240.
46
47
48 633 Holsman, R.H., Petchenik, J. & Cooney, E.E. 2010: CWD after "the fire": Six reasons why
49
50 634 hunters resisted Wisconsin's eradication effort. *Human Dimensions of Wildlife* 15:
51
52 635 180-193.
53
54
55
56
57
58
59
60
61
62
63
64
65

- 636 Hoover, C.E., Davenport, K.A., Henderson, D.M., Denkers, N.D., Mathiason, C.K., Soto, C.,
1
2 637 Zabel, M.D. & Hoover, E.A. 2017: Pathways of prion spread during early Chronic
3
4 638 Wasting Disease in deer. *Journal of Virology* 91: e00077-17.
5
6
7 639 Jennelle, C.S., Henaux, V., Wasserberg, G., Thiagarajan, B., Rolley, R.E. & Samuel, M.D.
8
9
10 640 2014: Transmission of Chronic Wasting Disease in Wisconsin white-Tailed deer:
11
12 641 Implications for disease spread and management. *Plos One* 9: e91043
13
14 642 Jewell, J.E., Conner, M.M., Wolfe, L.L., Miller, M.W. & Williams, E.S. 2005: Low
15
16 643 frequency of PrP genotype 225SF among free-ranging mule deer (*Odocoileus*
17
18 644 *hemionus*) with chronic wasting disease. *Journal of General Virology* 86: 2127-2134.
19
20
21 645 Johnson, C.J., Herbst, A., Duque-Velasquez, C., Vanderloo, J.P., Bochsler, P., Chappell, R.
22
23
24 646 & McKenzie, D. 2011: Prion protein polymorphisms affect Chronic Wasting Disease
25
26 647 progression. *Plos One* 6: e17450
27
28
29 648 Johnson, C.J., Pedersen, J.A., Chappell, R.J., McKenzie, D. & Aiken, J.M. 2007: Oral
30
31 649 transmissibility of prion disease is enhanced by binding to soil particles. *Plos*
32
33 650 *Pathogens* 3: e93
34
35
36 651 Johnson, C.J., Phillips, K.E., Schramm, P.T., McKenzie, D., Aiken, J.M. & Pedersen, J.A.
37
38 652 2006: Prions adhere to soil minerals and remain infectious. *Plos Pathogens* 2: e32
39
40
41 653 Kim, T.Y., Shon, H.J., Joo, Y.S., Mun, U.K., Kang, K.S. & Lee, Y.S. 2005: Additional cases
42
43 654 of Chronic Wasting Disease in imported deer in Korea. *Journal of Veterinary Medical*
44
45 655 *Science* 67: 753-759.
46
47
48 656 Kjær, L.J., Schaubert, E.M. & Nielsen, C.K. 2008: Spatial and temporal analysis of contact
49
50 657 rates in female white-tailed deer. *Journal of Wildlife Management* 72: 1819-1825.
51
52
53 658 Krumm, C.E., Conner, M.M., Hobbs, N.T., Hunter, D.O. & Miller, M.W. 2010: Mountain
54
55 659 lions prey selectively on prion-infected mule deer. *Biology Letters* 6: 209-211.
56
57
58
59
60
61
62
63
64
65

- 660 Krumm, C.E., Conner, M.M. & Miller, M.W. 2005: Relative vulnerability of chronic wasting
1
2 661 disease infected mule deer to vehicle collisions. *Journal of Wildlife Diseases* 41: 503-
3
4 662 511.
5
6
7 663 Landbruks- og matdepartementet. 2016: Regulation 11 July 2016 No 913 concerning
8
9 664 measures to reduce the spread of Chronic Wasting Disease (CWD).
10
11 665 <https://lovdata.no/dokument/SF/forskrift/2016-07-11-913?q=cwd>;
12
13
14 666 Lang, K.R. & Blanchong, J.A. 2012: Population genetic structure of white-tailed deer:
15
16 667 Understanding risk of chronic wasting disease spread. *Journal of Wildlife*
17
18 668 *Management* 76: 832-840.
19
20
21 669 Loe, L.E., Mysterud, A., Veiberg, V. & Langvatn, R. 2009: Negative density-dependent
22
23 670 emigration of males in an increasing red deer population. *Proceedings of the Royal*
24
25 671 *Society of London, Series B* 276: 2581-2587.
26
27
28 672 Long, E.S., Diefenbach, D.R., Wallingford, B.D. & Rosenberry, C.S. 2010: Influence of
29
30 673 roads, rivers, and mountains on natal dispersal of white-tailed deer. *Journal of Wildlife*
31
32 674 *Management* 74: 1242-1249.
33
34
35 675 Manjerovic, M.B., Green, M.L., Mateus-Pinilla, N. & Novakofski, J. 2014: The importance
36
37 676 of localized culling in stabilizing chronic wasting disease prevalence in white-tailed
38
39 677 deer populations. *Preventive Veterinary Medicine* 113: 139-145.
40
41
42 678 Mateus-Pinilla, N., Weng, H.Y., Ruiz, M.O., Shelton, P. & Novakofski, J. 2013: Evaluation
43
44 679 of a wild white-tailed deer population management program for controlling chronic
45
46 680 wasting disease in Illinois, 2003-2008. *Preventive Veterinary Medicine* 110: 541-548.
47
48
49
50 681 Miller, M.W. & Conner, M.M. 2005: Epidemiology of Chronic Wasting Disease in free-
51
52 682 ranging mule deer: Spatial, temporal, and demographic influences on observed
53
54 683 prevalence patterns. *Journal of Wildlife Diseases* 41: 275-290.
55
56
57
58
59
60
61
62
63
64
65

- 684 Miller, M.W., Hobbs, N.T. & Tavener, S.J. 2006: Dynamics of prion disease transmission in
1
2 685 mule deer. *Ecological Applications* 16: 2208-2214.
3
- 4 686 Miller, M.W., Swanson, H.M., Wolfe, L.L., Quartarone, F.G., Huwer, S.L., Southwick, C.H.
5
6
7 687 & Lukacs, P.M. 2008: Lions and prions and deer demise. *Plos One* 3: e4019
8
- 9 688 Miller, M.W. & Williams, E.S. 2003: Horizontal prion transmission in mule deer. *Nature*
10
11 689 425: 35-36.
12
13
- 14 690 Miller, M.W., Williams, E.S., Hobbs, N.T. & Wolfe, L.L. 2004: Environmental sources of
15
16 691 prion transmission in mule deer. *Emerging infectious diseases* 10: 1003-1006.
17
18
- 19 692 Miller, M.W., Williams, E.S., McCarty, C.W., Spraker, T.R., Kreeger, T.J., Larsen, C.T. &
20
21 693 Thorne, E.T. 2000: Epizootiology of Chronic Wasting Disease in free-ranging cervids
22
23 694 in Colorado and Wyoming. *Journal of Wildlife Diseases* 36: 676-690.
24
25
- 26 695 Milner, J.M., Wedul, S.J., Laaksonen, S. & Oksanen, A. 2013: Gastrointestinal nematodes of
27
28 696 moose (*Alces alces*) in relation to supplementary feeding. *Journal of Wildlife Diseases*
29
30 697 49: 69-79.
31
32
33
- 34 698 Mitchell, G.B., Sigurdson, C.J., O'Rourke, K.I., Algire, J., Harrington, N.P., Walther, I.,
35
36 699 Spraker, T.R. & Balachandran, A. 2012: Experimental oral transmission of Chronic
37
38 700 Wasting Disease to reindeer (*Rangifer tarandus tarandus*). *Plos One* 7: e39055
39
40
- 41 701 Monello, R.J., Galloway, N.L., Powers, J.G., Madsen-Bouterse, S.A., Edwards, W.H., Wood,
42
43 702 M.E., O'Rourke, K.I. & Wild, M.A. 2017: Pathogen-mediated selection in free-
44
45 703 ranging elk populations infected by chronic wasting disease. *Proceedings of the*
46
47 704 *National Academy of Sciences, USA* 114: 12208-12212.
48
49
50
- 51 705 Monello, R.J., Powers, J.G., Hobbs, N.T., Spraker, T.R., Watry, M.K. & Wild, M.A. 2014:
52
53 706 Survival and population growth of a free-ranging elk population with a long history of
54
55 707 exposure to Chronic Wasting Disease. *Journal of Wildlife Management* 78: 214-223.
56
57
58
59
60
61
62
63
64
65

708 Moore, S.J., Kunkle, R., Greenlee, M.H.W., Nicholson, E., Richt, J., Hamir, A., Waters,
1
2 709 W.R. & Greenlee, J. 2016: Horizontal transmission of Chronic Wasting Disease in
3
4 710 reindeer. *Emerging infectious diseases* 22: 2142
5
6
7 711 Moore, S.J., Vrentas, C.E., Hwang, S., West Greenlee, M.H., Nicholson, E.M. & Greenlee,
8
9 712 J.J. 2018: Pathologic and biochemical characterization of PrPSc from elk with PRNP
10
11 713 polymorphisms at codon 132 after experimental infection with the chronic wasting
12
13 714 disease agent. *BMC Veterinary Research* 14: 80
14
15
16
17 715 Mysterud, A. & Rolandsen, C.M. 2018: A reindeer cull to prevent chronic wasting disease in
18
19 716 Europe. *Nature Ecology and Evolution* 2: 1343-1345.
20
21
22 717 Mysterud, A. & Rolandsen, C.M. 2019: Fencing for wildlife disease control. *Journal of*
23
24 718 *Applied Ecology* 56: in press
25
26
27 719 Nalls, A.V., McNulty, E., Powers, J., Seelig, D.M., Hoover, C., Haley, N.J., Hayes-Klug, J.,
28
29 720 Anderson, K., Stewart, P., Goldmann, W., et al. 2013: Mother to offspring
30
31 721 transmission of Chronic Wasting Disease in Reeves' muntjac deer. *Plos One* 8: e71844
32
33
34 722 O'Rourke, K.I., Besser, T.E., Miller, M., Cline, T., Spraker, T., Jenny, A., Wild, M., Zebarth,
35
36 723 G. & Williams, E. 1999: PrP genotypes of captive and free-ranging Rocky Mountain
37
38 724 elk (*Cervus elaphus nelsoni*) with chronic wasting disease. *Journal of General*
39
40 725 *Virology* 80: 2765-2679.
41
42
43
44 726 O'Rourke, K.I., Spraker, T.R., Hamburg, L.K., Besser, T.E., Brayton, K.A. & Knowles, D.P.
45
46 727 2004: Polymorphisms in the prion precursor functional gene but not the pseudogene
47
48 728 are associated with susceptibility to chronic wasting disease in white-tailed deer.
49
50
51 729 *Journal of General Virology* 85: 1339-1346.
52
53
54 730 Packer, C., Holt, R.D., Hudson, P.J., Lafferty, K.D. & Dobson, A.P. 2003: Keeping the herds
55
56 731 healthy and alert: implications of predator control for infectious disease. *Ecology*
57
58 732 *Letters* 6: 797-802.
59
60
61
62
63
64
65

733 Pays, O., Benhamou, S., Helder, R. & Gerard, J.-F. 2007: The dynamics of group formation
1
2 734 in large mammalian herbivores: an analysis in the European roe deer. *Animal*
3
4 735 *Behaviour* 74: 1429-1441.
5
6
7 736 Perrott, M.R., Sigurdson, C.J., Mason, G.L. & Hoover, E.A. 2012: Evidence for distinct
8
9 737 chronic wasting disease (CWD) strains in experimental CWD in ferrets. *Journal of*
10
11 738 *General Virology* 93: 212-221.
12
13
14 739 Pirisinu, L., Tran, L., Chiappini, B., Vanni, I., Di Bari, M.A., Vaccari, G., Vikøren, T.,
15
16 740 Madslie, K., Våge, J., Spraker, T., et al. 2018: A novel type of Chronic Wasting
17
18 741 Disease detected in European moose (*Alces alces*) in Norway. *Emerging infectious*
19
20 742 *diseases* 24: 2210-2218.
21
22
23
24 743 Plummer, I.H., Johnson, C.J., Chesney, A.R., Pedersen, J.A. & Samuel, M.D. 2018: Mineral
25
26 744 licks as environmental reservoirs of chronic wasting disease prions. *Plos One* 13:
27
28 745 e0196745
29
30
31 746 Potapov, A., Merrill, E., Pybus, M. & Lewis, M.A. 2015: Empirical estimation of R0 for
32
33 747 unknown transmission functions: The case of Chronic Wasting Disease in Alberta.
34
35 748 *Plos One* 10: e0140024
36
37
38 749 Potapov, A., Merrill, E., Pybus, M. & Lewis, M.A. 2016: Chronic wasting disease:
39
40 750 Transmission mechanisms and the possibility of harvest management. *Plos One* 11:
41
42 751 e0151039
43
44
45 752 Pritzkow, S., Morales, R., Moda, F., Khan, U., Telling, G.C., Hoover, E. & Soto, C. 2015:
46
47 753 Grass plants bind, retain, uptake and transport infectious prions. *Cell reports* 11: 1168-
48
49 754 1175.
50
51
52
53 755 Prusiner, S.B. 1998: Prions. *Proceedings of the National Academy of Sciences, USA* 95:
54
55 756 13363-13383.
56
57
58
59
60
61
62
63
64
65

757 Race, B.L., Meade-White, K.D., Ward, A., Jewell, J., Miller, A.W., Williams, E.S.,
1
2 758 Chesebro, B. & Race, R.E. 2007: Levels of abnormal prion protein in deer and elk
3
4 759 with Chronic Wasting Disease. *Emerging infectious diseases* 13: 824-830.
5
6
7 760 Raymond, G.J., Raymond, L.D., Meade-White, K.D., Hughson, A.G., Favara, C., Gardner,
8
9 761 D., Williams, E.S., Miller, M.W., Race, R.E. & Caughey, B. 2007: Transmission and
10
11 762 adaptation of chronic wasting disease to hamsters and transgenic mice: evidence for
12
13 763 strains. *Journal of Virology* 81: 4305-4314.
14
15
16
17 764 Rhyan, J.C., Miller, M.W., Spraker, T.R., McCollum, M., Nol, P., Wolfe, L.L., Davis, T.R.,
18
19 765 Creekmore, L. & O'Rourke, K.I. 2011: Failure of fallow deer (*Dama dama*) to develop
20
21 766 Chronic Wasting Disease when exposed to a contaminated environment and infected
22
23 767 mule deer (*Odocoileus hemionus*). *Journal of Wildlife Diseases* 47: 739-744.
24
25
26
27 768 Robinson, S.J., Samuel, M.D., Johnson, C.J., Adams, M. & McKenzie, D.I. 2012a: Emerging
28
29 769 prion disease drives host selection in a wildlife population. *Ecological Applications*
30
31 770 22: 1050-1059.
32
33
34 771 Robinson, S.J., Samuel, M.D., O'Rourke, K.I. & Johnson, C.J. 2012b: The role of genetics in
35
36 772 chronic wasting disease of North American cervids. *Prion* 6: 153-162.
37
38
39 773 Robinson, S.J., Samuel, M.D., Rolley, R.E. & Shelton, P. 2013: Using landscape
40
41 774 epidemiological models to understand the distribution of chronic wasting disease in
42
43 775 the Midwestern USA. *Landscape Ecology* 28: 1923-1935.
44
45
46 776 Rorres, C., Romano, M., Miller, J.A., Mossey, J.M., Grubestic, A.H., Zellner, D.E. & Smith,
47
48 777 G. 2018: Contact tracing for the control of infectious disease epidemics: Chronic
49
50 778 Wasting Disease in deer farms. *Epidemics* 23: 71-75.
51
52
53 779 Samuel, M.D. & Storm, D.J. 2016: Chronic wasting disease in white-tailed deer: infection,
54
55 780 mortality, and implications for heterogeneous transmission. *Ecology* 97: 3195-3205.
56
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58
59
60
61
62
63
64
65
- 781 Schauber, E.M., Nielsen, C.K., Kjær, L.J., Anderson, C.W. & Storm, D.J. 2015: Social
782 affiliation and contact patterns among white-tailed deer in disparate landscapes:
783 implications for disease transmission. *Journal of Mammalogy* 96: 16-28.
- 784 Schwabenlander, M.D., Culhane, M.R., Hall, S.M., Goyal, S.M., Anderson, P.L., Carstensen,
785 M., Wells, S.J., Slade, W.B. & Armién, A.G. 2013: A case of chronic wasting disease
786 in a captive red deer (*Cervus elaphus*). *Journal of Veterinary Diagnostic Investigation*
787 25: 573-576.
- 788 Sharp, A. & Pastor, J. 2011: Stable limit cycles and the paradox of enrichment in a model of
789 chronic wasting disease. *Ecological Applications* 21: 1024-1030.
- 790 Singh, N.J., Börger, L., Dettki, H., Bunnefeld, N. & Ericsson, G. 2012: From migration to
791 nomadism: movement variability in a northern ungulate across its latitudinal range.
792 *Ecological Applications* 22: 2007-2020.
- 793 Spraker, T.R., Miller, M.W., Williams, E.S., Getzy, D.M., Adrian, W.J., Schoonveld, G.G.,
794 Spowart, R.A., O'Rourke, K.I., Miller, J.M. & Merz, P.A. 1997: Spongiform
795 encephalopathy in free-ranging mule deer (*Odocoileus hemionus*), white-tailed deer
796 (*Odocoileus virginianus*) and Rocky Mountain elk (*Cervus elaphus nelsoni*) in
797 northcentral Colorado. *Journal of Wildlife Diseases* 33: 1-6.
- 798 Stokstad, E. 2017: Norway seeks to stamp out prion disease. *Science* 356: 12
- 799 Storm, D.J., Samuel, M.D., Rolley, R.E., Shelton, P., Keuler, N.S., Richards, B.J. & Van
800 Deelen, T.R. 2013: Deer density and disease prevalence influence transmission of
801 chronic wasting disease in white-tailed deer. *Ecosphere* 4: 1-14.
- 802 Tamguney, G., Miller, M.W., Wolfe, L.L., Sirochman, T.M., Glidden, D.V., Palmer, C.,
803 Lemus, A., DeArmond, S.J. & Prusiner, S.B. 2009: Asymptomatic deer excrete
804 infectious prions in faeces. *Nature* 461: 529-532.

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47
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49
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51
52
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54
55
56
57
58
59
60
61
62
63
64
65
- 805 Uehlinger, F.D., Johnston, A.C., Bollinger, T.K. & Waldner, C.L. 2016: Systematic review
806 of management strategies to control chronic wasting disease in wild deer populations
807 in North America. *BMC Veterinary Research* 12: 1-16.
- 808 Våge, J., Hopp, P., Vikøren, T., Madslie, K., Tarpai, A., Moldal, T. & Benestad, S.L. 2018:
809 The surveillance programme for Chronic Wasting Disease (CWD) in free-ranging and
810 captive cervids in Norway 2017. Norwegian Veterinary Institute; Oslo.
- 811 Vaske, J.J. 2010: Lessons learned from human dimensions of Chronic Wasting Disease
812 research. *Human Dimensions of Wildlife* 15: 165-179.
- 813 Vaske, J.J., Miller, C.A., Ashbrook, A.L. & Needham, M.D. 2018: Proximity to chronic
814 wasting disease, perceived risk, and social trust in the managing agency. *Human
815 Dimensions of Wildlife* 23: 115-128.
- 816 Velásquez, C.D., Kim, C., Herbst, A., Daude, N., Garza, M.C., Wille, H., Aiken, J. &
817 McKenzie, D. 2015: Deer prion proteins modulate the emergence and adaptation of
818 Chronic Wasting Disease strains. *Journal of Virology* 89: 12362-12373.
- 819 Viljugrein, H., Hopp, P., Benestad, S.L., Nilsen, E.B., Våge, J., Tavorpanich, S., Rolandsen,
820 C.M., Strand, O. & Myrsterud, A. 2018: A method that accounts for differential
821 detectability in mixed samples of long-term infections with applications to the case of
822 Chronic Wasting Disease in cervids. *Methods in Ecology and Evolution*
823 doi.org/10.1111/2041-210X.13088
- 824 Vincent, J.P., Bideau, E., Hewison, A.J.M. & Angibault, J.M. 1995: The influence of
825 increasing density on body weight, kid production, home range and winter grouping in
826 roe deer (*Capreolus capreolus*). *Journal of Zoology* 236: 371-382.
- 827 Waddell, L., Greig, J., Mascarenhas, M., Otten, A., Corrin, T. & Hierlihy, K. 2018: Current
828 evidence on the transmissibility of chronic wasting disease prions to humans - A
829 systematic review. *Transboundary Emerging Disease* 65: 37-49.

830 Wahlström, L.K. & Liberg, O. 1995: Contrasting dispersal patterns in two Scandinavian roe
1 deer *Capreolus capreolus* populations. *Wildlife Biology* 1: 159-164.
2 831
3
4 832 Wasserberg, G., Osnas, E.E., Rolley, R.E. & Samuel, M.D. 2009: Host culling as an adaptive
5 management tool for chronic wasting disease in white-tailed deer: a modelling study.
6
7 833
8
9 834 *Journal of Applied Ecology* 46: 457-466.
10
11 835 Wild, M.A., Hobbs, N.T., Graham, M.S. & Miller, M.W. 2011: The role of predation in
12 disease control: A comparison of selective and nonselective removal on prion disease
13
14 836 dynamics in deer. *Journal of Wildlife Diseases* 47: 78-93.
15
16 837
17
18 838 Williams, A.L., Kreeger, T.J. & Schumaker, B.A. 2014: Chronic wasting disease model of
19 genetic selection favoring prolonged survival in Rocky Mountain elk (*Cervus*
20
21 839 *elaphus*). *Ecosphere* 5: 1-10.
22
23 840
24
25 841 Williams, E.S. 2005: Chronic wasting disease. *Veterinary Pathology* 42: 530-549.
26
27 842 Williams, E.S., Miller, M.W., Kreeger, T.J., Kahn, R.H. & Thorne, E.T. 2002: Chronic
28
29 843 Wasting Disease of deer and elk: A review with recommendations for management.
30
31 844
32
33 845 *Journal of Wildlife Management* 66: 551-563.
34
35 846 Williams, E.S. & Young, S. 1980: Chronic wasting disease of captive mule deer: a
36
37 847 spongiform encephalopathy. *Journal of Wildlife Diseases* 16: 89-98.
38
39 848 Williams, E.S. & Young, S. 1992: Spongiform encephalopathies in Cervidae. *Rev Sci Tech*
40
41 849
42
43 850 11: 551-567.
44
45 851 Wolfe, L.L., Watry, M.K., Sirochman, M.A., Sirochman, T.M. & Miller, M.W. 2018:
46
47 852 Evaluation of a test and cull strategy for reducing prevalence of Chronic Wasting
48
49 853 Disease in mule deer (*Odocoileus hemionus*). *Journal of Wildlife Diseases* 54: 511-
50
51 854 519.
52
53
54
55
56
57
58
59
60
61
62
63
64
65