

1 **RESPIRATION-RELATED CEREBRAL BLOOD FLOW VARIABILITY**
2 **INCREASES DURING CONTROL-MODE NON-INVASIVE VENTILATION IN**
3 **NORMOVOLEMIA AND HYPOVOLEMIA**

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24
25 **Running head:** CBF variability increases during controlled ventilation

26

26 **Abstract**

27 **Purpose** Increased variability in cerebral blood flow (CBF) predisposes to adverse
28 cerebrovascular events. Oscillations in arterial blood pressure and PaCO₂ induce CBF
29 variability. Less is known about how heart rate (HR) variability affects CBF. We
30 experimentally reduced respiration-induced HR variability in healthy subjects, hypothesizing
31 that CBF variability would increase.

32 **Methods** Internal carotid artery (ICA) blood velocity was recorded by Doppler ultrasound in
33 ten healthy subjects during baseline, control-mode, non-invasive mechanical ventilation
34 (NIV), i.e. with fixed respiratory rate, hypovolemia induced by lower body negative pressure,
35 and combinations of these. ICA beat volume (ICABV) and ICA blood flow (ICABF) were
36 calculated. HR, mean arterial blood pressure (MAP), respiratory frequency (RF), and end-
37 tidal CO₂ were recorded. Integrals of power spectra at each subject's RF±0.03Hz were used to
38 measure variability. Phase angle/coherence measured coupling between cardiovascular
39 variables.

40 **Results** Control-mode NIV reduced HR variability (-56%, p=0.002) and ICABV variability (-
41 64%, p=0.006) and increased ICABF variability (+140%, p=0.002) around RF.
42 NIV+hypovolemia reduced variability in HR and ICABV by 70–80% (p=0.002) and doubled
43 ICABF variability (p=0.03). MAP variability was unchanged in either condition. Respiration-
44 induced HR and ICABV oscillations were in inverse phase and highly coherent
45 (coherence>0.9) during baseline, but this coherence decreased during NIV, in normovolemia
46 and hypovolemia (p= 0.01).

47 **Conclusion** Controlling respiration in awake healthy humans reduced HR variability and
48 increased CBF variability in hypovolemia and normovolemia. We suggest respiration-induced
49 HR variability to be a mechanism in CBF regulation. Maintaining spontaneous respiration in
50 patients receiving ventilatory support may be beneficial also for cerebral circulatory purposes.

51

52 **Key words:** internal carotid artery, cerebral blood flow, respiration, heart rate variability,
53 positive pressure ventilation, hypovolemia.

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55

56 **Abbreviation list**

57	CBF	cerebral blood flow
58	ETCO ₂	end-tidal carbon dioxide
59	HF	high frequency
60	HR	heart rate
61	HRV	heart rate variability
62	ICA	internal carotid artery
63	ICABF	internal carotid artery blood flow
64	ICABV	internal carotid artery beat volume
65	MAP	mean arterial pressure
66	NIV	non-invasive ventilation
67	PaCO ₂	arterial partial pressure of carbon dioxide
68	PEEP	positive end expiratory pressure
69	RF	respiratory frequency
70	RSA	respiratory sinus arrhythmia
71	95% CI	95% confidence intervals

72

73

73 **Introduction**

74 Cerebral blood flow (CBF) preservation is an important clinical target in critically ill patients.
75 Traditionally, preserving patient mean arterial blood pressure (MAP) in the plateau region of
76 the CBF autoregulation curve and maintaining normocapnia has been a strategy for ensuring
77 adequate CBF. However, as both arterial blood pressure and the arterial partial pressure of
78 CO₂ (P_aCO₂) in humans are highly dynamic, there is an increasing interest in CBF variability
79 and its potential clinical significance. CBF variability has been extensively studied in relation
80 to arterial blood pressure variability, in order to assess the dynamic cerebral autoregulation.
81 Blood pressure variations have been connected to increased stroke incidence, brain damage,
82 and poor health outcomes (Jung and Kim 2013; Shimbo et al. 2012; Tatasciore et al. 2007).
83 Increased CBF variability is one of the mechanisms thought to underlie the relationship
84 between elevated blood pressure variability and brain dysfunction (Tzeng and MacRae 2013),
85 and thus to mediate stroke complications in patients (Jung and Kim 2013).

86 Cerebral autoregulation is less effective above 0.08 Hz (Zhang et al. 1998) and most effective
87 below 0.05 Hz (Claassen et al. 2016). High-frequency (HF, 0.15–0.4 Hz) oscillations in MAP,
88 resulting from HF oscillations in cardiac output, are thus considered to be transmitted to the
89 brain vasculature. However, little CBF variability is observed in the HF range in
90 spontaneously breathing healthy individuals (Rickards and Tzeng 2014). Kuo et al report a
91 HF component in middle cerebral artery velocity variability, however constituting only 16%
92 of its total variability (Kuo et al. 1998); the HF arterial blood pressure variability being
93 approximately 21%. Spontaneous respiration contributes significantly to haemodynamic
94 variability in the HF range (0.15–0.4 Hz), inducing fluctuations in heart rate (HR), left cardiac
95 stroke volume, cardiac output and MAP (Toska and Eriksen 1993). The observed low CBF
96 variability, contrasted by the significant respiration-induced hemodynamic variability of the
97 central circulation, leads to the speculation that a regulatory mechanism minimizes CBF
98 variations in the HF interval. Respiratory sinus arrhythmia (RSA), the HR variability (HRV)
99 in the HF interval, has been shown to stabilize cardiac output and thus MAP in the HF range
100 (Elstad et al. 2015; Toska and Eriksen 1993).

101 Several modelling methods have been employed in order to study the effect of beat-to-beat
102 MAP and breath-to-breath end-tidal CO₂ (ETCO₂) fluctuations on CBF variability (Mitsis et
103 al. 2004; Panerai et al. 2000). The effect of respiration and HRV on CBF variability however
104 is not well documented. Reduced short-term HRV has been observed in patients with

105 cardiovascular and cerebrovascular pathological conditions (Akil et al. 2015; Nagata et al.
106 2006). Reduced HRV is an independent prospective marker of stroke risk in healthy
107 individuals (Binici et al. 2011). Experimentally, HRV can be reduced by control-mode non-
108 invasive ventilatory support (NIV) (Elstad et al. 2015). The HF spectral power calculated
109 from power spectral analysis is an important tool for measurement of HRV (Larsen et al.
110 2010).

111 The purpose of this study was to examine the effect of spontaneous respiration and HRV on
112 CBF variability during normovolemia and mild hypovolemia. Internal carotid artery blood
113 flow (ICABF) variability at each subject's respiratory frequency (RF) was assessed in relation
114 to the respiration-induced variations in HR and MAP, using frequency domain analysis. Since
115 HRV in the HF-range (HF-HRV) during spontaneous breathing has been shown to reduce
116 variations in cardiac output (Elstad et al. 2015), which directly influences CBF (Ogoh et al.
117 2005), we hypothesized that respiratory-induced HRV (RF-HRV) may reduce ICABF
118 variability as well. To test our hypothesis, we trained subjects to accept low-pressure control-
119 mode NIV (i.e., with fixed tidal volume and respiratory rate) to prevent spontaneous initiation
120 of breathing and reduce HRV. The effects of reduced HRV on ICABF were studied both in
121 normovolemia and in hypovolemia induced by lower body negative pressure (LBNP).
122 Hypovolemia and controlled ventilation is a commonly encountered clinical situation e.g. in
123 the operating theatre; increased CBF variability under these circumstances may affect patient
124 outcome and result in adverse cerebrovascular effects. To the best of our knowledge, the
125 effects of hypovolemia and mechanical ventilation on CBF variability have only been studied
126 separately previously. In this protocol we investigated their combined effect on ICABF
127 variability.

128

128 **Materials and methods**

129 Steady-state data from these experiments have been published (Skytjoti et al. 2016). In this
130 study, we investigated the oscillatory responses of the cerebrovascular variables. Fifteen
131 young healthy volunteers were recruited and gave written informed consent to participate in
132 the study. All procedures were performed according to the Declaration of Helsinki. The
133 regional ethics committee approved the protocol and the procedures (NO: 2014/2228,
134 December 2014).

135 None of the subjects was a smoker or taking any medication. The subjects were instructed to
136 abstain from caffeinated beverages and strenuous physical activity for 12 hours before the
137 experiment. They also avoided food and drink for two hours and alcohol for 24 hours before
138 the experiment.

139 We analysed recordings from each subject in four different experimental conditions:
140 spontaneous breathing+normovolemia (baseline), spontaneous breathing+hypovolemia
141 (hypovolemia), control-mode NIV+normovolemia (NIV), and control-mode
142 NIV+hypovolemia. Five subjects were excluded because we did not obtain technically
143 successful recordings in all four experimental conditions, i.e., at least 2 minutes of continuous,
144 artefact-free beat-by-beat recordings of all cardiovascular variables. Thus, the analyses
145 presented here are from 10 subjects (4 males, 6 females), median age 22 years (range 20–30
146 years).

147 *Experimental protocol*

148 The experimental protocol is described in detail in a previous paper (Skytjoti et al. 2016). The
149 experimental challenges were control-mode (i.e no spontaneous breaths), pressure-regulated,
150 volume-controlled NIV (VIVO50, Diacor a/s, Norway) and LBNP (Hisdal et al. 2003), alone
151 or in combination. The subjects lay supine in an LBNP chamber wearing a facemask
152 throughout the procedure. During parts of the experiment NIV was applied with mandatory
153 respiratory frequency, tidal volume, positive end-expiratory pressure (PEEP), and maximal
154 inspiratory pressure set to match each subject's natural respiratory pattern. Spontaneous
155 inspiration was prevented by training the subjects in a preparatory session not to initiate
156 inspiration, but to accept the frequency and tidal volume administered by the ventilator.
157 Ventilator settings (median (range)) were: RF: 14 breaths per min (11–16); target tidal volume:
158 650mL (500–850). Maximum and minimum inspiratory pressures were set to 14 cmH₂O and
159 4.5 cmH₂O respectively for all subjects. PEEP values of 1.3 cmH₂O (0.9-1.4) were recorded

160 during normovolemia and 1.2 (0.8-1.4) during hypovolemia. The periods of LBNP (-30
161 mmHg, induced over 0.3 sec (Hisdal et al. 2003)) generated an acute central blood volume
162 shift corresponding to a depletion of 10–20% of the total blood volume (500–1000 mL) and
163 resulting in mild to moderate central hypovolemia (Hisdal et al. 2003).

164 An initial 10-min baseline period of normovolemia was followed by 10-min of simulated
165 central hypovolemia. A 10-minute recovery period of normovolemia followed. During each
166 interval, the subjects breathed spontaneously for 5 minutes and were subjected to control-
167 mode NIV for 5 minutes (Fig.1). Each subject underwent the procedure twice, with a few
168 minutes' pause between rounds. The first 30-min round was randomized to start with either
169 NIV or spontaneous breathing; in the second round the sequence was reversed.

170 *Recordings*

171 Mean blood velocity in the ICA (5 MHz probe, insonation angle: 45°, SD-100, Vingmed
172 Sound, Horten, Norway) was recorded beat-by-beat by a trained operator using Doppler
173 ultrasound. ICA was chosen over the widely used middle cerebral artery because the diameter
174 of the vessel was needed for blood flow calculations. The diameter of the subject's right ICA
175 was measured approximately 2 cm above the bifurcation of the common carotid artery (Willie
176 et al. 2012) before the beginning of the recordings using B-mode Ultrasound (10 MHz and 2.5
177 MHz, System Five, GE Vingmed Sound, Norway). Non-invasive finger arterial pressure was
178 recorded continuously (Finometer, Finapres Medical System, Netherlands), and beat-by-beat
179 MAP was calculated by numerical integration. Respiration was recorded breath-by-breath
180 with an elastic belt around the abdomen (Respiration and Body position Amplifier, Scan-Med
181 a/s, Norway). The signal from the belt resulting from the stretch is referred to as respiration.
182 The RF at maximal power was used in the calculations. HR was calculated from the R-R
183 interval in a three-lead ECG sampled at 300Hz. ETCO₂ in the facemask was recorded by a
184 capnograph (Artema MM201, Artema Medical AB, Sweden). Blood velocity wave,
185 respiration band signal, arterial blood pressure curve, stroke volume (Finometer), LBNP,
186 ETCO₂ and room temperature were sampled at 100 Hz. Mean ICA blood velocity, HR (ECG)
187 and MAP were also sampled beat-by-beat. The recordings were transferred on-line to a
188 recording computer running dedicated data collection and analysis software (Program for real-
189 time data acquisition, Morten Eriksen, Norway). ICA beat volume (ICABV) was calculated
190 beat-by-beat from blood velocity and the diameter of the ICA. ICABF was calculated beat-by-
191 beat from ICABV multiplied by instantaneous HR.

192 *Mathematical and statistical analyses*

193 All data was resampled at 4Hz employing an interpolation scheme. For each experimental
194 condition, we selected a continuous sequence of 2 minutes after manual inspection for
195 artefacts. The power spectrum, which is a plot of the variance of a time series as a function of
196 frequency, was computed by the Fast Fourier Transform algorithm. The frequency resolution
197 was kept the same in all conditions ($\sim 0.0083\text{Hz}$). Coherence analysis, which provides a
198 description of the relationship between two fluctuating variables in a specified range of
199 frequencies, was used to examine the association between cardiovascular variables. Using
200 cross-spectral analysis, it is possible to examine whether the variability of two distinct time
201 series is interrelated in the frequency domain.

202 The integrals (area under the curve) of the power spectra were calculated in the subsequent
203 three frequency intervals as estimates of variability in each frequency band. First, respiration-
204 induced variability in cardiovascular variables was calculated at each subject's peak RF \pm
205 0.03 Hz (RF-variability). Since spontaneous RF varied between subjects from 0.14 Hz to 0.3
206 Hz, we moved the RF-interval (RF \pm 0.03Hz) accordingly. The percentage of the total
207 respiration power included in the RF \pm 0.03Hz interval was calculated on the signal from the
208 Respiration and Body position Amplifier. In spontaneous breathing states this interval
209 included almost 80% of the respiration power (normovolemia: 77% (56%-85%), hypovolemia:
210 78% (65%-85%)). During control-mode NIV all respiration power was included in the
211 RF \pm 0.03Hz interval.

212 Integrals were also calculated for the HF interval (0.15–0.4 Hz, HF-variability), which is
213 widely used for the study of respiratory variability. Third, the total variability in the interval
214 0.05–0.5 Hz was calculated in order to quantify the changes in respiratory-induced variability
215 of the cardiovascular variables relative to the overall variability. The relative RF-variability of
216 each cardiovascular variable was then calculated as the fraction of the RF-variability (RF \pm
217 0.03Hz) to the total variability.

218 Interaction between cardiovascular variables was examined by computing phase angles and
219 coherence from the cross-spectra at peak RF (RF \pm 0.02 Hz), for the following pairs of
220 variables: Respiration–HR, Respiration–ICABV, Respiration–ICABF, HR–ICABV, and
221 MAP–ICABF. The phase angle is the time delay between two waves that oscillate at the same
222 frequency. Coherence provides a measure of the coupling between two signals over the range
223 of frequencies examined.

224 Averaged phase angles were calculated by weighting the phase angles with their squared
225 coherence. Two oscillating variables were considered to be in phase when the absolute phase
226 difference between them was less than 45° (0.79 rad) and in inverse phase when the absolute
227 phase difference was more than 135° (2.35 rad). Waves that meet in antiphase weaken each
228 other, a phenomenon called destructive interference. This has been observed for HR and
229 cardiac stroke volume at the RF, in effect diminishing cardiac output oscillations (Toska and
230 Eriksen 1993).

231 Numbers are medians and 95% confidence intervals (95% CI) calculated by Hodges-
232 Lehmann's estimate if not otherwise specified. For illustration purposes, integrals calculated
233 for each experimental condition were also normalized with respect to the median value during
234 baseline. Wilcoxon matched-pairs signed-rank test against a two-sided alternative (Hollander
235 and Wolfe 1999) was used to test the differences in ICABF variability between conditions
236 (StatXact, Cytel Studio 10, Cytel Inc., Cambridge, MA, USA) and the level of significance
237 was set at $p=0.05$ before analysis. In addition we report the p-values (calculated after analysis)
238 for the change in variability of the rest of the cardiovascular variables for informative
239 purposes.

240 **Results**

241 All ten subjects completed the protocol successfully and tolerated the abruptly induced central
242 hypovolemia without signs of presyncope. The ten subjects complied successfully with the
243 ventilator; they did not initiate inspiration but accepted passively the fixed respiratory rate and
244 tidal volume administered by the ventilator. ETCO_2 was decreased during control mode NIV
245 due to the slight hyperventilation which was necessary in order to diminish spontaneous
246 inspiratory effort. However, ETCO_2 and RF were stable during each of the 2-min time
247 intervals selected for frequency analysis in each experimental condition.

248 Raw data from one subject (recordings of ICABF, HR and ICABV) during the procedure are
249 presented in Fig.2.

250 *Cerebrovascular variability during baseline*

251 Table 1 shows steady state values of the studied variables during the four different
252 experimental states. Synchronous high-amplitude oscillations in HR and ICABV were
253 observed during baseline whereas little variability was observed in ICABF over the
254 respiratory cycle (Fig. 2, Fig. 3a). Inspiration coincided with an increase in HR and a decrease
255 in ICABV, and expiration with a decrease in HR and an increase in ICABV (Fig 3a).

256 The total, HF, and RF variability in HR and ICABV were maximal during spontaneous
257 breathing+normovolemia (Table 2, Fig 4a). The relative RF-HRV was 39% (23%–45%) of
258 the total variability (measured over the entire 0.05–0.5 Hz interval). Thus, respiration-
259 centered HRV is an important part of HRV. In contrast, the relative RF-ICABF variability
260 during baseline constituted only 13% (11%–15%) of the total ICABF variability.

261 Table 3 shows the changes in phase angle and coherence between the conditions for the
262 chosen pairs of variables. Respiration and HR oscillations were in phase and highly coherent
263 (median coherence: 0.93). Respiration was in inverse phase and highly coupled with ICABV
264 oscillations (median coherence: 0.94). Similarly, HRV and ICABV oscillations were in
265 inverse phase and highly coherent (median coherence: 0.90), the increases in HR coinciding
266 with drops in ICABV (Fig. 2a). These effects were also demonstrated by the power spectra,
267 where maximum power was observed for HR and ICABV at the RF during baseline. In
268 contrast, small variability in MAP and ICABF was observed (Fig 5a).

269 Coherence between MAP and ICABF was very low during spontaneous
270 breathing+normovolemia (Table 3), the condition when both these variables reached their

271 lowest variability. Low coherence may indicate a nonlinear relationship, an absence of a
272 relationship between two signals (Zhang et al. 1998), or a low signal-to-noise ratio, which is
273 common with spontaneous oscillations (Claassen et al. 2009).

274 *Effect of control-mode NIV on cerebrovascular variability*

275 A significant drop in HRV in all frequency bands, in HF-ICABV variability and in RF-
276 ICABV variability occurred during control-mode NIV (Table 2, Fig. 4b). Relative RF-HRV
277 was 33% (12%–40%), similar to during baseline. The decrease in the respiratory-induced
278 oscillations of HR and ICABV is also depicted in Fig. 3b.

279 RF-ICABF variability increased during control-mode NIV by about 140% (Table 2, Fig. 4b).
280 In eight out of ten subjects, a peak in the power spectrum of ICABF was observed at the set
281 RF (Fig. 5b); this peak was absent during spontaneous breathing. A similar peak at the set RF
282 was observed for MAP variability in six of the ten subjects (Fig. 5b). The relative RF-ICABF
283 variability was increased to 33% (24%–37%, $p=0.002$), compared to 13% at baseline.

284 Despite the reduction in RSA magnitude during control-mode NIV, respiration and HR
285 oscillations remained in phase and were highly coherent (median coherence: 0.91). In contrast,
286 coherence between respiration and ICABV as well as between HR and ICABV decreased
287 (Table 3) compared to during baseline.

288 *Effect of hypovolemia on cerebrovascular variability*

289 Both HF-ICABV variability and RF-ICABV variability decreased during hypovolemia (Table
290 2, Fig. 4c). The relative RF-HRV was 36% (19%–44%). ICABF variability did not change
291 with hypovolemia in either frequency interval. The relative RF-ICABF variability was 18%
292 (14%–21%), slightly increased from baseline ($p=0.05$).

293 RSA was maintained during hypovolemia (HR in phase with respiration and highly coherent,
294 Table 3). ICABV oscillations was in inverse phase and highly coherent with both respiration
295 and HR oscillations (Table 3).

296 *Effect of combined control-mode NIV and hypovolemia on cerebrovascular variability*

297 HF-HRV and RF-HRV were about 70–80% lower during control-mode NIV+hypovolemia
298 than during baseline, as was HF- and RF-ICABV variability (Table 2, Fig. 4d). In contrast,
299 RF-ICABF variability increased by about 100%. The relative RF-HRV was 22% (10%–37%)

300 of the total, i.e. lower than during baseline, and the relative RF-ICABF variability was 24%
301 (6%–30%).

302 During combined control-mode NIV and hypovolemia, an increased respiratory component
303 in ICABF variability appeared around the set RF in eight out of ten subjects (Fig. 5d), similar
304 to during NIV in normovolemia. No such peak in ICABF variability was present during
305 baseline or during hypovolemia alone, i.e. in conditions with spontaneous breathing. A similar
306 peak in MAP variability at the set RF was observed in seven of the ten subjects (Fig. 5d), but
307 on a group level, MAP variability did not differ significantly between experimental conditions
308 in either frequency interval (Table 2, Fig. 4d).

309 Coherence and phase angle between respiration and HR did not change, indicating the
310 presence of RSA though with a low magnitude. In contrast, the coherence between respiration
311 and ICABV, HR and ICABV and respiration and ICABF reached their lowest values in this
312 state (Table 3).

313 Discussion

314 In the present study we evaluated respiratory-induced HRV as a possible regulatory
315 mechanism of CBF and examined the effect of respiratory variations on CBF during
316 combined control-mode NIV and hypovolemia. We found that respiratory-induced ICABF
317 variability in spontaneously breathing subjects was minimal, constituting only a small fraction
318 of total ICABF variability both during normovolemia and hypovolemia. Control-mode NIV
319 reduced respiratory HRV and increased respiratory variability in ICABF, during
320 normovolemia and hypovolemia. We suggest that respiratory HRV during spontaneous
321 breathing minimizes CBF variability. This finding might be of clinical importance as
322 increased CBF variability has been connected to adverse cerebrovascular events.

323 During spontaneous breathing, HRV counteracted ICABV fluctuations, as drops in ICABV
324 coincided with increases in HR during inspiration, and vice versa during expiration, thus
325 minimizing fluctuations in ICABF. In contrast, during control-mode NIV with mandatory set
326 RF and tidal volume, respiration-induced variability in ICABF was increased by 140% during
327 NIV+normovolemia and 100% during NIV+hypovolemia. The coupling between respiration-
328 induced oscillations in ICABV and HR decreased markedly during control-mode NIV (both
329 in normovolemia and hypovolemia), from the situation of highly-coupled, inverse-phase
330 oscillations during spontaneous breathing+normovolemia. Our study thus demonstrates a
331 connection between decreased respiration-induced HRV and increased ICABF variability that
332 may be of clinical relevance.

333 Both cardiac stroke volume and cardiac output have been shown to be linearly related to
334 middle cerebral artery blood velocity (Bronzwaer et al. 2014; Ogoh et al. 2005). Increased
335 oscillations in these variables could therefore be linearly transmitted to middle cerebral artery
336 blood velocity and thus to CBF. We demonstrated that despite respiration-synchronous
337 oscillations in ICABV during spontaneous breathing, ICABF variability was minimal due to
338 buffering of the ICABV variability by HRV, which stabilized ICABF over the respiratory
339 cycle.

340 Possible physiological roles of RSA have been investigated both in humans and animal
341 models. Artificially induced RSA in a dog model optimized gas exchange and oxygen
342 transport and reduced intrapulmonary shunt (Hayano et al. 1996), but this could not be
343 demonstrated in humans (Tzeng et al. 2009). RSA has also been shown to buffer respiratory
344 variability in left cardiac output, thus stabilizing MAP and systemic flow (Elstad 2012; Elstad

345 et al. 2015). Minimization of cardiac work has been proposed as another function of RSA
346 (Ben-Tal et al. 2012). We here suggest minimization of respiration-related CBF variability as
347 an additional physiological role of RSA.

348 In the present study, control-mode NIV reduced RSA substantially compared to the
349 physiologic setting of spontaneous breathing. The main mechanism for this decrease in RSA
350 was probably the reduction in the central feed-forward drive due to the elimination of
351 spontaneous inspiratory effort (Beda et al. 2012; Elstad et al. 2015). The positive intrathoracic
352 pressure during NIV may have affected the pulmonary and cardiac stretch receptors. In pigs,
353 controlled mechanical ventilation reduced RSA amplitude and cardioventilatory coupling
354 compared to assisted mechanical ventilation (Beda et al. 2012). Our study thus, in line with
355 previous reports, shows that controlled mechanical ventilation reduces RSA, making control-
356 mode NIV a good protocol to explore the effects of RSA on ICABF variability in human
357 subjects.

358 Hypovolemia exaggerates the effects of positive pressure ventilation on the circulation. The
359 combination of hypovolemia and control-mode mechanical ventilation is frequent in the
360 operating theatre and not uncommon among critical care patients; thus our findings of
361 increased variability in cerebrovascular flow under such circumstances are of interest.
362 Because HRV decreases during hypovolemia (Elstad and Walloe 2015), the combination of
363 controlled ventilation and hypovolemia was hypothesized to induce a larger drop in RSA.
364 This hypothesis was verified (Table 2). However, despite a greater reduction in RSA during
365 control-mode NIV+hypovolemia than during NIV alone, CBF variability in our study was
366 similar in these two states.

367 Positive pressure ventilation and PEEP may affect cardiac stroke volume by impeding right
368 ventricular filling. Our study employed very low PEEP settings that did not likely impact
369 stroke volume much. In contrast hypovolemia induced by LBNP reduced stroke volume
370 markedly (Skytjoti et al. 2016).

371 It has been argued that increased CBF oscillations are not necessarily harmful for the brain
372 (Rickards and Tzeng 2014). Increased middle cerebral artery velocity oscillations have been
373 suggested to delay presyncopal symptoms during progressive hypovolemia in subjects
374 breathing through an inspiratory threshold device (Rickards et al. 2007). Amplification of the
375 respiratory pump with the inspiratory threshold device produced increased oscillations in CBF
376 velocity in their study, resembling the high-amplitude respiration-induced ICABV oscillations

377 we observed during baseline (Fig. 1, Fig. 2a). In contrast, the use of control-mode NIV in the
378 present study prevented spontaneous respiration and reduced the spontaneous RF- and HF-
379 ICABV variability. Concurrently, an increased respiratory variability component appeared in
380 ICABF.

381 During control-mode NIV, RF-ICABF variability increased, and a decrease in coupling
382 between HR and ICABV was observed. The relative RF-ICABF variability was also higher
383 during controlled than spontaneous breathing, indicating that the respiratory-induced CBF
384 variations became a more important component of total CBF variability during controlled
385 ventilation. Both the decreased coupling between HRV and ICABV variability and the cyclic
386 changes in intrathoracic pressure during NIV may have been responsible for the increase in
387 RF-ICABF variability. The effects of mechanical ventilation on the circulation are more
388 pronounced the higher the mean airway pressure, as higher intrathoracic pressure impedes
389 venous return and decreases right ventricular filling. Hypovolemia will exaggerate these
390 effects (Cheifetz 2014). However, while each cyclic increase in intrathoracic pressure
391 increases right ventricular afterload, it also improves left ventricular filling and decreases left
392 ventricular afterload. Thus, during hypovolemia, mechanical ventilation induces larger
393 respiratory oscillations in cardiac output (Michard 2005; Rimehaug et al. 2016), and these
394 may be transferred to the CBF. We demonstrated an increase in RF-ICABF variability during
395 control-mode NIV. Increased variability in MAP and cardiac output may have contributed
396 though we were unable to demonstrate this in our study.

397 *Clinical implications*

398 HRV is affected by several pathophysiological conditions. Reductions in HRV have been
399 associated with poor clinical outcomes after acute myocardial infarction (Balanesu et al.
400 2004) and have been reported to be a predictor of post stroke mortality (Makikallio et al.
401 2004). Several studies have linked lower HRV with brain dysfunction (Biswas et al. 2000;
402 Katz-Leurer et al. 2014; Kholod et al. 2013). A decrease in HRV was associated with
403 cerebrovascular pathology in normotensive diabetic patients (Nagata et al. 2006).

404 In healthy spontaneously breathing subjects, minimal variability is observed in CBF (Kuo et
405 al. 1998; Rickards et al. 2007). We found that ICABF oscillations increased around the set RF
406 during control-mode NIV, both in normovolemia and in hypovolemia. This observation could
407 be of clinical importance as it may indicate that CBF regulation might be disturbed by the
408 applied therapy in patients treated with controlled mechanical ventilation for extended periods

409 of time. Elderly patients in particular may be more vulnerable, also due to a higher prevalence
410 of medical comorbidities. A recent study indicated an association between impaired
411 intraoperative CBF autoregulation and postoperative cognitive dysfunction in patients over 65
412 years old (Goettel et al. 2017).

413 Associations between cerebral pathology and exaggerated blood pressure variations have been
414 reported and possible underlying mechanisms are larger CBF fluctuations, neurohumoral
415 activation, endothelial dysfunction, inflammatory mediators, and oxidative stress (Jung and
416 Kim 2013; Shimbo et al. 2012; Tatasciore et al. 2007; Tatasciore et al. 2008). Optimizing
417 intravascular volume, avoiding excess sedation, and using lung-protective ventilation
418 strategies with supported rather than controlled ventilation may contribute to improved
419 cerebral outcomes in critical care patients.

420 *Considerations*

421 CBF is highly dependent on $P_a\text{CO}_2$ (Panerai et al. 2010) and any kind of physical stimulation.
422 The experiments were performed in a quiet room, in resting subjects who had familiarized
423 themselves with the equipment and procedures. To minimize confounding factors, we
424 recorded ETCO_2 as an estimate of the $P_a\text{CO}_2$ and ensured that there were no significant
425 changes during each 2-min time interval selected for frequency analysis in each experimental
426 state. However there was a decrease in ETCO_2 from baseline to NIV and to
427 NIV+hypovolemia. Since ETCO_2 necessarily can be determined only once per breath, we
428 could not study the effect of ETCO_2 fluctuations on ICABF variability in the HF range. To
429 evaluate whether cyclical changes in arterial partial pressure of CO_2 actually affect CBF
430 would require continuous invasive measurements. It has been shown that the effect of breath-
431 by-breath ETCO_2 fluctuations on the middle cerebral artery blood flow velocity are
432 considerable in the very low frequencies and in the low frequencies, implying that slow
433 variations in ETCO_2 have a larger impact on middle cerebral artery blood flow velocity
434 (Mitsis et al. 2006). Mitsis et al showed that ETCO_2 variations have a considerable effect on
435 CBF at frequencies below 0.04 Hz (Mitsis et al. 2004). Taking into account the negligible
436 effect of ETCO_2 in the HF range, we assumed that ETCO_2 fluctuations did not affect
437 cerebrovascular respiratory variability.

438 Another methodological concern was whether the diameter of the ICA, which was measured
439 once in the beginning of each experiment, remained stable throughout the experiment or
440 changed in response to changes in ETCO_2 and LBNP. $P_a\text{CO}_2$ change <1.3 kPa is considered

441 unlikely to cause ICA diameter changes (Sato et al. 2012; Willie et al. 2012), and no change
442 in ICA diameter was reported between baseline and -35mmHg LBNP (Ogoh et al. 2015). As
443 the largest change in ETCO₂ in our experiments was 1.1 kPa and the LBNP level was -30
444 mmHg we assumed that ICA diameter remained the same.

445 We calculated RF variability at subjects' peak $RF \pm 0.03\text{Hz}$ in addition to in the HF interval. In
446 spontaneously breathing subjects, the RF varies with time, and therefore spontaneous
447 respiratory variability may be widely distributed. To take this into account, we calculated the
448 percentage of the total respiration power (respiration signal) included in the $RF \pm 0.03\text{Hz}$
449 interval. Almost 80% of the respiration power was included. During control-mode NIV with
450 or without hypovolemia, all respiratory variability was included in the $RF \pm 0.03\text{Hz}$ interval.
451 The measured change in RF-HRV would probably have been larger if it was calculated over a
452 wider interval during spontaneous breathing.

453 Our findings during NIV directly apply only if conditions are similar, i.e. healthy subjects, no
454 sedation or paralytics, marginal PEEP, controlled respiratory rate, and low-grade pressures.
455 The subjects did not initiate inspiration, but our study design did not hinder diaphragmatic
456 muscle activity participating in the respiratory cycle. Thus, the applied interventions were
457 milder than those often used in clinical settings, where more pronounced effects of controlled
458 mechanical ventilation would be expected. This is suggested also by animal studies (Beda et
459 al. 2012).

460 *Conclusion*

461 Respiratory and cardiovascular variability may introduce changes in cerebral hemodynamics
462 despite several counteracting mechanisms, such as the cerebral autoregulation of CBF. We
463 suggest that respiration-induced HRV could be an additional mechanism that contributes to
464 stabilization of short-term CBF variability in spontaneously breathing, healthy individuals. A
465 reduction in HRV, experimentally induced in our study by control-mode NIV, was
466 accompanied by a significant increase in CBF variability in both normovolemia and mild
467 hypovolemia.

468

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470

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474

475 **Disclosures**

476 The authors declare that there is no conflict of interest.

477

478 **Ethical approval**

479 All procedures performed in the study involving human participants were in accordance with
480 the ethical standards of the institutional and /or national research committee and with the 1964
481 Helsinki declaration and its later amendments or comparable ethical standards.

482

483 **Authors' contributions**

484 All authors have contributed to the design of the study as well as the acquisition, analysis and
485 interpretation of data. M. S. drafted the manuscript and all authors have revised it critically
486 and approved the version to be published.

487

487

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Table 1. Cardiovascular and respiratory variables in the four different conditions: Normovolemia with and without control-mode NIV, hypovolemia with and without control-mode NIV. Healthy human subjects, N=10.

	Normovolemia		Hypovolemia	
	Spontaneous breathing	Control-mode NIV	Spontaneous breathing	Control-mode NIV
ICABF (ml min ⁻¹)	251 (190–285)	226** (163–253)	228 (179–285)	210** (149–245)
ICABV (ml)	4.8 (3.1–5.4)	4.2* (2.8–5.2)	3.8** (2.7–4.4)	3.3** (2.2–3.9)
HR (bpm)	56.6 (46.6–59.6)	54.7 (45.5–59.4)	64.0** (55.2–67.4)	65.1** (55.9–68.0)
MAP (mmHg)	78.3 (69.9–81.2)	77.4 (71.2–80.7)	80.3 (72.9–83.3)	78.8 (72.2–81.7)
ETCO₂ (kPa)	4.9 (4.4–5.1)	4.3** (4.0–4.6)	4.7** (4.2–4.9)	4.0** (3.5–4.2)

Data are medians and 95% Confidence Intervals calculated by Hodges Lehmann estimate. NIV: non-invasive ventilatory support, ICABF: internal carotid artery blood flow, ICABV: internal carotid artery beat volume, HR: heart rate, MAP: mean arterial pressure, ETCO₂: end-tidal CO₂. Significance level compared to baseline: * p<0.05, **p≤0.01.

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Table 2. Cardiovascular variability estimated from integrals of the power spectra at the respiratory frequency ± 0.03 Hz, in the high-frequency interval (0.15–0.4 Hz) and total variability (0.05–0.5Hz). Healthy human subjects, N=10.

	Normovolemia		Hypovolemia	
	Spontaneous breathing	Control-mode NIV	Spontaneous breathing	Control-mode NIV
Variability at respiratory frequency ± 0.03 Hz				
HR (bpm ²)	4.1 (2.0–5.9)	1.8** (0.6–2.2)	2.6 (1.2–3.7)	1.1** (0.4–1.5)
MAP (mmHg ²)	1.0 (0.6–1.4)	1.1 (0.5–2.0)	1.9 (0.7–4.6)	1.7 (0.6–2.5)
ICABV (ml ²)	0.042 (0.013–0.062)	0.015** (0.006–0.021)	0.013** (0.005–0.021)	0.009** (0.004–0.012)
ICABF (ml min ⁻¹) ²	14.0 (5.1–19.4)	33.7** (12.4–47.4)	25.6 (6.2–32.3)	29.1* (7.6–43.4)
Variability in the high-frequency interval				
HR (bpm ²)	7.4 (3.1–10.9)	2.8** (1.3–3.4)	4.1* (1.7–4.6)	1.8** (0.9–2.5)
MAP (mmHg ²)	2.3 (1.4–2.7)	2.3 (1.2–6.4)	3.9* (1.8–7.1)	4.0 (1.7–9.4)
ICABV (ml ²)	0.065 (0.024–0.094)	0.025* (0.013–0.033)	0.023** (0.01–0.034)	0.015* (0.009–0.021)
ICABF (ml min ⁻¹) ²	44.9 (14.4–65.5)	68.0 (22.0–86.4)	48.0 (18.1–75.8)	47.5 (21.2–85.3)
Total Variability				
HR (bpm ²)	12.7 (5.1–17.4)	4.5** (2.6–5.6)	7.7 (4.7–9.3)	7.2 (2.5–9.2)
MAP (mmHg ²)	9.0 (3.8–11.7)	5.0 (3.9–16.9)	12.0 (3.9–24.0)	12.7 (5.2–25.7)
ICABV (ml ²)	0.100 (0.048–0.135)	0.045 (0.02–0.058)	0.046 (0.02–0.061)	0.034* (0.017–0.047)
ICABF (ml min ⁻¹) ²	100.5 (37.1–158.7)	123.3 (39.9–148.5)	128.8 (37.0–160.1)	97.8 (52.7–322.7)
RF (Hz)	0.23 (0.2–0.24)	0.23 (0.21–0.25)	0.21** (0.18–0.22)	0.23 (0.21–0.25)

Values are medians and 95% confidence intervals calculated by Hodges-Lehmann estimates. NIV: non-invasive ventilatory support, HR: heart rate, MAP: mean arterial pressure, ICABV: internal carotid artery beat volume, ICABF: internal carotid artery blood flow, RF: respiratory frequency. Significance level compared to baseline: * $p < 0.05$, ** $p < 0.01$.

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Table 3. Phase angles and coherences between cardiorespiratory variables during normovolemia and hypovolemia, with and without NIV.

	Normovolemia		Hypovolemia	
	Spontaneous Breathing	Control-mode NIV	Spontaneous Breathing	Control-mode NIV
RESP-HR				
Phase angle	0.46 (0.7,0.3)	0.76 (1.27,0.47)	0.59 (1.06,0.36)	0.79 (1.37,0.63)
Coherence	0.93 (0.87–0.97)	0.91 (0.84–0.94)	0.91 (0.85–0.95)	0.92 (0.88–0.94)
RESP-ICABV				
Phase angle	2.89 (2.70–3.21)	2.79 (2.32–3.23)	3.16 (2.76–3.42)	3.66 (2.55–3.83)
Coherence	0.94 (0.85–0.96)	0.78 (0.48–0.84)*	0.84 (0.70–0.88)	0.62 (0.30–0.77)*
RESP-ICABF				
Phase angle	3.19 (1.87–3.70)	4.67 (3.58–5.02)	4.36 (4.15–4.53)	5.03 (4.26–5.47)
Coherence	0.77 (0.64–0.84)	0.87 (0.69–0.89)	0.70 (0.52–0.73)	0.66 (0.43–0.78)
HR-ICABV				
Phase angle	3.28 (3.16–3.50)	3.77 (3.45–4.12)	3.62 (3.46–4.04)	3.77 (3.15–4.71)
Coherence	0.90 (0.80–0.97)	0.82 (0.53–0.89)*	0.90 (0.76–0.92)	0.65 (0.30–0.76)*
MAP-ICABF				
Phase angle	6.23 (1.54–5.84)	5.30 (4.31–6.09)	0.71 (6.02–1.47)	5.93 (4.77–6.28)
Coherence	0.36 (0.20–0.45)	0.49 (0.31–0.59)	0.45 (0.25–0.54)	0.49 (0.18–0.63)

Number of subjects =10. Coherence-weighted phase angles in rad (on a unit circle from 0 to 2π). Data are medians and 95% confidence intervals calculated by Hodges-Lehmann estimates. NIV: non-invasive ventilatory support, RESP: respiration, HR: heart rate, ICABV: Internal carotid artery beat volume, ICABF: Internal carotid artery blood flow. Significance level compared to baseline: * $p=0.01$.

Figure Captions

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607 **Fig. 1** Study protocol displaying subjects' breathing conditions and lower body chamber
 608 pressure during 10 min of normovolemia, 10 min of simulated hypovolemia and 10 min of
 609 normovolemia–recovery. This sequence was run twice in each subject, once starting with
 610 controlled non-invasive ventilation, i.e. with set respiratory frequency and tidal volume (NIV),
 611 and once starting with spontaneous breathing. LBNP: lower body negative pressure

612 **Fig. 2** Recordings (raw data) from one subject during 10 min of normovolemia and 10 min of
 613 simulated hypovolemia induced by lower body negative pressure, with and without non-
 614 invasive control-mode ventilatory support (NIV). During spontaneous
 615 breathing+normovolemia there were large synchronous fluctuations in heart rate (HR) and
 616 internal carotid artery beat volume (ICABV) but minimal oscillations in internal carotid artery
 617 blood flow (ICABF). Controlled mechanical ventilation and hypovolemia, separately and
 618 combined, reduced HR and ICABV oscillations

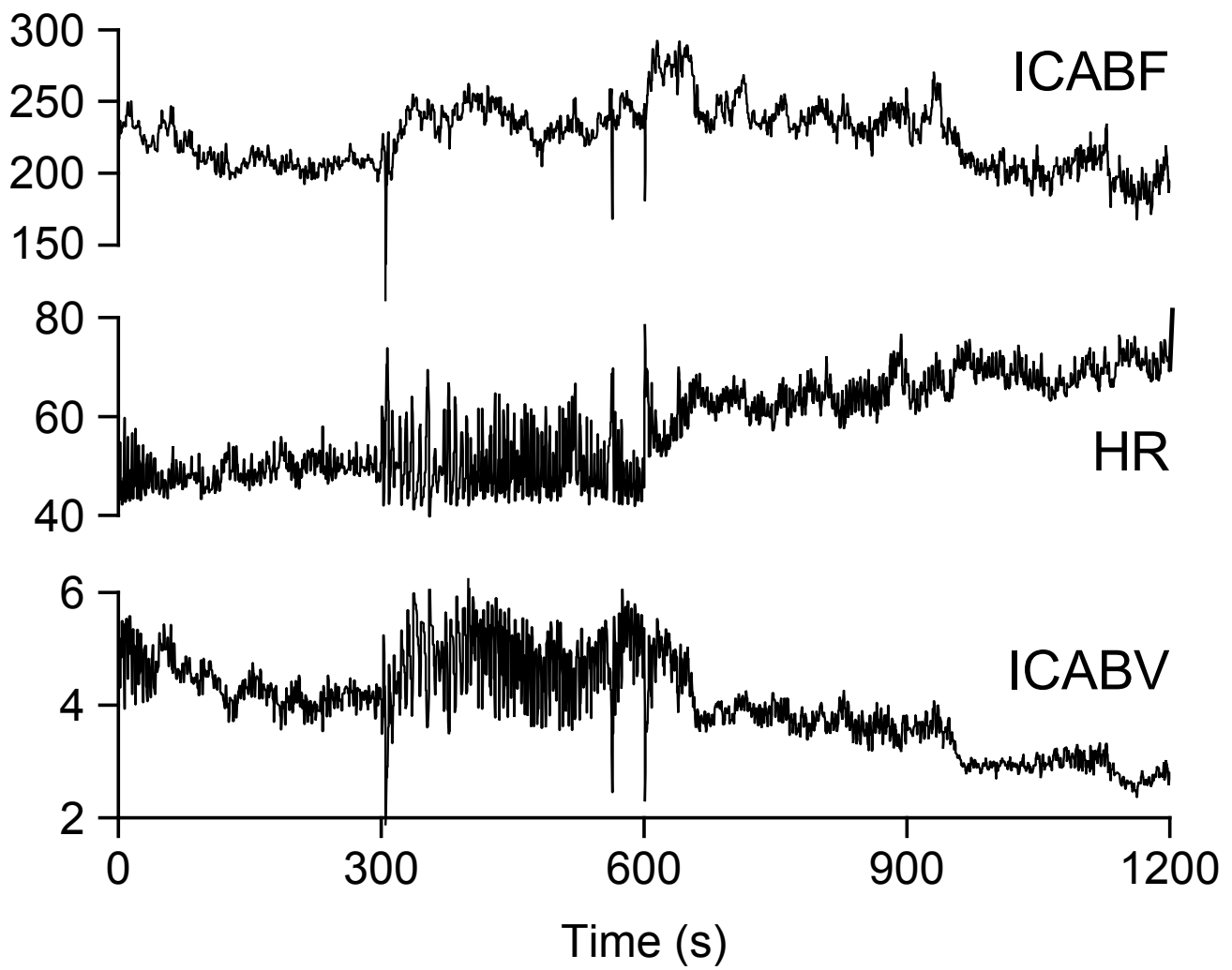
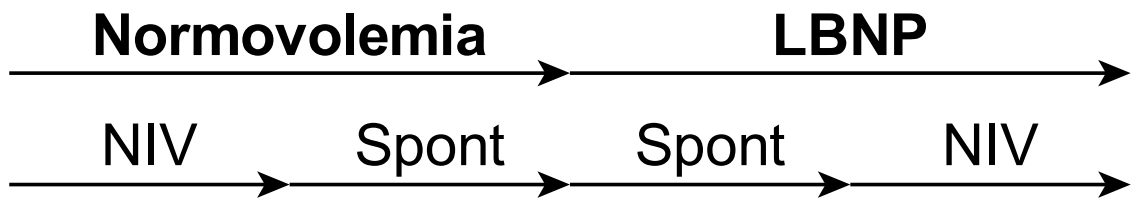
619 **Fig. 3** Oscillations in HR, ICABV, ICABF and MAP during 30 sec (6-8 respiratory cycles) in
 620 four states (a. Baseline: spontaneous breathing+normovolemia, b. NIV: control-mode
 621 NIV+normovolemia, c. Hypovolemia: spontaneous breathing+hypovolemia, and d. Control-
 622 mode NIV+hypovolemia), in the same subject as Fig. 2. Top graph shows respiration (upward
 623 stroke: inspiration, downward stroke: expiration). Inspiration (dotted lines) coincided with
 624 increases in HR and decreases in ICABV. Expiration coincided with decreases in HR and
 625 increases in ICABV. HR: heart rate, ICABV: internal carotid artery beat volume, ICABF:
 626 internal carotid artery blood flow, MAP: Mean arterial pressure, NIV: controlled non-invasive
 627 ventilation

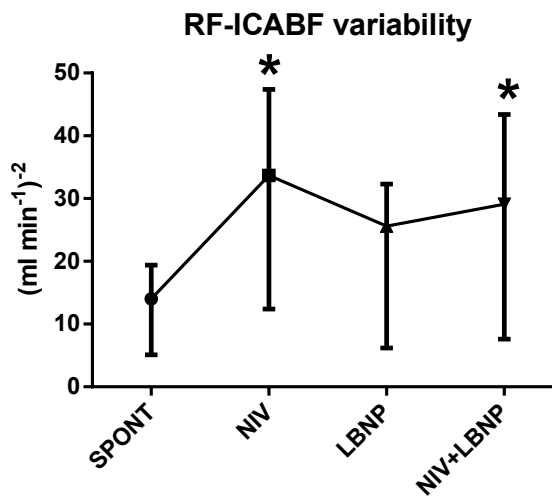
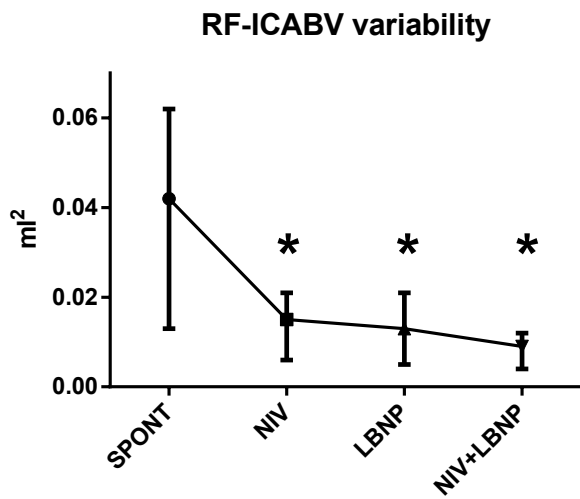
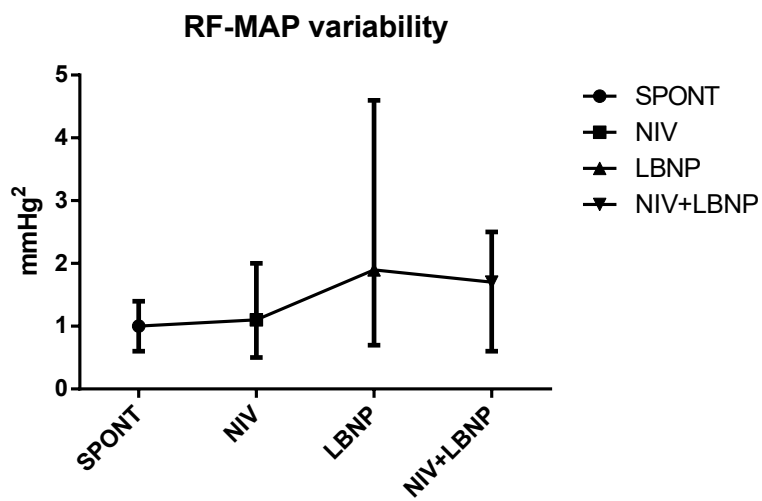
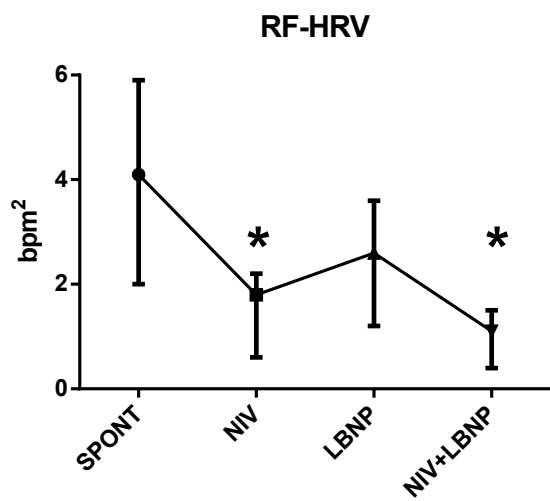
628 **Fig. 4** Respiratory variability (integrals of power spectra) changes in heart rate (HR), mean
 629 arterial pressure (MAP), internal carotid artery beat volume (ICABV) and internal carotid
 630 artery blood flow (ICABF) at peak respiratory frequency in four states (a. Baseline:
 631 spontaneous breathing+normovolemia, b. NIV: control-mode NIV+normovolemia, c.
 632 Hypovolemia: spontaneous breathing+hypovolemia, and d. Control-mode NIV+hypovolemia).
 633 Variability was normalized with respect to the median value during Baseline. Data from 10
 634 healthy humans 20–30 years old. Values are medians and 95% confidence intervals calculated
 635 by Hodges-Lehmann estimates. *Significant difference from spontaneous
 636 breathing+normovolemia (Wilcoxon matched-pairs signed-rank test)

637 **Fig. 5** Power spectra (HF-interval: 0.15-0.4Hz) of cardiovascular variables during four states
638 (a. Baseline: spontaneous breathing+normovolemia, b. NIV: control-mode
639 NIV+normovolemia, c. Hypovolemia: spontaneous breathing+hypovolemia, and d. control-
640 mode NIV+hypovolemia), in the same subject as Fig. 2 and Fig. 3. The RF-interval (0.24–0.3
641 Hz) is noted. HR: heart rate, ICABV: internal carotid artery beat volume, ICABF: internal
642 carotid artery blood flow, MAP: Mean arterial pressure, RF-SPONT: spontaneous respiratory
643 frequency, RF-NIV: set respiratory frequency during controlled non-invasive ventilation

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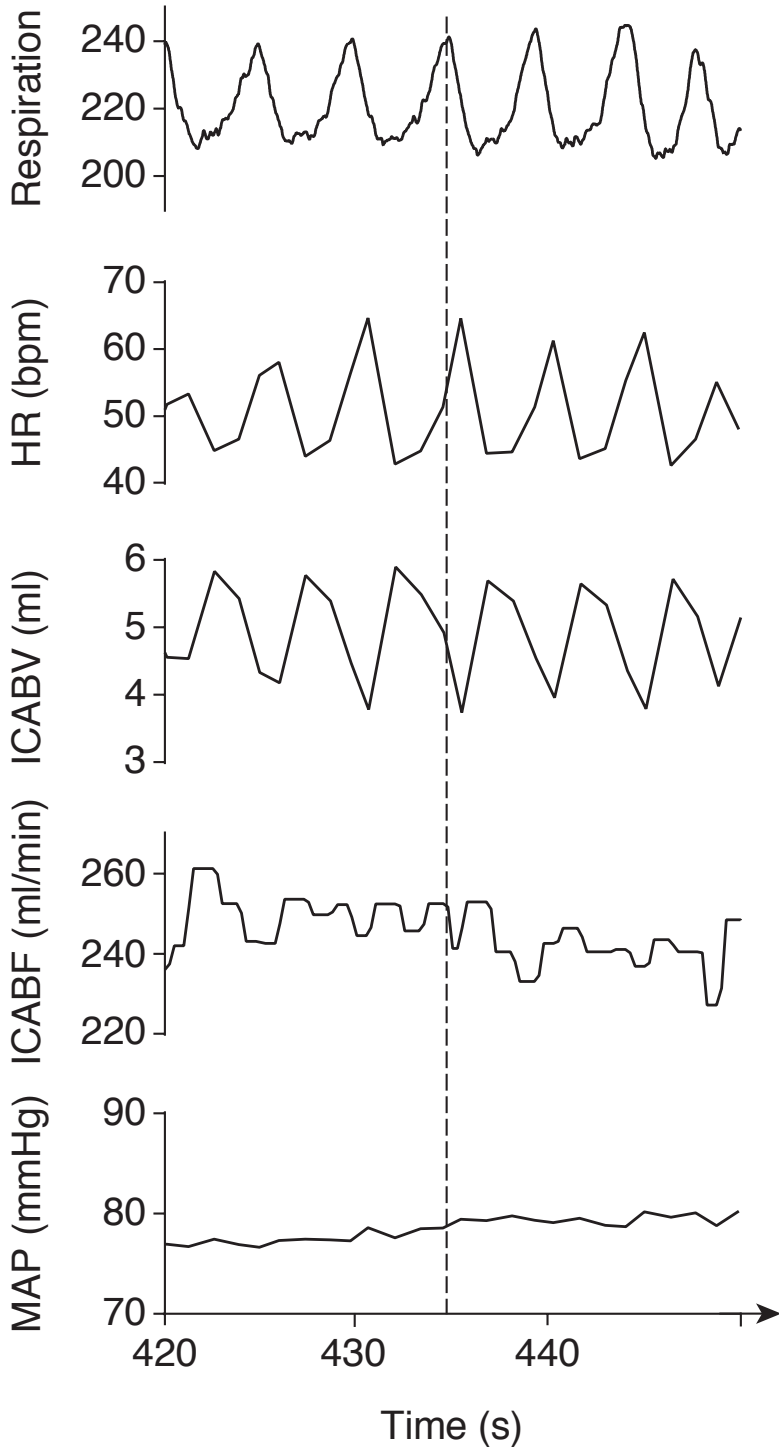
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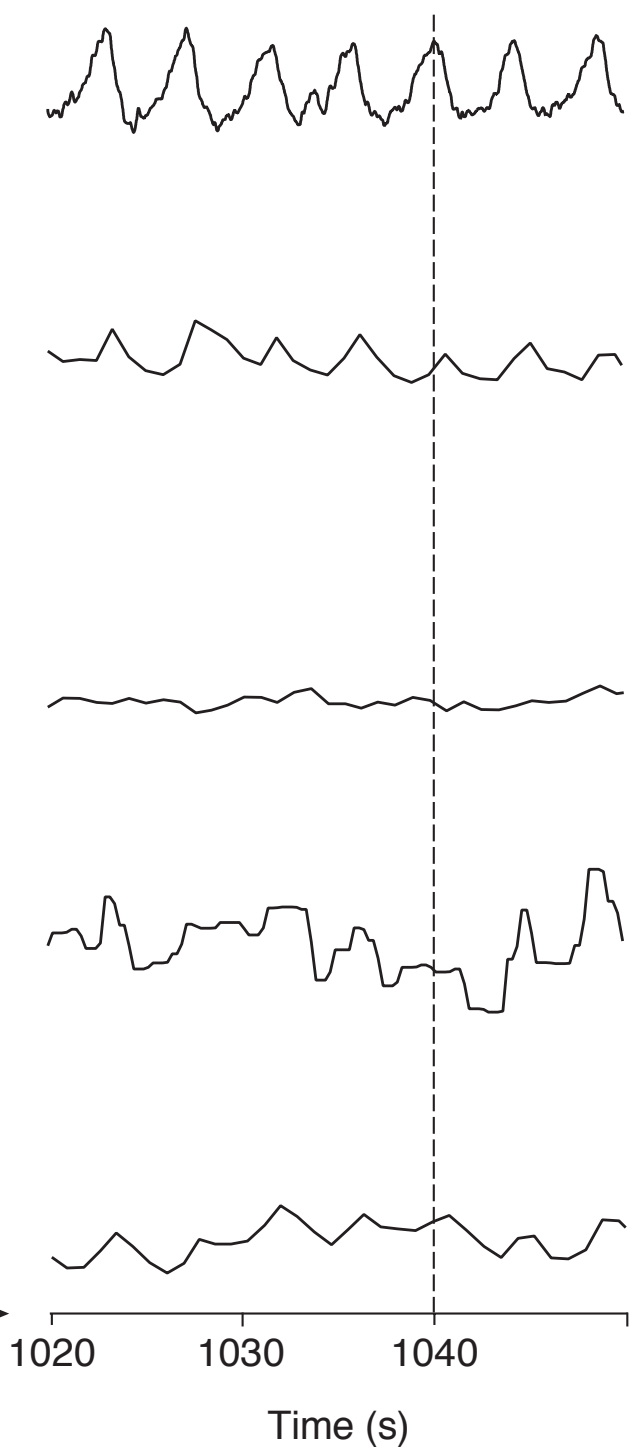
Normovolemia

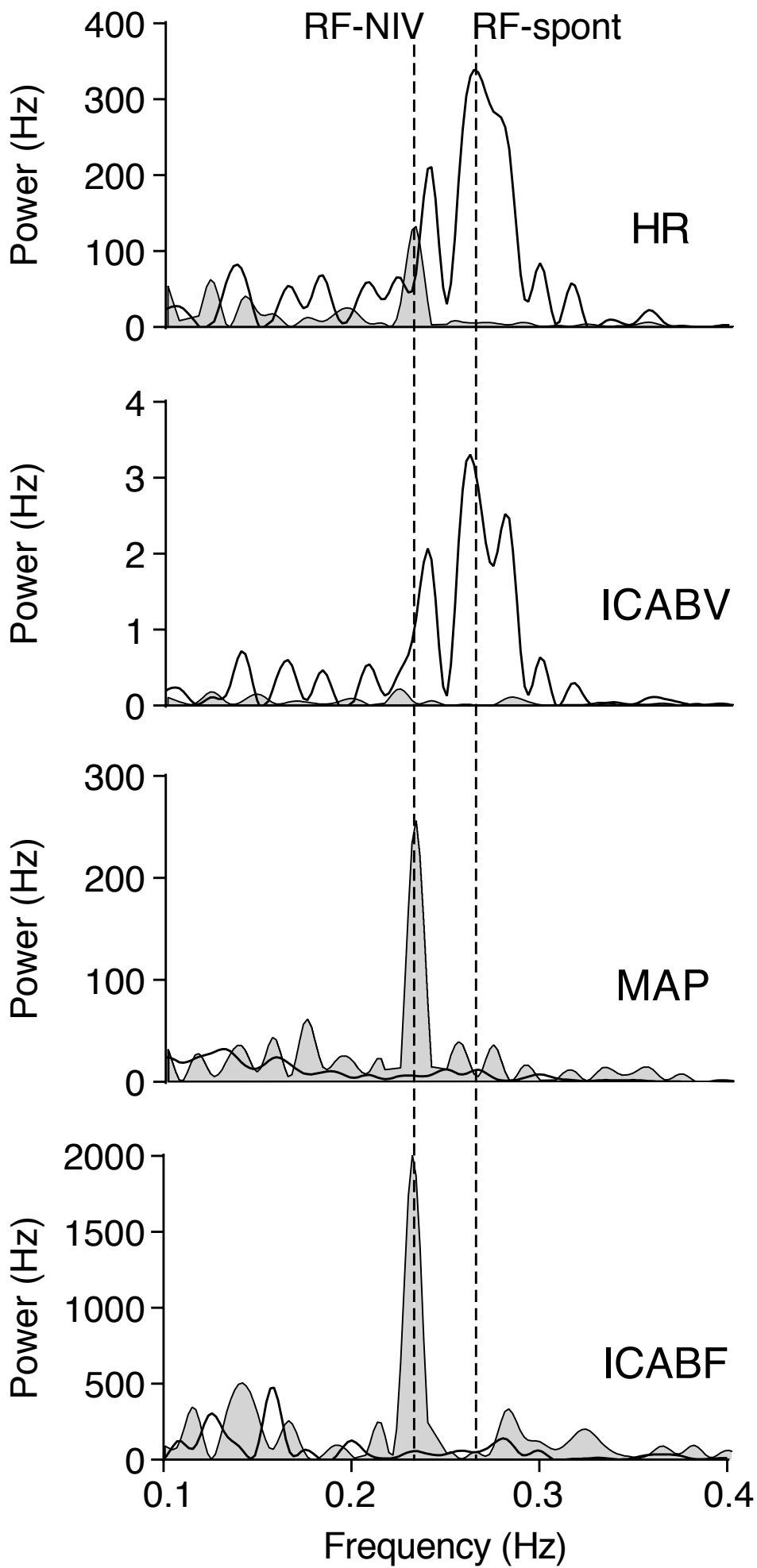
Spontaneous breathing



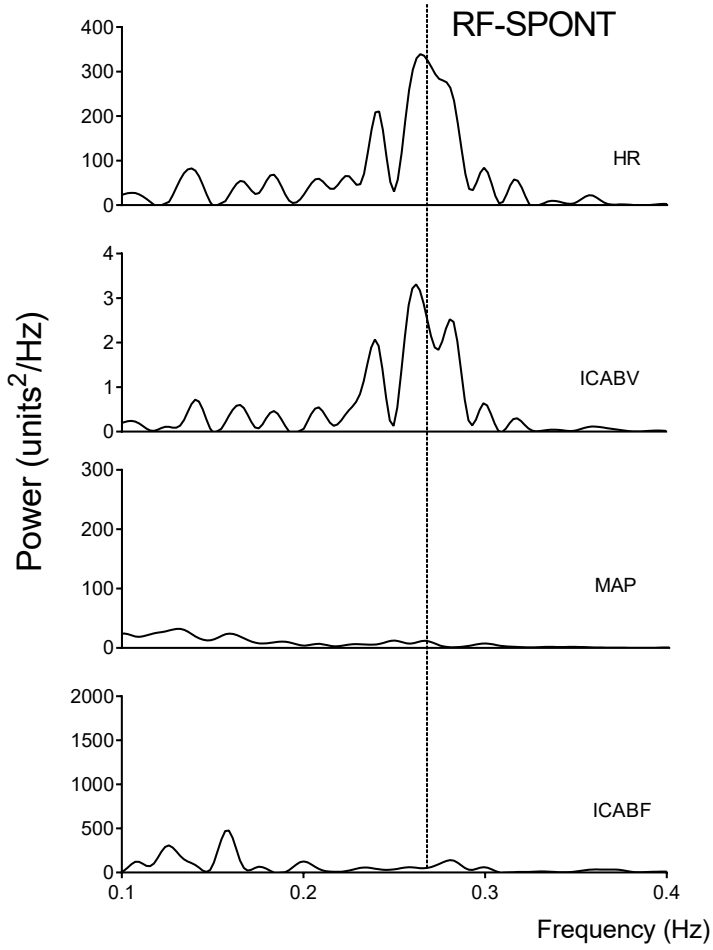
Hypovolemia

NIV

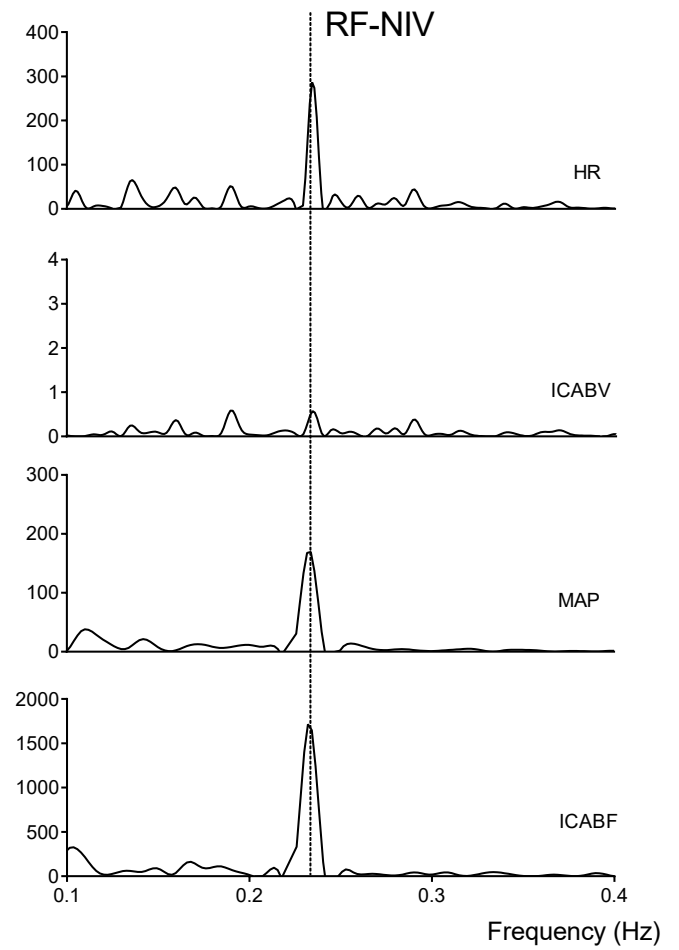




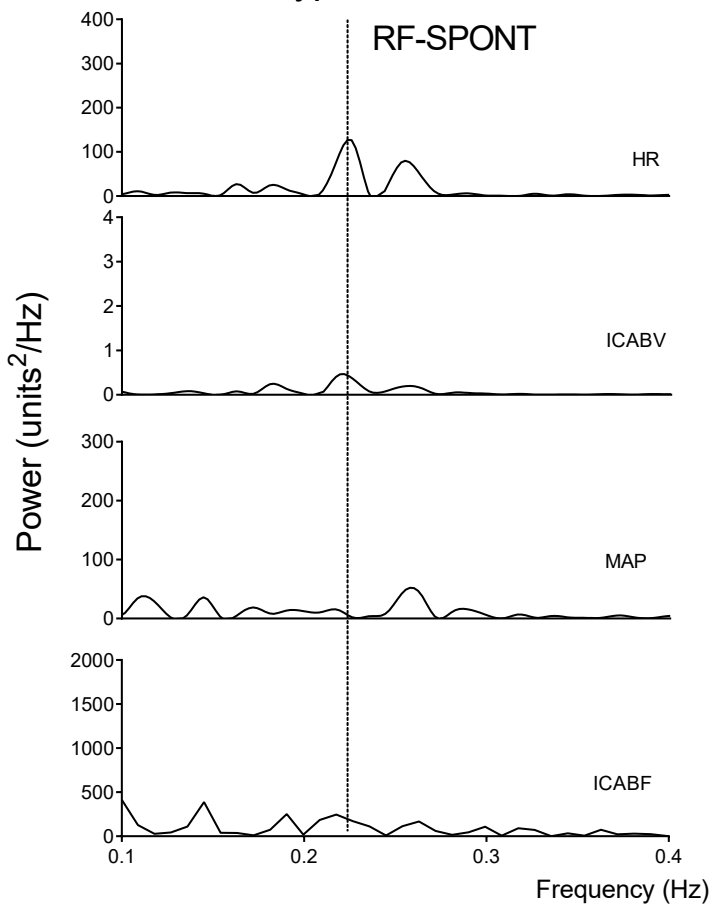
a. Baseline



b. NIV



c. Hypovolemia



d. NIV+hypovolemia

