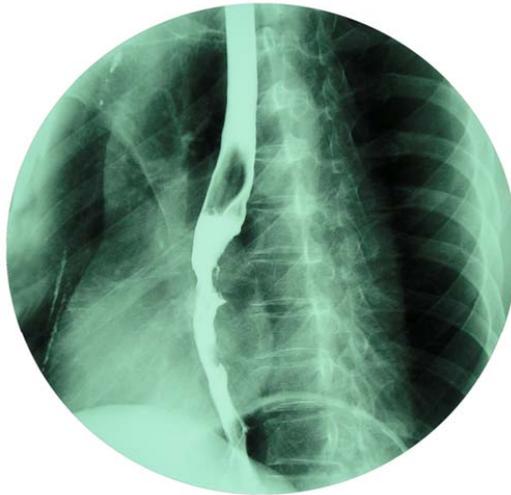


Clinicopathological significance of seprase, dipeptidyl peptidase IV and urokinase-type plasminogen activator in esophageal carcinomas



Mariusz Adam Goscinski

Thesis

**Faculty Division the Norwegian Radium Hospital, Rikshospitalet HF,
Faculty of Medicine**



**UNIVERSITY
OF OSLO**



**Department of Surgical Oncology
and Department of Pathology
The Norwegian Radium Hospital,
Rikshospitalet HF, Oslo, Norway**



© **Mariusz Adam Goscinski, 2009**

*Series of dissertations submitted to the
Faculty of Medicine, University of Oslo
No. 750*

ISBN 978-82-8072-312-3

All rights reserved. No part of this publication may be reproduced or transmitted, in any form or by any means, without permission.

Cover: Inger Sandved Anfinsen.
Printed in Norway: AiT e-dit AS, Oslo, 2009.

Produced in co-operation with Unipub AS.
The thesis is produced by Unipub AS merely in connection with the thesis defence. Kindly direct all inquiries regarding the thesis to the copyright holder or the unit which grants the doctorate.

*Unipub AS is owned by
The University Foundation for Student Life (SiO)*

*To
Alexandra, Zofia, Anna, Jan and Stanislaw*

TABLE OF CONTENTS

1. PREFACE.....	7
<i>Acknowledgements</i>	7
<i>Abbreviations</i>	8
<i>TNM Classification / Staging</i>	10
2. LIST OF PAPERS	13
3. GENERAL INTRODUCTION	15
<i>Epidemiology of esophageal cancer</i>	16
<i>Squamous cell carcinoma</i>	17
Microscopic features	17
Risk factors	18
<i>Adenocarcinoma</i>	18
Microscopic features	18
Risk factors	18
<i>Barrett's esophagus</i>	19
Dysplasia in Barrett's esophagus	20
Macroscopic appearance of Barrett's esophagus	20
Early esophageal cancer.....	23
Esophageal cancer diagnostics	23
<i>Esophageal cancer research areas</i>	24
<i>Treatment of esophageal carcinoma</i>	26
Single modality treatments.....	26
<i>Surgery</i>	26
<i>Endoscopic Mucosal Resection</i>	26
<i>Laser therapy</i>	27
<i>Photodynamic therapy</i>	27
Multimodal treatment.....	28
<i>Radio- and chemotherapy</i>	28
Palliative treatment	29
<i>Tumor invasion and development of metastases</i>	29
<i>Membrane proteases</i>	30
Seprase / FAP- α	31
Dipeptidyl peptidase IV (DPPIV, CD 26).....	32
uPA	32
Serine protease complexes	33
4. AIMS OF THE STUDY.....	35
5. MATERIALS AND METHODS	37
<i>Patients and tissue samples</i>	37
Cell lines	38
Antibodies	38
Laboratory methods	38
<i>Immunohistochemistry</i>	38
<i>Semi-quantitative analysis of immunohistochemistry</i>	39
<i>Immunoblotting</i>	39
<i>Quantitative real-time RT-PCR analysis</i>	39

	<i>Enzymatic activity biochemical assay</i>	40
	<i>Statistical analysis</i>	40
6.	SUMMARY OF PAPERS	41
	PAPER I	41
	PAPER II	42
	PAPER III	43
	PAPER IV	44
7.	GENERAL DISCUSSION	45
	<i>Main findings</i>	46
	<i>Serine proteases in dysplastic-, cancer- and stromal cells of SCC and AC</i>	47
	<i>Clinical study interpretation</i>	50
8.	CONCLUDING REMARKS AND PERSPECTIVES	53
9.	REFERENCE LIST	55
10.	PAPERS I – IV	75

1. PREFACE

Acknowledgements

The work presented in this thesis would not have been possible without the fellowship from the Surgical Oncology and Pathology Departments of the Norwegian Radium Hospital, Rikshospitalet HF, Faculty of Medicine, University of Oslo and the Norwegian Cancer Society.

I would like to thank the head of the Surgical Oncology Department, Prof. Dr. Med. Karl-Erik Giercksky, who inspired me to start these doctoral studies and guided me through good and hard times during the last few years, in addition to providing financial support. Kalle, your trust in my person and competence helped not only immensely in my professional life, but were also of great value in my personal life.

I also wish to thank, Prof. Dr. Med. Jahn Marthin Nesland, Head of the Department of Pathology, for his never failing enthusiasm for this project and for sharing his valuable knowledge of esophageal pathology, as well as funding the necessary research material.

I am also grateful to Dr. Med. Zhenhe Suo from the same department for his contribution with methods, antibodies and Chinese tissue material and for arranging a memorable professional visit to China for me. All three were excellent supervisors.

My gratitude goes also to my colleagues and co-authors at the Departments of Pathology, Surgery, Biochemistry and Statistics of the Norwegian Radium Hospital, Rikshospitalet HF, who contributed in various helpful ways to this thesis.

I have highly appreciated the help from the technical assistants, both at the Department of Pathology of the Radium Hospital, Rikshospitalet HF, and the Department of Pathology of Anyang Tumor Hospital, Henan Province, China.

Special thanks to Inger-Liv Nordli, Mai Nguyen, Ellen Hellesylt, Martina Skrede, Elisabeth Emilsen and Hiep Phuc Dong for their excellent laboratory work.

I am particularly thankful to Dr. Stein Gunnar Larsen, Dr. Malgorzata Zakrzewska, Dr. Monika Slominska-Wojewodzka, Dr. Antoni Wiedlocha, Dr. Vivi Ann Flørenes, Prof. Dr. Med. Wen-Tien Chen, Dr. Trond Warloe, and Prof. Dr. Med. Kjell Elgjo for their constructive criticism and our discussions which widened my scientific horizon.

Special thanks are also dedicated to José-Maria Bassols (Chema) who always readily assisted me with his technical expertise.

Finally, I would like to express my endless gratitude to my beloved wife Alexandra for her invaluable help, understanding, patience and love, in addition to the linguistic revision of the texts.

Oslo, October 2008



Mariusz Adam Goscinski

Abbreviations

ACS	American Cancer Society
ADAM	Disintegrin And Metalloprotease Family
AJCC	American Joint Committee of Cancer
Bcl-2	B-cell CLL/lymphoma 2
Bcl-X	Anti-apoptotic Gene
b-FGF	Basic Fibroblast Growth Factor
BMI	Body Mass Index
BMP-6	Bone Morphogenetic Protein 6
c-Myc	Proto-oncogene Retrovirus-associated DNA Sequences
Cox-2	Cyclooxygenase 2
DAB-2	Disabled Homolog 2, Mitogen-responsive phosphoprotein
DPPIV	Dipeptidyl Peptidase IV
ECM	Extracellular Matrix
EDTA	Ethylenediaminetetraacetic Acid
EGFR	Epidermal Growth Factor Receptor
EMR	Endoscopic Mucosal Resection
ERCC3	Excision Repair Cross-complementing Rodent Repair Deficiency
FAP- α	Fibroblast Activation Protein- α
FAS-L	Fas Ligand (TNF superfamily, member 6)
GERD	Gastroesophageal Reflux Disease
GUS	Beta-Glucuronidase
HER2	Human Epidermal Growth Factor Receptor 2
HET1A	Immortalized Esophageal Keratinocytes (Cell Line)
HPV	Human Papillomavirus
hTERT	Human Telomerase Reverse Transcriptase
HUVEC	Human Umbilical Vein Endothelial Cells
iNOS	Inductible Nitric Oxide Synthase
INT-2	Fibroblast Growth Factor 3
KYSE	Human Japanese Esophagus Carcinoma Squamous Cell (Cell Line)
MMP	Matrix Metalloproteinase
MT-MMP	Membrane-type Matrix Metalloproteinase
Nd:YAG	Neodymium-doped Yttrium Aluminium Garnet
NF κ B	Nuclear Factor Kappa B
NSAID	Non-steroidal Anti-inflammatory Drug
P16	Tumor Suppressor Gene
p16MST1	Tumor Suppressor Gene
PCR	Polymerase Chain Reaction
PDT	Photodynamic Therapy
PVDF	Polyvinylidene Fluoride
QRT-PCR	Quantitative Real-time Polymerase Chain Reaction
RAR β	Retinoic Acid Receptor Beta
RFA	Radiofrequency Ablation
RT-PCR	Real-time Polymerase Chain Reaction
SCC	Squamous Cell Carcinoma

SIM	Specialized Intestinal Metaplasia
SIMP	Serine-type Integral Membrane Peptidase Family
TBS	Tris Buffered Saline
TGF- α	Transforming Growth Factor Alpha
TGF- β	Transforming Growth Factor Beta
TGH	Triiodothyronine, Glucagon and Heparin Mixture
TNM	Classification of Malignant Tumors
TP	Thymidine Phosphorylase Gene
TRAIL	Tumor Necrosis Factor-related Apoptosis-inducing Ligand
TRIS	Tris(hydroxymethyl)aminomethane
UICC	International Union Against Cancer
uPA	Urokinase-type Plasminogen Activator
uPAR	Urokinase-type Plasminogen Activator Receptor
VEGF	Vascular Endothelial Growth Factor
WHO	World Health Organization

TNM Classification / Staging

Histological classification of carcinomas according to UICC Global Cancer Control, TNM Atlas (UICC 2005).

Anatomical subsites of the esophagus:

- Cervical esophagus
- Intrathoracic esophagus
 1. Upper thoracic portion
 2. Mid-thoracic portion
 3. Lower thoracic portion

T – Primary Tumor

- Tx Primary tumor cannot be assessed
- T0 No evidence of primary tumor
- Tis Carcinoma *in situ*
- T1 Tumor invades lamina propria or submucosa
- T2 Tumor invades muscularis propria
- T3 Tumor invades adventitia
- T4 Tumor invades adjacent structures

N – Regional Lymph Nodes

- Nx Regional lymph nodes cannot be assessed
- N0 No regional lymph nodes metastasis
- N1 Regional lymph node metastasis

Regional lymph nodes:

- Cervical esophagus:
 - Scalene
 - Internal jugular
 - Upper and lower cervical
 - Periesophageal
 - Supraclavicular
- Intrathoracic esophagus:
 - Upper periesophageal
 - Subcarinal
 - Lower periesophageal
 - Mediastinal lymph nodes
 - Perigastric lymph nodes

M – Distant Metastasis

Mx	Distant metastasis cannot be assessed
M0	No distant metastasis
M1	Distant metastasis

Tumors of the upper thoracic esophagus:

M1a	Metastasis in cervical lymph nodes
M1b	Other distant metastasis

Tumors of the mid-thoracic esophagus:

M1a	Not applicable
M1b	Non-regional lymph nodes or other distant metastasis

Tumors of the lower thoracic esophagus:

M1a	Metastasis in celiac lymph nodes
M1b	Other distant metastasis

pTNM – Pathological Classification

The pT, pN and pM categories correspond to the TNM categories (UICC 2005).

AJCC/UICC Stage Grouping for Esophageal Cancer (AJCC 2002, UICC 2005)

Cancer		
Stage 0	-	Tis N0 M0
Stage I	-	T1 N0 M0
Stage IIA	-	T2 N0 M0
		T3 N0 M0
Stage IIB	-	T1 N1 M0
		T2 N1 M0
Stage III	-	T3 N1 M0
		T4 Any N M0
Stage IV	-	Any T Any N M1
Stage IVA	-	Any T Any N M1a
Stage IVB	-	Any T Any N M1b

2. LIST OF PAPERS

Paper I

Goscinski MA, Suo ZH, Nesland JM, Flørenes VA, Giercksky KE.

Dipeptidyl peptidase IV expression in cancer and stromal cells of human esophageal squamous cell carcinomas, adenocarcinomas and squamous cell carcinoma cell lines. APMIS 2008;116:823-31.

Paper II

Goscinski MA, Suo ZH, Nesland JM, Chen WT, Zakrzewska M, Wang J, Zhang S, Flørenes VA, Giercksky KE.

Seprase, DPPIV and uPA expression in dysplasia and invasive squamous cell carcinoma of the esophagus. A study of 229 cases from Anyang Tumor Hospital, Henan Province, China. Oncology 2008;75:49-59.

Paper III

Goscinski MA, Suo ZH, Flørenes VA, Vlatkovic L, Nesland JM, Giercksky KE.

Fap- α and uPA show different expression patterns in premalignant and malignant esophageal lesions. Ultrastruct Pathol 2008;32:89-96.

Paper IV

Goscinski MA, Larsen SG, Warloe T, Stoldt S, Nesland JM, Suo ZH, Giercksky KE.

Adenocarcinomas on the rise – does it influence survival from oesophageal cancer? Scand J Surg 2008, submitted.

3. GENERAL INTRODUCTION

Once the diagnosis of cancer is established, it is of utmost importance to determine whether the disease is local or already has spread to lymph nodes or distant organs (Fidler & Ellis 1994). Some scientists suggest that in nearly 50% of patients, surgical excision of primary malignant tumors is not curative because metastases have already occurred by that time (Sugarbaker et al. 1977, Sugarbaker 1979, Fidler & Balch 1987). Currently, metastases are the cause of 90% of human cancer mortality (Mehlen et al. 2006). Recent studies and clinical experiences show that metastases without any doubt are the most dreaded aspect of cancer.

In the 20th century, many different hypotheses about metastases were put forward. One of the first approaches was Stephen Paget's "seed and soil" hypothesis (Paget 1889), describing organs that are affected by disseminated cancer. Later, scientists such as James Ewing, Dale Rex Coman, Irving Zeidman, Barbara Lucke and others, suggested different theories about the origin of metastases (Fidler 2003). Isaiah J. Fidler reported in 1970-1973 that metastasis can result from the survival of only a few tumor cells (Fidler 1970, Weiss 1986, Fidler 1990). After the year 2000, research concerning tumor progression and metastasis development confirmed the theory that at the time of diagnosis, many human tumors are heterogeneous and include numerous cell subpopulations. The tumors also contain the so-called "cancer stem cells", which display different biological characteristics and metastatic potential (Bonnet & Dick 1997, Fidler 2003).

Today, research on malignant expansion is in progress and mainly concentrates on early cancer diagnosis and cancer therapy directed against host factors – angiogenesis and organ growth factors, as it is believed to provide a basis for treatment that will give better results than conventional therapy (Simone et al. 1998, Fidler 2003).

As mentioned above, there are many hypotheses and approaches trying to shed light upon the process of neoplastic disease development. Some of the theories focus particularly on the formation of metastases from the primary tumor through an exceedingly complex process. It includes a series of sequential steps such as invasion of adjacent tissue, intravasation, transport through the circulatory system and arrest at a secondary site, extravasation and growth in a secondary organ (Folkman 1986, Liotta 1986, Nicolson 1988). To complete this process, degradation of the extracellular matrix (ECM) is decisive (Folkman 1986, Takino 2007). Basic and clinical research has therefore been concentrated on the role of tumor-associated proteolytic systems – proteases and anti-proteases, particularly membrane proteases. Some of the most extensively studied proteases are dipeptidyl peptidase IV (DPPIV), urokinase-type plasminogen activator (uPA) and, recently, seprase (surface expressed protease, also known as fibroblast activation protease α (FAP- α)) (Vivier et al. 1991, Morimoto et al. 1994, Hansen et al. 1994, Rettig et al. 1994, Scalan et al. 1994, Mathew et al. 1995, Chen 2003, Chen & Kelly 2003).

Many studies have shown that in several tumors the expression of membrane proteases in cancer cells or stromal cells adjacent to cancer sites correlates with an

increasing tumorigenicity (Nishino et al. 1988, Scalani et al. 1994, Nekarda et al. 1998, Okada et al. 2003, Iwasa et al. 2003, Kikkawa et al. 2005). However, the pathophysiological significance of the serine proteases in esophageal carcinoma has not yet been fully elucidated.

Epidemiology of esophageal cancer

Esophageal cancer is turning into one of the more common cancers in the world. The incidence shows great geographical variations, but on the whole it is stated to be number six in frequency (Stewart & Kleihues 2003, Siewert et al. 2004, NCI 2006). The majority of cases are diagnosed in developing countries, where it is the fourth most frequently occurring cancer. The area with the highest reported incidence is known as the “Esophageal cancer belt of South-Central Asia”, which extends from Turkey through northern Iran, Afghanistan and southern Russia to northern China (Ghadirian 1982, Saidi 1999, Hajian 2002). The invasive cancer incidence rate in these areas is more than 200 per 100 000 inhabitants, compared to Europe and the USA where it is 2.6-11.1 cases and 2.6-5.9 cases per 100 000 inhabitants respectively, with considerable racial and regional differences (NCI 2006). In particular, the area located in the southern part of the Taihang Mountains on the borders of Henan, Shansi and Hopei provinces in China has one of the highest incidence and mortality rate for esophageal carcinoma in the world. In Linxian county in Henan province, the age-adjusted mortality rate for esophageal carcinoma has been reported to be 151/100 000 for males and 115/100 000 for females (Lu et al. 1988, Stoner et al. 2001, He et al. 2008).

There are two different histological types of esophageal carcinoma: squamous cell carcinoma (SCC) and adenocarcinoma (AC). In Western countries, the number of SCCs and ACs is almost equivalent, while in developing countries, SCC is the most prevalent one (Pera et al. 1993, Devesa et al. 1998, Blot & McLaughlin 1999, Daly et al. 2000, Corley et al. 2001, Parkin et al. 2002, Pera et al. 2005, de Jonge et al. 2006, Yee et al. 2007, Holmes et al. 2007).

Since the entire esophageal tract is normally lined with squamous epithelium, SCC can occur anywhere along the length of the esophagus. AC, on the other hand, starts in tissue lined by columnar epithelium, which normally does not cover the esophagus. In order for an AC to develop, columnar epithelium must replace an area of squamous epithelium in a metaplastic process (Barrett’s metaplasia), often followed by dysplasia (non-invasive neoplastic transformation with a potential for malignant progression). This occurs mainly in the lower esophagus, where most ACs are found (Stein et al. 1993, Cote et al. 2003, de Jonge et al. 2006).

Squamous cell carcinoma

The esophagus is normally lined with stratified squamous nonkeratinizing epithelium (Fig.1A) (Norton et al. 2000). SCC arises from this epithelial layer, probably in response to chronic toxic irritation. In many cases, esophageal SCC is the end result of a progression through increasingly severe degrees of dysplasia to carcinoma *in situ*, resulting in invasive carcinoma (Fig. 1B). Clinical and epidemiological studies have confirmed that squamous epithelial dysplasia of the esophagus is a precancerous lesion and that approximately 70% of patients with squamous dysplasia later develop SCC (Mukada et al. 1978, Munoz et al. 1982, Mandard et al. 1984, Kuwano et al. 1988).

Epithelial dysplasia has previously been classified into low-, moderate- and high grades (Oehlert et al. 1979, Riddell et al. 1983, Tosi et al. 1989, de Dombal et al. 1990, WHO 1990, Lewin & Appelman 1996). Low grade dysplasia usually affects less than half of the epithelial thickness, while high grade dysplasia affects more than half of it. Moderate grade dysplasia is borderline or intermediate between the two other grades. Sometimes, separating one grade from another is difficult, as the lines of demarcation are not always clear. Nowadays, many chose to use thus the two-grade system, low- and high grade dysplasia (Riddell et al. 1983, Tosi et al. 1987 & 1989, Rubio et al. 1989, de Dombal et al. 1990, Burke et al. 1991).

When it comes to the transition from high grade dysplasia to carcinoma *in situ*, no clear distinction can be made either macro- or microscopically. Both stages display a similar histological pattern and probably require similar surveillance and treatment (Lewin et al. 1998).

Microscopic features

Preinvasive lesions are characterized by cellular atypia and abnormal intraepithelial architecture. These features progress until atypical cells have spread to all epithelial layers and the architecture no longer can be identified. These characteristics are clearly visible in high grade dysplasia (carcinoma *in situ*), whereas low grade dysplasia still has a component of differentiated squamous cells in the upper layers, and abnormal cells are limited to the lower half of the epithelium (Crissman et al. 1987 & 1989, Mills et al. 2000, WHO 2000).

The histologic appearance of invasive SCC is related to invasion of atypical cells into lamina propria or beyond it (Schlemper et al. 1997). Usually, a large pushing front of numerous cohesive squamous cells can be observed, but the cells may also invade as small cell nests or as individual cells infiltrating through the stroma. Invasive carcinomas generate a surrounding inflammation, where a mixture of various leukocytes can be observed (Fig. 1B and 1C) (Crissman et al. 1987 & 1989, WHO 2000).

Risk factors

Use of tobacco and alcohol are among the highest risk factors regarding SCC. Consumption of salty and spicy food, as well as low fiber intake have also been positively identified as risk factors for developing SCC (Soler et al. 2001, Enzinger & Mayer 2003, Syrjänen 2002, Green et al. 2003). Further risk factors are summarized in Table 1.

Adenocarcinoma

In contrast to SCC, the incidence of AC nowadays is rising rapidly to the point where this tumor accounts for 50% or more of all esophageal cancers in the USA and other industrialized countries (Lagergren et al. 1999, Bollschweiler 2001, de Jonge et al. 2006, Merry et al. 2007, Hashibe et al. 2007). ACs occur almost exclusively in the distal part of the esophagus. The tumor is first seen as a thickened plaque-like white mucosa. Larger lesions form white exophytic polypoid masses with well demarcated borders. Occasionally, carcinomas appear papillary or may be multifocal. AC spreads through the esophagus into periesophageal tissues (Fenoglio-Preiser et al. 1989).

Microscopic features

The majority of ACs is classified as low-, moderate to well differentiated carcinomas and form glands with columnar epithelium. The epithelial cells contain nuclei with a coarse chromatin pattern, nucleoli and cytoplasm in which mucin can be found (Fig. 1F). A minority of tumors displays the diffuse type pattern with signet ring cells (Pettersen 1932, Adams et al. 1945, Spin 1973, Thompson et al. 1983, Thurberg et al. 1999, Nakagawa et al. 2000).

Risk factors

The major acknowledged risk factor for AC is the pathological and clinical alteration of the lower esophagus, also known as Barrett's esophagus, coexisting with gastroesophageal reflux disease (GERD), wherein columnar epithelium replaces the squamous epithelium that normally lines the distal esophagus (Lagergren et al. 1999, Heath et al. 2000, Ye et al. 2004, Nakajima & Hattori 2004, Oberg et al. 2005, Layke, et al. 2006). Other risk factors are summarized in Table 1.

Table 1. Risk factors for esophageal carcinoma.

SCC	AC
Tobacco smoking*	GERD
Alcohol consumption	Barrett's esophagus
Consumption of salt-cured, salt-pickled and moldy food**	High BMI
Low fiber intake	Tobacco smoking*
Consumption of hot beverages	Use of NSAID
Achalasia	Work in stooped posture
Previous head and neck cancer	Genetic alterations
Plummer-Vinson syndrome	
HPV (human papilloma virus) infection	
Genetic alterations	

* Nitrosamines, polycyclic aromatic hydrocarbons, aromatic amines, aldehydes and phenols

** Nitrosamines and fungal toxins

Barrett's esophagus

The importance of Barrett's esophagus rests on the fact that local occurrence of columnar metaplasia can develop into AC. The concept of Barrett's esophagus has changed significantly since its first description by Numan R. Barrett in 1950 and 1957 (Barrett 1950 & 1957). In 1950, in "Chronic Peptic Ulcer of the Oesophagus and Oesophagitis", Barrett reviewed several published cases, looked at preserved pathologic specimens and found that esophageal ulcers were surrounded by columnar mucosa. He concluded that "these cases were examples of congenital short esophagus – in which part of the stomach extends upwards into the mediastinum, or even to the neck, and that in this stomach a typical chronic gastric ulcer can form". He suggested that the ulcer was a separate entity from reflux esophagitis (Barrett 1950, Cameron 2001). In 1957, Barrett revised his theory and noted that the columnar lining was a continuous sheet extending from the esophago-gastric junction upwards, and that it was thus columnar cell-lined esophagus extending into the mediastinum and not the stomach (Barrett 1957). As a result of these observations, the term "Barrett's esophagus" has become well established in medical literature to indicate columnar metaplasia of the distal esophagus associated with chronic GERD. Ever since, several different concepts of Barrett's esophagus have emerged (Cohen et al. 1963, Trier 1970, Bremner et al. 1970, Naef & Savary 1972, Iascone et al. 1983, Gillen et al. 1988).

Based on the potential development of malignancy, the heterogenous spectrum of Barrett's metaplastic mucosa is classified into two distinct types: columnar epithelium, with specialized intestinal metaplasia (SIM), including goblet and columnar non-goblet cells, and columnar epithelium, without specialized intestinal metaplasia and lacking goblet cell-type elements (Barrett 1957, Siewert & Dittler 1993, Koppert et al. 2005, Fléjou 2005). The goblet cells are presently regarded as the hallmark in the histological identification of Barrett's esophagus and in the selections of high-risk patients for endoscopic surveillance (Chaves et al. 1999). The columnar epithelium with SIM is the most common and distinctive epithelium type found in Barrett's esophagus (Fig. 1D) (Spechler et al. 1994, Spechler 2003).

Dysplasia in Barrett's esophagus

Dysplasia in Barrett's esophagus can be classified as low-, moderate or high grade. In low grade dysplasia, simple tubules with little branching can be demonstrated. Atypical cells are present in the glands, and the epithelium is usually pseudostratified. Mitotic figures usually are sparse but can be present in the superficial half of the mucosa (Lewin 1998, Goldblum 2003).

High grade dysplasia (carcinoma *in situ*) shows severe cellular atypia and varying degrees of abnormal architecture. The nuclei are often enlarged, vesicular with irregularly clumped chromatin and contain large, distinct and irregular nucleoli (Fig. 1D and 1E) (Smith et al. 1984, Schmidt et al. 1985, Spechler & Goyal 1986, Reid et al. 1988). Moderate grade dysplasia is a borderline condition between low- and high grade dysplasia.

Macroscopic appearance of Barrett's esophagus

Endoscopy reveals columnar epithelium in the distal esophagus with a characteristic red color which contrasts sharply with the pale appearance of adjacent squamous epithelium (Spechler & Goyal 1996). Although endoscopic examination can usually distinguish columnar epithelium from squamous epithelium, it is difficult to tell apart different types of columnar epithelium by endoscopic appearance alone. SIM and gastric columnar epithelium can only be distinguished by histological examination of biopsy specimens. Gastric epithelium may normally line a short segment of the distal esophagus. Thus, endoscopists usually diagnose Barrett's esophagus only when columnar epithelium extending well above the gastroesophageal junction can be verified. The criteria for diagnosing Barrett's esophagus based on the length of the columnar lining have varied, ranging from 2-5 cm or more, measured above the gastroesophageal junction. Nowadays, we distinguish only two types of Barrett's esophagus: the long segment, where columnar lining extends more than three cm from the gastroesophageal junction, and the short segment, where columnar lining extends up to three cm from the gastroesophageal junction (Schnell et al. 1992, Levine et al. 1993, Spechler 1994 & 2003, Hirota et al. 1999, Spechler & Goyal 1996).

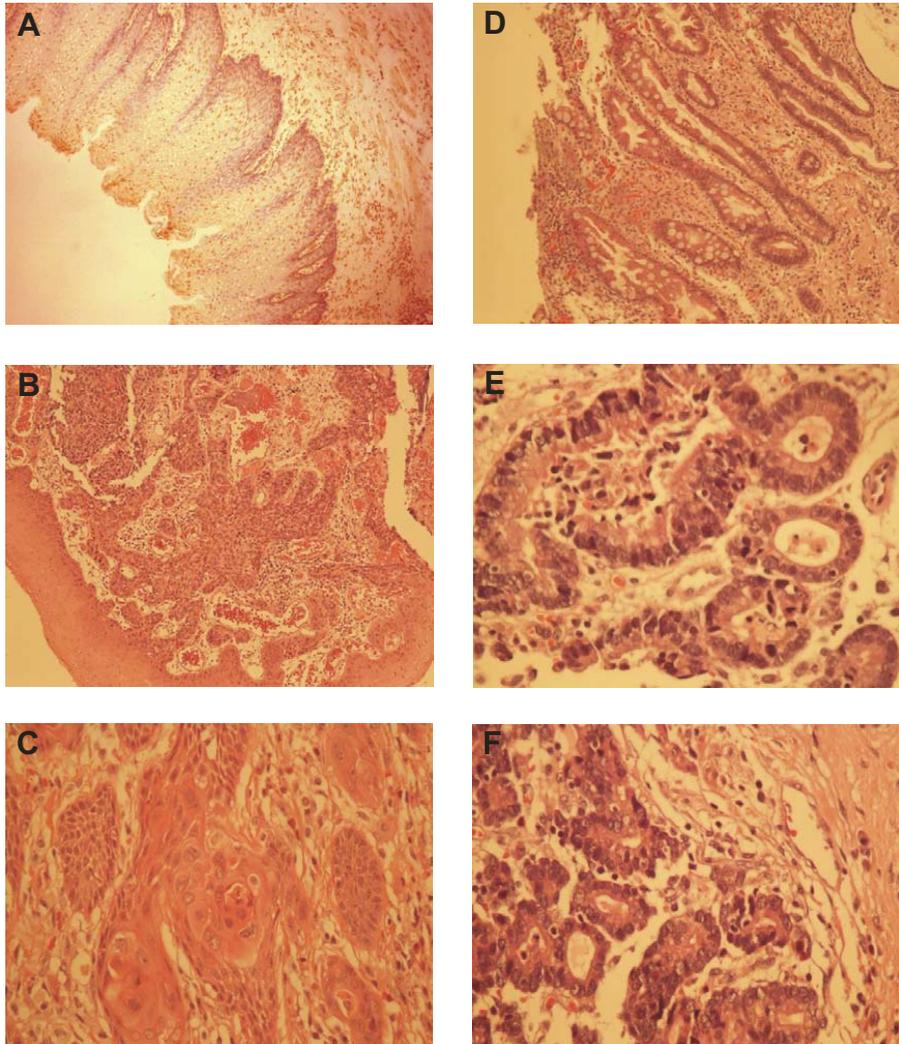


Figure 1. Microscopic illustrations of SCC and AC (original magnification x 200). Normal esophageal epithelium (A). Transition of normal esophageal epithelium toward SCC (B). Well differentiated SCC (C). Barrett's esophagus with intestinal metaplasia within columnar epithelium (D). High grade dysplasia within columnar epithelium (E). Moderately differentiated AC (F).

Early esophageal cancer

Early-stage esophageal cancer is almost asymptomatic with minimal findings in endoscopy. However, a risk for malignant spreading into vessels and lymph nodes is present even in small tumors (Peters et al. 1994, van Sandick et al. 1998, Stein et al. 2000 & 2005, Buttar et al. 2001).

The accepted definition of early esophageal cancer is based on two related elements: superficial extension of the tumor in mucosa and submucosa, and complete or almost complete absence of lymph node metastasis. Most cases correspond therefore to stage I cancer (T1N0M0), according to the TNM/AJCC/UICC staging system (see above) (AJCC 2002, UICC 2005).

This definition does not apply to all superficial SCC tumors (T1). The lymph node involvement is negligible only when the tumor is limited to the mucosa. There are three successive phases in the invasion of mucosa: intraepithelial cancer (carcinoma *in situ*), without affection of the basal membrane; microinvasive cancer with invasion into lamina propria; and intramucosal / transmucosal cancer, invading the muscularis mucosa. No spreading to lymph nodes is associated with intraepithelial cancer, and only a small number of lymph nodes can present metastasis in microinvasive cancer. Intramucosal cancer presents a higher degree of lymph node metastases and reaches the submucosal mucous glands through intraductal propagation (Hölscher et al.1997, Ell et al. 2000, Rice et al. 2001, May et al. 2002, Stein et al. 2003).

Severe dysplasia is considered as carcinoma *in situ* and is often found in patients with multicentric esophageal cancer. Tumors extending into the submucosa have a risk rate for lymphatic invasion varying from 30% to 50%. Submucosal SCC cannot be considered as an early cancer, but must instead be treated as an advanced cancer (Streitz et al. 1991, Nigro et al. 1999, Tajima et al. 2000, Buskens et al. 2004). In AC that has developed in columnar cell-lined esophagus, a clear-cut distinction between superficial (T1) and non-superficial (T2) cancer is based on whether it invades the muscularis propria or not (Lambert R, 1995, Hölscher et al.1997).

Esophageal cancer diagnostics

Direct diagnostic techniques:

- Esophagoscopy with biopsy followed by a pathological evaluation
- Barium X-ray
- Computed tomography with multi-slice technique

Diagnostic techniques used to determine the extent of disease:

- Endoscopic ultrasound
- PET and PET/CT scans
- Bronchoscopy
- Thoracoscopy and laparoscopy

Esophageal cancer research areas

It is well known that several environmental factors can lead to the development of esophageal carcinoma. Environmental carcinogens are able to affect the genetic material of host cells, generating an abnormal regulation of multiple genes (Stoner et al. 2007). Genetic alterations observed in esophageal carcinomas are based on alterations in tumor suppressor genes leading to uncontrolled cell proliferation and terminating apoptosis, loss of cell cycle control as a result of disruption of G1/S cell cycle checkpoints and changes in oncogene functions leading to deregulation of cell signaling pathways (Stoner et al. 2007). Several previous studies have reported numerous genetic alterations associated with esophageal carcinomas, as summarized in Table 2 (Lu et al. 1988, Hollstein et al. 1988 & 1991, Jiang et al. 1993, Guo et al. 1993, Gao et al. 1994, El-Rifai et al. 1998, Moskaluk 1998, Tanaka et al. 1999, Hiyama et al. 1999, Xing et al. 1999, van Dekken 1999, Raida et al. 1999, Zimmermann et al. 1999, Kimura et al. 1999, Lu 2000, Mandard et al. 2000, Selaru 2002, Kuwano et al. 2005).

The discovery of new tumor cell markers, e.g. growth factor receptors, angiogenic and apoptotic factors or transmembrane proteases, has become an important current research topic (Table 3) (Gottlinger et al. 1986, Robaszkiewicz et al. 1991, Reid et al. 1992 & 2000, Traweek et al. 1993, Galipeau et al. 1996, Teodori et al. 1998, Hanahan & Weinberg 2000, Lam 2000, Mandard et al. 2000, Blant et al. 2001, Xu Y et al. 2002, Klein et al. 2002, Yu et al. 2003, Hosch et al. 2003). The expression differences in molecular markers in premalignant and malignant tumor stages, as well as in various histological tumor types are essential for the development of more effective and early diagnostic methods and less harmful therapies for esophageal carcinoma.

In combination with surgery, gene therapy, immunotherapy and new types of chemotherapy, including tyrosine kinase inhibitors or examination of new combinations of existing drugs with irradiation (multimodal therapy), are the current treatment trends (Shaheen & Ransohoff 2002, AJCC 2002, Enzinger & Mayer 2003, Swisher et al. 2003, Burmeister et al. 2005, Kleinberg et al. 2004, Koshy et al. 2004, Posner et al. 2005, Souza & Spechler 2005, ACS 2008).

Table 2. Genetic alterations associated with esophageal carcinoma (modified after Stoner & Gupta 2001).

SCC
<p>p53 mutations Loss of p16MST1 and / or p15, and / or RARβ and disabled-2 (DAB-2) Amplification of INT-2, EGFR, cyclin D₁ and c-Myc Altered expression of the cyclin D1 Altered expression of apoptosis related genes: bcl-2, caspase 3, TRAIL, Fas-L, Fas Elevations in hTERT, BMP-6, iNOS, COX-2 and β-catenin levels Loss of heterozygosity on chromosomes 1p, 3p, 4, 5q, 9, 11q, 13q, 17q and 18 Enhanced expression of the transcription activator, NFκB</p>
AC
<p>Gains of chromosomes 6p, 7pq, 8q, and 17q High-level amplifications at 8q23~q24.1, 15q25, 17q12q21 and 19q13.1 Losses of chromosomes 4pq, 5q, 18q, 19p, 20, 21, and Y</p>

Table 3. Molecular markers in esophageal cancer (modified after Stoner & Gupta 2001).

Markers
<p>Growth factor receptors EGFR HER2/neu</p>
<p>Angiogenetic factors Cox-2 TP VEGF b-FGF TGF-α, TGF-β</p>
<p>Cell cycle regulators Cyclin D1 p21, p27, p53</p>
<p>Apoptotic factors Bcl-2 Bcl-X P16 Survivin</p>
<p>DNA repair system ERCC3</p>
<p>Matrix metalloproteinases MMP-2, MMP-7, MMP-9</p>

Treatment of esophageal carcinoma

Single modality treatments

Surgery

Surgical resection has long been the mainstay of curative treatment and remains the standard treatment for stage T1-T3, N0, M0 tumors. It is contra-indicated in locally advanced tumors (T4) and when lymph node and distant metastases have developed (Fumagalli 1996, Veuillez et al. 2007). Total or partial esophagectomy is most commonly performed on esophageal and gastric cardia carcinomas, as well as in patients with Barrett's esophagus associated with severe dysplasia, undilatable strictures and benign, obstructing tumors (Fumagalli 1996).

Usually, resection of the thoracic esophagus is made by a right- or left-sided transthoracic 'en bloc' resection. A transhiatal resection is an option for cancer of the distal part of the esophagus (Skinner 1983, Orringer 1984, Killinger et al. 1996, Veuillez et al. 2007).

Esophageal cancer involves a risk for lymph node metastasis development, and the quality of the lymphadenectomy associated with esophagectomy is therefore important. Two-field lymphadenectomy (thoracic and abdominal) for cancers from the distal and middle part of the esophagus and three-field lymphadenectomy (thoracic, abdominal and cervical) for cancers of the proximal part of the esophagus are recommended (Fumagalli 1996, Lerut et al. 2004, Veuillez et al. 2007).

The results depend on the extension of the disease, possible co-morbidities and the surgeon's experience. The post-operative mortality rate currently ranges from 4% to 10%, depending on the surgical centers (patient volume and team experience) (Collard et al. 2001, Bumm & Wong 1994). After R0 resection, the reported 5-year overall survival rate is greater than 95% for stage 0, between 50% and 80% for stage I, 30% and 40% for stage IIA, 10% and 30% for stage IIB and between 10% and 15% for stage III (Enzinger & Mayer 2003, NCI 2006, Veuillez et al. 2007).

Endoscopic Mucosal Resection

This method is restricted to superficial cancers limited to the mucosa, submucosa or precancerous lesions and is not recommended for tumors invading the muscularis mucosa or if lymph node metastasis is suspected. The procedure is usually performed after confirmation of the extent of the lesion by iodine staining. An endoscopic mucosal resection (EMR) tube with a snare which seizes the lifted mucosa including its lesion is used. Finally, the mucosa is resected by electric current (Makuuchi et al. 1992, Yokoyama et al. 1995, Makuuchi 1996).

The rate of success is more than 90% when indications are respected, and the immediate morbidity rate is low (perforation less than 1%, death 0.1%). However, delayed complications are more common and are reported in about 6% of cases

(stenosis, recurrences, bleeding) (Kodama et al. 1998, Inoue 1998, Veuillez et al. 2007).

The problems arise when the lesion is not completely removed and additional excision is needed such as piecemeal resection carrying the possibility of tumor tissue implantation. In these cases, supplementary treatments such as photodynamic therapy and radiofrequency ablation (RFA) are often warranted.

Laser therapy

Laser therapy is today considered a palliative endoscopic therapy (Spencer et al. 2002). The tumor is vaporized or coagulated under direct vision with no mechanical stress on the esophageal wall. This technique is suitable for patients with exophytic tumors. Successful tumor recanalization can be achieved in more than 90% of the patients, and a subsequent return to eating solids can be obtained in the majority. Laser therapy does, however, need to be repeated every 4–6 weeks as the tumor regrows. This problem may be solved by combining it with adjuvant radiotherapy (Spencer et al. 2002). Palliative laser therapy has largely been replaced by self-expanding stents or been combined with these.

Photodynamic therapy

Photodynamic therapy (PDT) is generally used with the intent of curing early stages of esophageal cancer and as a palliative treatment for advanced and recurrent cancers. PDT is initiated with the administration of a photosensitizer (sodic porfimer, 5-ALA, meta-tetra hydroxyphenyl chlorine) and local tumor treatment is activated later using appropriate laser monochromatic light (Sibille et al. 1995, Veuillez et al. 2007).

The advantage of some of the PDT drugs is that they destroy cancer cells with less harm to normal cells. One drawback is that the photosensitizer must be activated by laser light; therefore only superficial cancers can be treated. The light cannot reach cancers that have expanded deeper into the esophageal wall or spread to other organs. As a palliative therapy, PDT does not eliminate all cancer tissue, but partially removes it and relieves the dysphagia.

When PDT is used on superficial cancers, the rate of success for this technique is approximately 100% but, depending of the drug used, includes a risk for symptomatic stenosis in nearly one-third of the cases (Barr 2003). In a single study, the 5-year survival rate was 74% (Sibille et al. 1995).

PDT, EMR and RFA, either alone or in combination, are simple, efficient and non-mutilating procedures that could be used on all localized lesions defined by a battery of markers to be precancerous even in patients with severe co-morbidity.

Multimodal treatment

Radio- and chemotherapy

When surgery is contra-indicated and the patients have no signs of distant metastases, infiltrating esophagus cancers may be treated using radio- and chemotherapy (Veuillez et al. 2007). The first report, by Herskovic in 1992, demonstrated that combined platinum-based radio- and chemotherapy treatment (50 Gy + five days with infusion of 5-fluorouracil (5-FU) and one day of cisplatin), followed by two cycles of the same chemotherapy regimen, generated better results than external irradiation alone. The radio- and chemotherapy combination resulted in a 25% 5-year overall survival rate compared to 0% for radiotherapy alone (Herskovic et al. 1992, Al-Sarraf et al. 1997). This study established the radio- and chemotherapy combination as a standard treatment for localized esophageal cancer (Stages II and III) (Veuillez et al. 2007). Other investigations, however, showed that there was no benefit in terms of overall survival in favor of secondary resection for patients responding to radio- and chemotherapy combination treatment (Stahl et al. 2005, Bedenne et al. 2007).

Recently, recommendations on the multimodal treatment (surgery and chemoradiation) of esophageal carcinoma have been published by Seitz and have been proposed as a general strategy for treatment of esophageal cancer (Seitz et al. 2006). Superficial cancers (*in situ* or T1-m1 or T1-m2, where “m” stands for the degree of infiltration into the mucosa) do not require multimodal treatment. Tumors covering more than two cm may be treated by mucosectomy. However, if the pathological examination reveals a more invasive tumor (T1-m3 or T2) with a risk for lymph node metastasis development, additional treatment is necessary and esophagectomy or radio- and chemotherapy combination has to be discussed depending on the condition of the patient.

Stage I (T1–T2, N0) invasive intrathoracic cancers are subject to surgical resection as standard treatment. In case of lymph node metastasis, adjuvant chemotherapy may be considered. In case of contra-indications to surgical resection, a radio- and chemotherapy combination may be appropriate.

Stage II (T1N1, T2N1, T3N0) invasive intrathoracic cancers usually undergo surgical resection preceded by neoadjuvant chemotherapy as standard treatment. Contra-indications to surgery entail the administration of radio- and chemotherapy.

Stage III (T3N1, T4N0–N1) intrathoracic invasive SCCs are subject to radio- and chemotherapy as standard treatment. In case of incomplete response or early recurrence, a salvage surgical excision must be considered.

Regarding AC, neoadjuvant chemotherapy before and after surgical resection is a valid option and by many considered the standard treatment (Cunningham et al. 2006). Locally inoperable cancers may be treated with a radio- and chemotherapy combination.

Radio- and chemotherapy combination is recommended when surgical resection is not possible for cervical cancers.

Non-operable patients without metastasis usually undergo a radio- and chemotherapy combination or receive an esophageal stent and occasionally an endotracheal stent in addition. The choice of treatment depends on the presence of esophago-tracheal or bronchial fistulae.

Patients with metastatic disease may be treated with radiotherapy or radio- and chemotherapy. An esophageal stent followed by chemotherapy is another option.

Common to all regimens is a disappointing low 5-year survival rate when clinical signs of cancer initiate treatment. Future strategy directed against symptoms (reflux) and dysplasia signs in premalignant lesions, combined with markers of malignant development and local treatment could improve the survival rate.

Palliative treatment

Stenting has become the treatment of choice for strictures with inoperable esophageal cancer (Guo et al. 2007 & 2008). However, stricture recurrence remains a challenge after a stent placement. Previously, endoscopic insertion of rubber stents was the first choice, but more recently, expanding metal stents have been introduced, as they are easier and safer to insert and less initial dilatation is required. Once in position, they expand across the tumor, but as experience shows, the swallowing quality is often not better, and the patients may have as many problems as with the rubber stents (Spencer et al. 2002).

Tumor invasion and development of metastases

Tumor invasion into the collagenous matrix and metastases represent the main problems in the treatment of carcinoma patients. The majority of patients with newly diagnosed carcinomas already present clinically detectable metastases (Fidler et al. 1978, Fidler & Hart 1982, Fidler & Balch 1987, Weiss 2000). In the tumor invasion process, cancer cells infiltrate the adjacent ECM by using several types of matrix degrading enzymes such as metalloproteinases, cysteine-, aspartic-, threonine and serine proteinases (Fig. 2) (Duffy 1987, Zukker 1988, Brunner & Preissner 1994, Keppler et al. 1994, Sloane et al. 1994, MacDougall & Matrisian 1995, Birkedal-Hansen 1995, Hewitt & Danø 1996). These enzymes are produced by the tumor cells and / or the surrounding host cells, and they cooperate with other proteins, for example integrins, cadherins and immunoglobulins, which may facilitate the dissolution process of the ECM (Hynes 1992, Natali et al. 1992, Morino et al. 1995, Takeichi 1991 & 1995 Rucklidge et al. 1994).

Next, neovascularization takes place, and cancer cells subsequently invade blood vessels in order to move to another organ. Adherent to the blood vessel walls,

malignant cells leave the primary circulation and migrate into the neighboring tissue, in a process called extravasation. They thus establish a new tumor site in the organism. The metastatic process is characterized by a highly selective competition, favoring the survival of a small subpopulation of metastatic cells (Liotta 1986, Fidler & Hart 1982). The metastatic subpopulation is abundant in the primary tumor tissue early in its growth, but less than 0.01% of malignant cells entering circulation actually form metastases (Kerbel et al. 1990, Fidler & Ellis 1994).

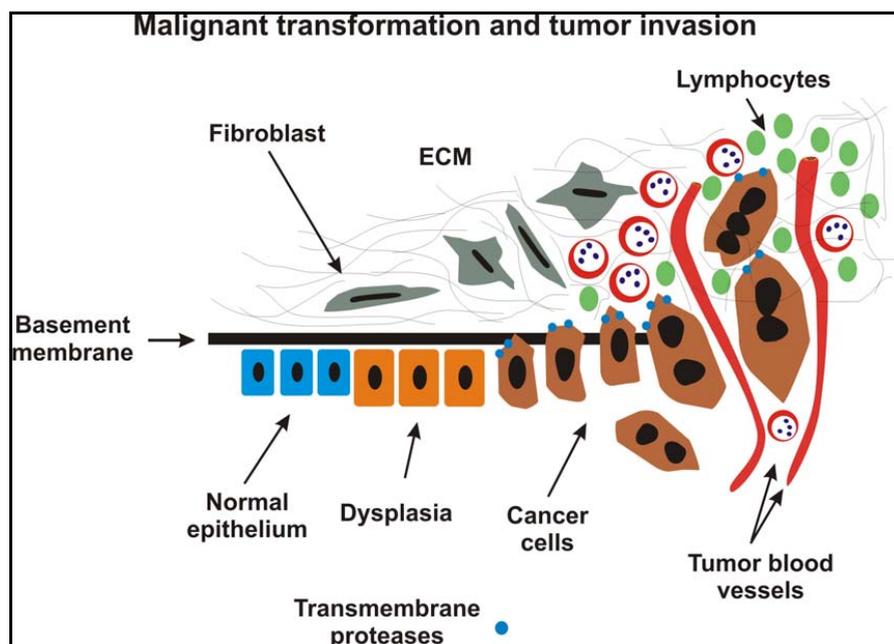


Figure 2. Invasive carcinoma development (modified after Liotta & Kohn 2001).

Membrane proteases

Membrane-bound proteases are widely spread among the different cell systems. Their expression in particular cell types is finely regulated, reflecting the specific functional cell implications and engagement in defined physiological pathways (Sedo et al. 2001). It has been reported that the proteases play a crucial role, both as effectors and regulatory molecules in protein turnover, ontogeny, inflammation, tissue remodeling, cell migration and tumor invasion (Sedo et al. 2001).

Several families of membrane proteases have been identified on the basis of their proteolytic activities, biologic functions and structural organization (Chen 2003, Chen & Kelly 2003):

1. Membrane-type matrix metalloproteinases (MT-MMPs)
2. Disintegrin and metalloproteinases (ADAM family)
3. Meprins
4. Secretases
5. Metallo- and serine peptidases (SIMP)

It is generally known that a given membrane protease may have several functions (diversity) and that more than one protease or protease family may mediate the same function (redundancy) (Bauvois B 2001). Soluble counterparts of some membrane proteases have been found intracellularly as well as in extracellular fluids, including blood plasma (Rettig et al. 1988, Sedo et al. 1996, Chiravuri et al. 1999, Abbott et al. 2000, Tang et al. 2000, Goldstein & Chen 2000, Chen 2003).

Localization of enzymes is critical for their function in cellular activities. It has been shown before that MT-MMPs and SIMP may have a prominent role in processing soluble factors as well as in degrading the components of the ECM (Sato et al. 1994). This study aims to examine this area in more detail and is focused on three members of serine protease family which are introduced below.

Seprase / FAP- α

FAP- α was first identified as an inducible antigen expressed on reactive stromal fibroblasts (Rettig et al. 1988 & 1993, Garin-Chesa et al. 1990). In parallel, seprase was originally isolated as a 170-kDa transmembrane protease from the malignant melanoma cell line LOX (Aoyama & Chen 1990, Monsky et al. 1994, Kelly et al. 1994). Further molecular cloning of FAP- α and seprase revealed the identical gene and protein (Scanlan et al. 1994, Piñero-Sánchez et al. 1997, Chen & Kelly 2003, Chen 2003). The gene is localized on the long arm of chromosome 2 (2q23). Seprase / FAP- α is a type II transmembrane protein of 760 amino acids, anchored in the plasma membrane by a short transmembrane domain, intracellularly exposing an amino terminal sequence, whereas a catalytic domain with a carboxyl-terminus remains extracellularly (Park et al. 1999, Levy et al. 1999). It displays both prolyl dipeptidyl peptidase and gelatinase activities. The protease appears as a homodimer (170 kDa) containing two 97 kDa subunits. Glycosylation and dimerization of the enzyme are necessary for its protease activity (Sun et al. 2002, Kelly 2005).

In humans, seprase has been identified in tissue remodeling sites, reactive stromal fibroblast in 90% of malignant epithelial tumors and sarcomas, granulation tissue of healing wounds and fetal mesenchymal tissue. The immunopathological expression of seprase has previously been investigated in gastric and colon cancer, as well as in melanoma, ovarian and breast cancer and its overexpression was associated with malignant phenotype (Okada et al. 2003, Iwasa et al. 2003). Seprase is not expressed in normal adult human tissue (Garin-Chesa et al. 1990, Rettig et al. 1993, Scalan et al. 1994, Ariga et al. 2001).

Dipeptidyl peptidase IV (DPPIV, CD 26)

DPPIV was first identified in 1966 as glycylproline naphthylamidase (Hopsu-Havu & Glenner 1966). DPPIV is an integral membrane glycoprotein with type II topology. Native human DPPIV is a 110 kDa protein, while the active form is a 200-220 kDa homodimer that exhibits the dipeptidyl peptidase activity (Piazza et al. 1989, Johnson et al. 1993, Piñeiro-Sánchez et al. 1997). The DPPIV structure contains 766 amino acid residues. The human gene of DPPIV is localized on the long arm of chromosome 2 (2q24.3). Peptidase is expressed constitutively on brush border membranes of intestine and kidney epithelial cells and transiently in activated T-cells and migratory endothelial cells (Vivier et al. 1991, Yaron & Naider 1993, Morimoto et al. 1994).

It was shown that DPPIV, in addition to its typical dipeptidyl aminopeptidase activity, may possess endopeptidase activity as well (Bermpol et al. 1998). DPPIV expression and activity was observed in numerous types of human malignancies (basal cell carcinoma, prostate-, ovarian- and thyroid carcinoma) as well as in blood plasma of cancer patients (Hirai et al. 1999, Wilson et al. 2005, Ozog et al. 2006). In general, higher DPPIV expression is associated with more aggressive tumor behavior.

In contrast, in ovarian carcinoma cell lines, DPPIV overexpression was associated with a decrease in invasive potential, change in morphology, reduction of intraperitoneal dissemination of carcinoma cells and prolongation of survival time in vivo (Kajiyama et al. 2002, Kikkawa et al. 2005). Functional studies have also demonstrated that loss of DPPIV expression during malignant transformation of melanocytes is accompanied by growth factor independence, whereas its experimentally induced re-expression leads to the suppression of tumorigenicity, reversal of a block in differentiation and re-emergence of requirements for exogenous growth factor (Wesley et al. 1999, Pethiyagoda et al. 2000).

Finally, a soluble form of DPPIV modulating the responsiveness of T-cells to specific antigens has been detected in blood plasma (Tanaka et al. 1994).

uPA

The serine protease family includes also uPA, a glycoprotein with a molecular weight of 55 kDa, known to operate extracellularly. It is activated on the cell surface, binding to a specific receptor (uPAR), which is linked to the plasma membrane and may form complexes with seprase, DPPIV and β 1 integrin. uPA cleaves plasminogen to form active plasmin, breaking down most ECM components, including type IV collagen, laminin and fibronectin (Hansen et al. 1994).

An elevated level of uPA has been involved in the development of invasiveness in numerous neoplasms, including breast-, ovarian-, gastric- and colorectal cancer, as well as SCC and AC (Nishino et al. 1988, Sier et al. 1991, Hewin et al. 1995, Torzewski et al. 1997, Nekarda et al. 1998, Artym et al. 2002).

Serine protease complexes

Former studies have revealed that serine proteases are able to form complexes. Gherzi and Dong, for example, have shown that seprase and DPPIV are simultaneously involved in the degradation of the collagenous matrix (Gherzi et al. 2002). The two proteases form a complex localized at invadopodia of cells migrating on collagenous fibers that elicits both endo- and exopeptidase activities (Chen 2003). The complex, described as a 400 kDa molecule, develops into a potent ECM degrading factor (Piñero-Sánchez et al. 1997, Mueller et al. 1999, Gherzi et al. 2002).

Immunoprecipitation, immunofluorescence and cell surface crosslinking experiments demonstrated another seprase-DPPIV complex with $\alpha 3\beta 1$ integrins, additionally facilitating invasion into the collagenous matrix (Mueller et al. 1999, Chen 2003). Interestingly, $\alpha 3\beta 1$ integrin is even able to bind uPAR (Zhang et al. 2003). This binding demonstrates the possibility of creating a supramolecular complex consisting of seprase-DPPIV-uPAR/uPA- $\alpha 3\beta 1$ integrin, exposing common proteolytic activity (Scanlan et al. 1994, Gherzi et al. 2002, Artym et al. 2002, Kelly T 2005).

A recent study, performed on HUVEC, has shown a seprase-DPPIV complex (~ 820 kDa) localized at an invadopodia-like protrusion of endothelial cells involved in the invasion of the ECM (Gherzi et al. 2006). *In vivo* examination on invasive breast ductal carcinoma specimens has shown a distribution of the complex on the endothelial cells of capillaries, but not on large blood vessels, contributing thus to understanding the role of serine proteases in the angiogenetic process (Gherzi et al. 2006).

A simplified interaction model between the serine proteases, integrins and ECM is illustrated in Figure 3.

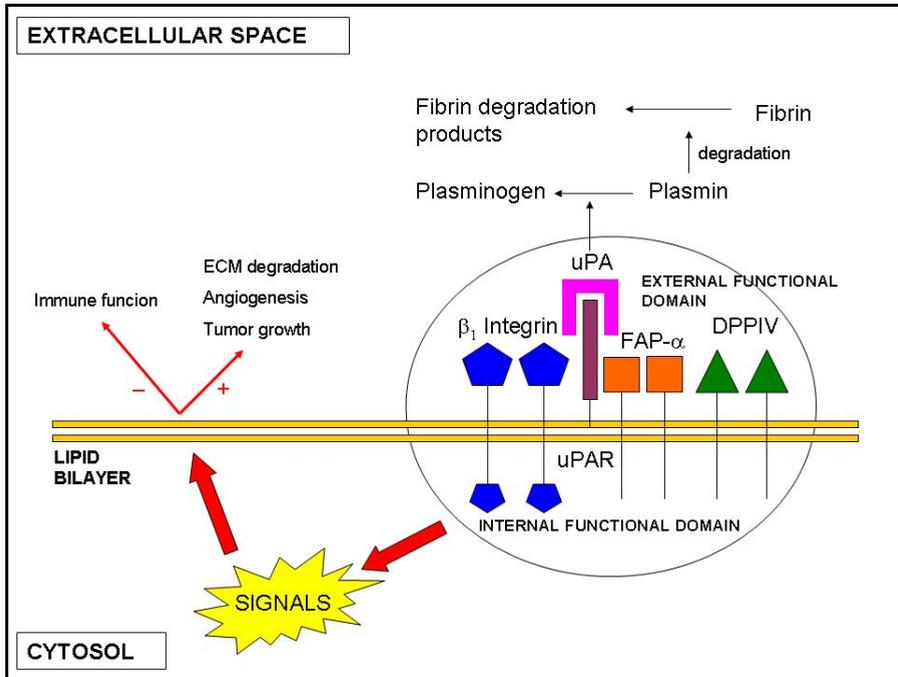


Figure 3. Possible serine protease complex. DPPIV, Fap- α , uPA / uPAR and β_1 integrin with proteolytic, adhesive and signaling capabilities. The complex may act intra- and extracellularly, demonstrating a common enzymatic activity (modified after Kelly 2005).

4. AIMS OF THE STUDY

Little information about expression patterns of the serine protease family members in esophageal carcinomas was available when this study was initiated. Though uPA has previously been analyzed in esophageal carcinomas, the role of seprase and DPPIV in this carcinoma type is not yet known. No systemic study of expression patterns of all three serine proteases together in the upper gastrointestinal tract and their clinical association was available. Thanks to the fact that the study partly was a collaboration project between the Norwegian Radium Hospital, Rikshospitalet HF in Oslo, Norway and Anyang Tumor Hospital, Henan Province in China, we were able to collect extensive material from this particular tumor type. This gave us the unique opportunity to study the expression of serine proteases in normal, premalignant and malignant stages of both SCC and AC in order to better understand their involvement in neoplastic progression and the possible use as marker of malignant or metastatic potential.

Furthermore, from a clinical point of view, no complete report has been published until today, assessing patients treated at the Norwegian Radium Hospital, Rikshospitalet HF with esophageal carcinoma. Using our clinical databases and tissue material, we examined clinical and histopathological information taking into account all the esophageal cancer patients treated at the Norwegian Radium Hospital, Rikshospitalet HF between 1987-2007 in order to analyze epidemiology, occurrence of precancerous stages and treatment results.

The specific aims were to:

- Compare seprase, DPPIV and uPA expression in dysplastic and cancer cells of SCC, as well as in stromal cells adjacent to premalignant and malignant sites
- Examine seprase and uPA expression in Barrett's esophagus, dysplasia and AC, as well as in stromal cell bordering neoplastic alterations
- Compare DPPIV expression in cancer and stromal cells of SCC and AC
- Demonstrate stromal serine protease expression
- Describe and analyze clinicopathological features of the patients with esophageal carcinoma

5. MATERIALS AND METHODS

Patients and tissue samples

This study included two independent patient groups:

1. Norwegian patients who were diagnosed with esophageal dysplasia or esophageal cancer in the period from January 1987 until December 2007 and underwent periodic surveillance or received partial or complete treatment at the Surgical Oncology Department of the Norwegian Radium Hospital, Rikshospitalet HF in Oslo, Norway (Papers I, III and IV).
2. Chinese patients who were diagnosed with esophageal dysplasia or esophageal cancer between 2003 and 2005 and underwent periodic surveillance or received treatment in Anyang Tumor Hospital, Henan Province, China (Paper II).

Diagnostic tests and surgical or oncological treatments were performed depending on the clinical symptoms and on the stage of advancement of the disease, consistent with the conventional rules of therapy of esophageal cancer, including ongoing clinical trials and accepted palliative care.

Tissue samples of Barrett's esophagus and dysplasia were obtained from patients who underwent diagnostic tests as a result of increasing symptoms of dysphagia or a local irritation in the esophagus. Chest and esophageal x-rays with contrast, blood tests and endoscopy with a subsequent pathological evaluation of tissue samples were performed at the Norwegian Radium Hospital, Rikshospitalet HF and at Anyang Tumor Hospital, depending on the samples' origin. The dysplasia samples were obtained by gastroscopy with a flexible gastroscope. They were fixed in formalin, embedded in paraffin and stained with haematoxylin and eosin.

Tissue samples of advanced esophageal carcinomas (SCC and AC) were obtained from nonoperated patients by gastroscopy (as a diagnostic test before choosing treatment) and from patients who underwent potentially curative resection of the esophagus (preceded by adequate diagnostic tests) in the above mentioned hospitals. Potentially curative resection was defined as removal of all gross tumor tissue, histologically confirmed absence of tumor tissue at the surgical margins and absence of distant metastases (Torzewski et al. 1997). The patients received either no treatment prior to surgery or underwent neo-adjuvant radiotherapy. The gastroscopies were carried out using a flexible gastroscope. Esophagectomies were performed through laparotomy and a right- or left-sided thoracotomy with an abdominal approach through the diaphragm. Subsequent reconstitution was completed mostly by means of esophagogastronomy, using the gastric tube through the retrosternal route, with construction of a cervical anastomosis. The surgery samples underwent the same fixation procedures as the dysplasia samples and were embedded in paraffin as a single sample (Norwegian tissue material), or were stored as tissue arrays (Chinese tissue material); each array contained tissue samples from 37 to 70 different patients.

As control group, samples of normal esophageal epithelium were collected from Norwegian patients, primarily operated at the Department of Surgical Oncology of the Norwegian Radium Hospital, Rikshospitalet HF for esophageal cancer between 1993 and 1999. Tissue samples were taken from normally looking squamous epithelium, located >5 cm from the tumor site. These samples were prepared for investigation in the same way as the samples described before. No tumor tissue was observed in the obtained samples, either macroscopically or microscopically.

In accordance with the WHO (WHO 1990), the dysplasia samples were histologically categorized into three groups: low-, moderate- and high grade dysplasia. The WHO classification also divides SCCs and ACs into three groups: well-, moderately- and poorly differentiated. Clinical classification was completed using TNM staging according to UICC Global Cancer Control (UICC 2005).

Cell lines

Cells deriving from well-, moderately- and poorly differentiated esophageal SCC respectively (KYSE450, KYSE140 and KYSE70 cell lines), and cells from normal esophageal epithelium (HET1A) were used in our study. The cell lines were provided by Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany.

Antibodies

We used the following antibodies in our experiments:

- Rabbit polyclonal antibody, isotype IgG, directed against human FAP- α , purchased from Abcam, Cambridge, UK
- Monoclonal rat antibody (clone E26), directed against the 200-220 kDa form of DPPIV obtained from Prof. W-T Chen (Department of Medicine, State University of New York, Stony Brook, N.Y., USA)
- Anti-human dipeptidyl peptidase IV, polyclonal goat antibody, obtained from R&D Systems, Minneapolis, MN
- Rabbit polyclonal antibody, directed against human HMW-scuPA (54 kDa), HMW-tsuPA (52 kDa) and LMW-scuPA (33 kDa), purchased from Abcam, Cambridge, UK

Laboratory methods

Immunohistochemistry

Tissue samples were fixed in 4% buffered formalin, embedded in paraffin, cut at four microns, dried in the oven at 70 °C overnight before deparaffinization and rehydration through decreasing concentrations of alcohol to running tap water. The slides required no pre-treatment. The EnVision+ system from Dako Cytomation was used according to the kit manual, and haematoxylin was used for counterstaining. Appropriate

negative and positive controls were applied according to the antibodies' specificity (M&M, papers I-III).

Semi-quantitative analysis of immunohistochemistry

The seprase-, FAP- α -, DPPIV- and uPA expressions were semi-quantified using a visual grading system in which the staining intensity was categorized into four groups; 0, 1+, 2+, 3+, where group 0 was defined as having a complete absence of staining. Groups 1-3 were defined as groups with positive staining of increasing intensity as compared to the positive control. The number of positive cells was also categorized into four groups, where group 0 was defined as total absence of positive cells, and groups 1-3 were defined as groups with positive cells counted in percentages; 0 = 0%, 1 = <25%, 2 = 25-50%, 3 = >50%. The outcome was calculated by multiplying the corresponding values from staining intensity by the number of positive cells and was subsequently divided into four final groups: 0, 1+, 2+ and 3+.

In order to guarantee quality control, two independent pathologists performed the sample interpretation separately. Diverging cases were discussed until an agreement was reached.

Immunoblotting

Cells from cell lines were lysed in cold TGH buffer (1% Triton X-100, 10% glycerol, 20mM Hepes, pH 7.2, 100mM NaCl) containing 1mM phenylmethylsulfonyl fluoride, 10 μ g/ml leupeptin and 1mM Na₃VO₄. After shaking, the lysate was incubated for 60 min at 4 °C with rocking. Insoluble materials were removed by centrifugation (14.000 rpm x g, 20 min) at 4 °C, and the supernatant was collected. Total protein concentration in each sample was estimated with the Bradford analysis (Bio-Rad). SDS-PAGE was performed in 12% gels as described by Laemmli (Laemmli 1970), and the proteins in the gel were transferred to a polyvinylidene difluoride (PVDF) membrane (Immobilon-FL, Millipore, Bedford, MA). After one h blocking with 5% non-fat dry milk powder in TBS (TRIS Buffered Saline consisted of 137 mM NaCl, 25 mM TRIS and 2.7 mM KCL, pH 7.6) containing 0.05% Tween-20, the membranes were incubated at 4 °C overnight with tested antibodies at adequate dilution. The membranes were then washed three times for 10 min with PBS (Phosphate Buffered Saline consisting of 137 mM NaCl, 12 mM phosphate and 2.7 mM KCL, pH 7.6)/0.1% Tween-20 and finally incubated with the appropriate secondary antibody conjugated to horseradish peroxidase for one h at room temperature. The membranes were then again washed three times for 10 min with TBS/0.05% Tween-20. A chemiluminescent detection reagent (ECL Plus, Western blotting detection system, GE Healthcare) was used for peroxidase signal detection. To ensure equal loading of proteins, the same membranes were reprobred with rabbit polyclonal anti ERK-2 (SC-154) antibody (Santa Cruz Biotechnology, CA, USA) at dilution 1:50000 in 5% milk.

Quantitative real-time RT-PCR analysis

Total cellular RNA was extracted from cell lines (KYSE450, KYSE140, KYSE70 and HET1A) using the TRIZOL reagent (Invitrogen, Carlsbad, CA). The high capacity cDNA reverse transcription kit (Applied Biosystems, Foster City, CA) was used to reverse-transcribe obtained RNA (0.8 μ g) in a 20 μ l reaction mixture using random

primers. The real-time PCR analyses were performed using TaqMan Fast Universal PCR Master Mix and TaqMan Gene Expression Assays for FAP- α , DPPIV, uPA and GUS (Applied Biosystems). A total of 5 μ l cDNA, diluted at 1:10, was used in 25 μ l PCR mixtures with 900 nM of each primer and 250 nM TaqMan probe. The reactions were carried out in a 7900HT Fast Real-time PCR system (Applied Biosystems) with the following program: 95 °C for 20 s followed by 40 cycles of 95 °C for one s, 60 °C for 20 s. Each sample was run in triplicate. The threshold cycle (CT) values of the amplification reactions were determined automatically using RQ Manager 1.2 software (Applied Biosystems). The FAP- α , DPPIV and uPA relative mRNA expression level was normalized with respect to the beta-glucuronidase (GUS) gene, which had stable transcript levels under these experimental conditions. The tumor / normal ratio of the normalized target transcript expression was calculated by means of the $2^{-\Delta\Delta C}$ method (Livak & Schmittgen 2001) from three independent experiments.

Enzymatic activity biochemical assay

For enzymatic activity assays, near-confluent cells from KYSE450, KYSE140, KYSE70 and HET1A cell lines were incubated for 18 h in fresh medium (RPMI 1640, Invitrogen) containing 1% FCS. Then, media were collected and cells were lysed in lysis buffer (0.1 M NaCl, 10 mM Na₂HPO₄, 1% Triton X-100, 1 mM EDTA). The DPPIV-like activity was assayed by measuring the cleavage of Gly-Pro-NH-Np substitute (Gly-Pro-4-nitroaniline, Sigma) at pH 8.0 by either cell lysate or medium collected from cells after 18 h incubation, according to the standard procedure provided by Sigma. Twenty-five μ l of sample (medium or lysate) were incubated in 0.1 M Tris (pH 8.0) with 0.5 mM Gly-Pro-NH-Np in a total volume of 200 μ l at 37 °C for different time points in flat-bottom 96-well microplates. Activity was determined by measuring absorption at 405 nm with Biotrak Microplate Reader (Amersham). uPA activity was measured in an analogous way (using the chromogenic substrate) with the CHEMICON uPA Activity Assay Kit. Total protein concentration in samples was assayed by the Bradford method (Bradford 1976).

Statistical analysis

Associations between variables were assessed using Chi-square tests (Pearson and linear-by-linear association). Differences between quantitative variables in independent groups were tested by Mann-Whitney tests. They were all two-sided tests. Comparison of survival between the groups was performed using log-rank tests. Survival curves were calculated with the Kaplan-Meier product-limit method. A p-value of <0.05 was considered statistically significant. Statistical analyses were performed using SPSS 13.0 and 15.0 for Windows.

6. SUMMARY OF PAPERS

PAPER I

DPPIV is a transmembrane serine protease involved in the process of tumor invasion and development of metastasis in human cancers. In this article we investigated the expression of DPPIV in cancer and stromal cells of both esophageal AC and SCC. We analyzed tissue material from 159 patients treated in our hospital between January 1987 and November 2002 (SCC (n=90) and AC (n=69)) using immunohistochemistry. The patients were divided into two groups to obtain more specific results: irradiated and non-irradiated patients (n=46 and n=113 respectively).

We also performed Western blotting on SCCs and normal esophageal epithelium cell lines, as well as on fresh frozen tissues from ACs and normal esophageal epithelium. Results of immunostaining were compared with the patients' clinicopathological features.

Immunohistochemistry of the whole population revealed differences between DPPIV expression in AC and SCC cells: DPPIV expression was higher in ACs than in SCCs. In contrast, DPPIV expression in stromal cells was higher in SCC stroma than in AC stroma. In the whole population, DPPIV was also significantly associated with lymph node metastases, distant metastases and age. The expression pattern revealed that a high expression of DPPIV involved a higher risk of distant- and lymph node metastases. The correlation between DPPIV expression and age was proportionally inverted: carcinoma cells of older patients contained a lower level of protease.

The separate analyses of SCC and AC showed a correlation between the expression of DPPIV in cancer cells and distant metastases only in the AC group. We did not find any associations between DPPIV expression in cancer cells and clinicopathological features in SCCs. Further analyses of the two groups showed no associations between DPPIV expression in stromal cells and clinicopathological features. Radiotherapy had no impact on DPPIV expression in the analyzed tissue samples. There was no correlation between DPPIV expression in cancer or stromal cells and survival of the patients.

Immunoblotting did not show any detectable DPPIV in the cell line originating from normal esophageal epithelium or fresh-frozen tissue samples of normal esophageal epithelium. In contrast, DPPIV was present in all carcinoma samples. SCC cell lines showed a decreasing intensity of DPPIV from well to poorly differentiated carcinomas. In the fresh-frozen cancer tissue samples, the highest DPPIV expression was detected in moderately differentiated carcinomas. Poorly and well differentiated carcinomas showed a lower expression of DPPIV.

Thus, differences in the DPPIV level in esophageal carcinomas compared with normal epithelium showed that esophageal malignancies were associated with an increased amount of cell surface-bound DPPIV.

PAPER II

In an attempt to elucidate the role of the serine proteases seprase, DPPIV and uPA in the degradation of the ECM and in the progression of esophageal SCC, we studied the expression of the proteases in esophageal dysplasia, invasive carcinomas and normal epithelium by using immunohistochemistry. Tissue samples from different stages of dysplasia (n=85) and from advanced SCCs at different stages (n=144) were obtained from patients treated at Anyang Tumor Hospital, Henan Province in China. Additionally, we performed real-time RT-PCR, Western blotting and enzymatic activity biochemical assays on cell lines originating from different SCC grades and from normal esophageal epithelium.

Seprase, DPPIV and uPA immunoreactivity were demonstrated in dysplastic and cancer cells as well as in stromal cells adjacent to dysplasia and cancer sites, but not in normal epithelium. A significant association between uPA expression and sex, tumor size and histological classification in carcinomas was shown. Increased expression of DPPIV in cancer cells correlated with longer patient survival. Furthermore, no significant associations between seprase and clinicopathological features either in dysplasia or in carcinomas could be found. Finally, using Western blotting, real-time RT-PCR and enzymatic activity assays, we demonstrated higher levels of the proteases in SCC cell lines than in normal esophageal epithelial cell lines.

These results showed that seprase, DPPIV and uPA were expressed in both premalignant and malignant forms of SCC, but lacking in normal esophageal epithelium, confirming thus their involvement in the SCC neoplastic progression. This fact indicates the possibility that these serine proteases play a role in the transformation from normal to dysplastic epithelium.

PAPER III

We examined FAP- α and uPA expression in premalignant and malignant stages of esophageal AC by immunohistochemistry. The findings were compared with clinicopathological features. Tissue samples of Barrett's esophagus with and without dysplastic changes were obtained from 22 patients, whereas tissue samples of irradiated and non-irradiated ACs of different histological grades were collected from 69 patients. We also performed Western blotting on fresh-frozen tissue samples obtained from both carcinoma tissue and normal esophageal epithelium.

There were no significant associations between the protein expressions in metaplastic and dysplastic cells and the histological grading or development of cancer. In contrast, stromal uPA expression was significantly higher in metaplastic and low- / moderate grade dysplasia than in high grade dysplasia. No such association could be found for FAP- α .

We did not find any correlations between FAP- α and uPA expression and clinicopathological features in carcinoma cells. Division of AC patients into two subgroups, irradiated patients (n=18) and non-irradiated patients (n=51), did not reveal any changes in the analyses.

Examination of the stromal protease expression on the other hand, exhibited significant correlations between FAP- α and depth of tumor invasion, as well as uPA and lymph node metastasis. High stromal FAP- α expression was detected in T1 tumors, compared to a low FAP- α activity in T2-T4 tumors. Stromal uPA overexpression was found in tissue samples of patients without lymph node metastases, while low uPA expression was associated with lymph node metastases.

The analysis of patients relating to radiotherapy revealed that non-irradiated patients had a higher stromal FAP- α concentration than irradiated patients.

We did not find any correlation between FAP- α or uPA expression and survival of the patients, but due to the modest number of patients, a type II error can not be excluded.

Immunoblotting showed higher protease expression in carcinoma tissues than in normal esophageal epithelium.

These results suggest that FAP- α and uPA expression in metaplastic, dysplastic and esophageal cancer tissue, including stroma adjacent to cancer sites, is associated with the early neoplastic progression of esophageal lesions.

PAPER IV

Esophageal cancer is not a common cancer in Norway, but remains a problem due to low survival rates and considerable treatment morbidity. We analyzed and compared clinical and pathological data of patients treated at the Norwegian Radium Hospital, Rikshospitalet HF, Oslo, during the last two decades (1988-1997 and 1998-2007). During the last 20 years, we observed a significant change in the occurrence of esophageal SCCs in relation to ACs in the Norwegian population. The AC incidence has increased from 5-10% to more than 50% in the last decade, while the incidence of SCCs has decreased. Additionally, a change in the pattern has also been observed: ACs occur in younger patients and are primarily related to GERD.

An important aspect of this two-decade study was to evaluate the general impact of anti-reflux therapy and surveillance of premalignant lesions and if this could be reflected in tumor stage, patient demographics and treatment results. This was not a randomized study but it gave us a possibility to get an impression of the effect of preoperative radiation on some biological markers and on the final treatment outcome, survival and recurrence rates. Based on the observations of our patients, we concluded that no significantly improved survival could be demonstrated from preoperative irradiation, but a trend towards improvement was seen. Finally, we found that the change from SCC to AC did not lead to improved long time survival in our patients.

Our results allowed us to observe that the most important epidemiological difference between SCC and AC is the strong association between GERD and AC. Furthermore, Barrett's esophagus was clearly associated with AC in more than one-fourth of the cases and probably in a lot more due to the fact that small Barrett's lesions are often completely supplanted by tumor tissue. Almost all patients suffered from dysphagia or dysphagia-related symptoms at the time of diagnosis, demonstrating that even for AC, usually in the distal esophagus, this is a late symptom. A very modest increase in early cancer lesions referred in the last decade demonstrates that an increased vigilance of GERD-induced lesions may be of importance for decisive local treatment instead of esophagectomy following dysphagia diagnosed advanced tumors.

In conclusion, upcoming studies should focus on developing strategies for early detection of the disease, using specific markers appearing in precancerous stages, separating thus completely or partially premalignant from malignant stages.

7. GENERAL DISCUSSION

The serine proteases' ability to degrade the ECM is considered to be a prerequisite for invasion and spreading of cancer cells (Duffy 1987, Zukker 1988, Birkedal-Hansen 1995, Hewitt & Danø 1996). Numerous factors are involved in this process, but the direct and indirect interaction between the proteases and the basement membrane seems to play a pivotal role in the initiation of tumor cell invasion (Duffy 1987).

Previous studies on serine proteases focused mainly on enzymes derived from tumor cells (Nishino et al. 1987, Torzewski et al. 1997, Kikkawa et al. 2004). Recently, however, enzymes produced by stromal cells bordering tumor sites have also been considered as a strong factor implicated in tumor progression (Iwasa et al. 2003, Mori et al. 2004). Although serine protease expression has been investigated in several carcinomas, there is still limited knowledge about their presence in esophageal malignancies.

Thanks to our collaboration with Anyang Tumor Hospital in China, we have access to a large series of esophageal carcinomas allowing us to study serine protease expression in premalignant and malignant stages of both SCC and AC. SCC, previously considered a dominant form of esophageal cancer worldwide, shows today a slightly decreasing incidence in the Western world, while it remains the major esophageal carcinoma type in the developing countries (Devesa et al. 1998, Blot & McLaughlin 1999). The SCC occurrence in certain areas of Asia or Africa is so high that screening programs could be considered a method for detecting premalignant and early stages of carcinoma, trying thus to reduce mortality among specific populations (Sammon 1998, Saidi 1999, Wang et al. 2002, Yang et al. 2002,). The Western world, on the other hand, suffers from increasing numbers of AC, whose association with GERD and Barrett's esophagus is commonly known. The latter could be considered a premalignant stage and by many suggested to undergo strict surveillance (Blot & McLaughlin 1999, Lagergren et al. 1999). The long-time effects on survival from esophageal cancer of such surveillance regimens have so far not been convincingly demonstrated.

We still believe that monitoring precancerous stages is important for early detection and local intervention and thus reducing mortality and morbidity. No blood test has yet been found which could distinguish premalignant from malignant stages. Histological biopsies and morphological evaluation still remain the major diagnostic methods (Dong et al. 2002).

In the current study, we focused on the analysis of three interesting indicators (seprase / FAP- α , DPPIV and uPA) and their possible role in the evolution of premalignant and malignant stages of both major types of esophageal carcinoma, SCC and AC. We compared their expression levels with clinicopathological features in a series of patients, using the clinical databases of the two previously mentioned cancer hospitals (papers I-III).

The analysis of clinical information related to Norwegian patients (Paper IV) has given a substantial insight into the rapidly changing esophageal carcinoma epidemiology in Norway and our diagnostic and treatment methods over the last two decades. Both pathological and clinical studies contributed to a better understanding of the evolution and differences between the two types of esophageal carcinomas and their premalignant stages. Results obtained from histopathological and biochemical studies pointed to the identification of possible markers that could be used in future clinical work to characterize precancerous stages as well as invasive cancers.

Main findings

Main findings concerning the expression of serine proteases in the analyzed tissue samples:

- Seprase, DPPIV and uPA immunoreactivity was found in dysplastic and SCC cells as well as in stromal cells adjacent to dysplasia and cancer sites, but not in normal epithelium (Paper II).
 - *A significant association between uPA expression in carcinoma cells and sex, tumor size and histological classification in SCCs was found.*
 - *High expression of DPPIV in carcinoma cells correlated with longer survival of the patients with SCC.*
 - *No significant associations between seprase expression and clinicopathological features either in dysplasia or in carcinomas were found.*
 - *SCC cell lines had higher levels of seprase, DPPIV and uPA than normal esophageal epithelial cell lines.*

- DPPIV displayed a significantly higher level in ACs as compared to SCCs, while no DPPIV was detected in normal esophageal epithelium (Paper I).
 - *Cellular overexpression of DPPIV in patients with AC was associated with distant metastases.*
 - *Radiotherapy in patients with SCC or AC had no impact on DPPIV expression in the analyzed tissue samples.*

- Seprase and uPA were detected in metaplastic-, dysplastic- and AC cells, as well as in stroma bordering metaplasia-, dysplasia- and carcinoma sites (Paper III).
 - *Stromal seprase expression was associated with depth of tumor invasion, while stromal uPA expression correlated with lymph node metastases in AC.*
 - *Stromal uPA expression in cells with dysplasia correlated with histological grading.*
 - *Higher protease expression in AC cells than in normal esophageal epithelium was found.*

Main findings concerning the analyses of the clinicopathological material from referred patients (Paper IV):

- Esophageal cancer showed an increased occurrence of AC in the distal part of the esophagus and, in the last two decades, was frequently associated with Barrett's esophagus in the Norwegian population.

- No significantly improved survival of the patients could be demonstrated from preoperative irradiation, but a trend towards improvement was seen.
- The change from SCC to AC did not lead to improved long time survival of the patients.

Serine proteases in dysplastic-, cancer- and stromal cells of SCC and AC

When analyzing tissue samples of different stages of dysplasia evolving into SCC (Paper II), we detected the strongest serine protease immunostaining in grade III dysplastic cells (carcinoma *in situ*) and the weakest in grade I, which turned out to be a common pattern for all three enzymes. The same staining pattern was also found in stromal cells adjacent to dysplasia sites, while normal epithelium did not show any staining at all. Considering the absence of serine protease expression in normal epithelium, the point of transformation from normal to dysplastic epithelium may be crucial for the activation of serine proteases, and, subsequently, their extracellular and pericellular proteolytic activities. Furthermore, the activity, or most likely, the production of serine proteases in both dysplastic cells and stromal cells adjacent to dysplasia increased in parallel to the advancement of dysplasia, showing a clear, positive association between the serine protease level and the degree of premalignancy. Finding serine proteases in premalignant and stromal cells indicates a close relationship between dysplastic and stromal cells, where dysplastic cells possibly stimulate stromal cells to generate e.g. serine protease activity. In relation to this and to what will be discussed further below, we thus concluded that serine proteases contribute to all stages of both premalignant and malignant progression in SCCs, and not only to the latter, as was previously suggested (López-Otín & Matrisian 2007).

On the other hand, when focusing on the expression of seprase and uPA in Barrett's metaplasia and dysplasia, eventually developing into AC (Paper III), no apparent association between protease expression and dysplasia stage could be identified, although the presence of proteases was evident in both dysplastic and stromal cells. Like in the case of SCCs, this finding too points towards an interplay between premalignant and stromal cells in this kind of carcinoma. It is noteworthy that this common trait concerned two histologically different premalignant conditions. With regards to this finding, we believe that the transmembrane proteases exist as signaling molecules on dysplastic cells and stromal cells adjacent to dysplasia in precancerous stages of both SCC and AC, probably interacting with other molecular factors exhibiting enzymatic activity, contributing to the development of invasive carcinomas.

The common immunostaining pattern for all tested proteases observed in dysplastic and stromal cells (Paper II) supported former studies, indicating the presence of supramolecular complexes such as seprase-uPA, seprase-DPPiV or even seprase-DPPiV-uPA on the cell membrane (Fig. 3) (Scanlan et al. 1994, Ghersi et al. 2002 & 2006, Chen & Kelly 2003, Kelly 2005). Although we did not specifically investigate the presence of such complexes in dysplastic or stromal cells, the similar enzymatic

activity of all three proteases detected in DPPIV-like and uPA enzymatic activity assays performed on cell lines reinforced the hypothesis about serine protease complexes.

Analysis of DPPIV alone in carcinoma cells (Paper I) showed that DPPIV expression was significantly higher in ACs than in SCCs, indicating that tumors originating from histologically different epithelial types may influence the production or expression of protease to a different degree. We presume that metaplasia occurring in the distal esophagus, where columnar cells replace stratified epithelium, and where dysplasia and AC frequently develop, significantly enhances DPPIV production in affected cells. This assumption is in agreement with Chaves' study (Chaves et al. 1999) in which she detected DPPIV in Barrett's esophagus and AC related to it, thus showing a direct association between these two conditions. Enhanced protease production may even be a necessary condition for the metaplastic process *per se*. The lower level of DPPIV found in SCCs in our study does not lessen the proteases' role in the development of this carcinoma type, but indicates that, in addition to DPPIV, other cell-surface-associated proteolytic enzymes could strongly be involved in the growth of SCCs, or, that SCCs require less DPPIV activity compared to ACs in order to develop equal invasive potential.

Furthermore, we found that DPPIV was also present in stromal cells bordering cancer sites (Paper I), similarly to previously mentioned findings in dysplastic changes. Stromal DPPIV presence in SCC, as well as in AC tissue samples, may indicate that cancer cells of histologically different carcinoma types can even influence adjacent stromal cells and initiate an additional stromal production of proteases. This finding is similar to other studies showing the presence of the DPPIV homologue, seprase, in fibroblasts, macrophages and endothelial cells adjacent to tumor sites in human gastric and colorectal cancer, where stromal seprase expression was associated with lymph node and liver metastases (Iwasa et al. 2003, Okada et al. 2003, Mori et al. 2004). Our results indicate that stromal DPPIV (no association between stromal DPPIV and clinicopathological features) is probably not directly, or only modestly responsible, for ECM degradation in esophageal carcinomas. It could rather be a useful activator of other enzymes, i.e. metalloproteinases or DPPIV-associated serine proteases which serve as workhorses.

Investigation of seprase, DPPIV and uPA occurrence in SCCs (Paper II) and of FAP- α and uPA in ACs (Paper III) showed again expression of the serine proteases not only in carcinoma cells but also in neighboring stromal cells. The immunoreactivity of uPA and DPPIV in SCC cells was characterized by a decreasing intensity pattern from well differentiated towards poorly differentiated carcinomas. As for seprase, although an expression of more than 93% was observed in SCC samples, we could not demonstrate any distinct expression pattern. AC cells, in contrast, despite abundant FAP- α and uPA presence, showed no correlation between serine protease expression and carcinoma advancement (Paper III). Interestingly, stromal FAP- α and uPA expression was inversely correlated to T and N and, in consequence, to tumor progression in general.

We suggested previously that cancer cells are able to stimulate the production of proteases and other enzymes essential to ECM degradation in stromal cells, which is consistent with other studies (Sawhney et al. 1992, Iwasa et al. 2003). The stromal serine protease expression could also originate from serine proteases which are mainly produced by cancer cells and may attach to stroma during their activation (Chen et al. 2006). Regardless of the cause of its expression, stromal protease expression denotes that stroma is able to partly assume control over the serine protease production in advanced cancer stages, possibly as a result of a reduced serine protease production in highly undifferentiated carcinoma cells. Intensification of such stromal behavior may be dependent on the carcinoma histology. For example, the fact that DPPIV and uPA expression in SCC cells decreases, whereas tumor aggressiveness increases, clearly illustrates this aspect (Paper II). A question arises at this point: do stromal cells take over the serine protease production from poorly differentiated carcinoma cells in SCCs and thus act as a major factor contributing to tumor invasion in highly invasive carcinomas?

As for ACs, lower stromal FAP- α and uPA expression positively correlated with tumor invasion and lymph node metastases, indicating that AC cells still possess more control than stromal cells concerning the serine protease production. This hypothesis is supported by results where DPPIV expression in stromal cells was greater in SCCs than in ACs and in cancer cells lower in SCCs than in ACs (Paper I). We showed thus an obvious difference in the serine proteases' biological behavior in two histologically different esophageal carcinomas. This conclusion derives from analyses of both carcinoma cells and stromal cells adjacent to cancer sites, the latter seeming to play a more important role in neoplastic progression than was previously assumed (López-Otín & Matrisian 2007).

Considering the fact that the analyzed serine proteases all belong to one family, that DPPIV and seprase share nearly 50% of sequence identity and that the enzymatic activity assay results performed on esophageal carcinoma cell lines and normal esophageal epithelium were similar for all three enzymes, we presume that the proteases operate in an analogous way regarding ECM degradation. Altogether, these findings point to a complex interaction between the serine proteases themselves and between premalignant, malignant and stromal cells in the development of neoplastic process.

The serine protease involvement in tumor progression appears to be evident. Previous studies presented them mostly as tumor promoters (Hewin et al. 1996, Cheng et al. 1998, Iwasa et al. 2003). However, contradictory findings concerning DPPIV, exposing it as a tumor suppressor, were also reported (Wesley et al. 1999, Pethiyagoda et al. 2000, Kikkawa et al. 2005), whereas seprase and uPA were only associated with an increased tumorigenicity. In our study, we observed that high uPA and DPPIV expressions in SCC cells correlated with a better prognosis (Paper II), which corresponds to the tumor suppressor hypothesis. However, this finding has to be interpreted with caution, as we cannot exclude the possibility that even stromal serine protease in carcinomas may significantly contribute to preserving their malignancy and thus tumor development. In such a case, it is important that not only

malignant but also stromal cells adjacent to lesions be analyzed for protease expression.

In our study we have analyzed three proteins considered to be important in tumor invasion and as prognostic markers. However, many other molecular factors are involved in neoplastic progression. Recently, many potential predictive and / or prognostic markers for esophageal carcinoma have been identified (Vallböhmer & Lenz 2006). For example, the p53 molecule, with major functions in regulating apoptosis and in G₁-S cell cycle transition, is early and often mutated in SCCs and ACs (Shimada et al. 2000). Rb and p16, central regulators of the G₁ cell cycle check point, are also altered in both histological esophageal carcinoma subtypes (Roncalli et al. 1998, Sturm et al. 2001). Human epidermal growth factor receptor 1 seems to be a reliable prognostic and predictive marker in esophageal cancer patients (Mendelsohn & Baselga 2000 & 2006, Miyazano et al. 2004). Alterations of other markers, such as angiogenetic (b-FGF, TGF- α , TGF- β , VEGF and Cox-2) or apoptotic (Bax) markers are frequently associated with the overall outcome of SCCs and ACs (Inoue et al. 1997, Shih et al. 2000, Aloia et al. 2001, Ikeguchi et al. 2001, Sturm et al. 2001, Shimada et al. 2001, Ogata et al. 2003, Fukai et al. 2003, Kuo et al. 2003, Kleespies et al. 2004, Han et al. 2005). As for the transmembrane protease family, MMP-2, MMP-7 and MMP-9 were studied in esophageal cancer and found to be negative prognostic indicators (Tanioka et al. 2003, Ishibashi et al. 2004, Sharma et al. 2004). Thus, despite the indisputable biological importance, the value of these and several other investigated molecules as clinical biomarkers for prognosis, prediction and early detection is still under evaluation.

Clinical study interpretation

The trend in the incidence rates of esophageal cancer generally increased in the last two decades in many Western countries. While rates for esophageal SCC remained stable or increased slightly, occurrence of AC in the distal esophagus augmented significantly. At the time of diagnosis, most esophageal cancer patients already suffer metastatic disease (Layke & Lopez 2006, Stoner et al. 2007). This results in a poor prognosis: only one of five esophageal cancer patients survives more than three years after initial diagnosis (Younes et al. 2002, Polednak 2003, Stoner et al. 2007).

We observed a distinct trend of increasing esophageal AC and decreasing SCC in the referred population during the last 20 years. In order to evaluate the rate, risk factors, advancement and treatment of both esophageal carcinomas, we analyzed the clinical data and cancer tissue samples of 347 patients treated in our hospital between 1988 and 2007. We wanted to know if the change in carcinoma type influenced survival from esophageal cancer in the last two decades.

Our observations confirmed the previously mentioned tendency of increasing esophageal AC and slightly decreasing SCC. This is consistent with other studies in Europe and in the USA (Devesa et al. 1998, El-Serag et al. 2002, Brown et al. 2008). We found that over one-fourth of patients with AC reported a Barrett's esophagus

history, mostly connected to GERD symptoms. Also, AC was diagnosed more frequently among young patients (30-55 years) (Paper IV). As for SCC, tobacco and alcohol were the two most recognized risk factors. This significant change in tumor type seems to be strongly related to changes in Norwegian lifestyle. In the last decades, the total number of smokers constantly diminished, whereas increasingly more Norwegians suffer from obesity (FHI 2004 & 2007, SSB 2008). Weight gain has been postulated to enhance gastroesophageal reflux, which can lead to a sequence of changes from Barrett's metaplasia to dysplasia and cancer.

In Norway, treatment of GERD has been taken seriously, and considerable resources have been allocated to this problem. If the diagnosis is made, treatment with proton pump inhibitors (PPI) is refunded by the health authorities. Endoscopy and regular follow-up with biopsies of Barrett's changes and dysplasia are recommended as well as surgery when drug therapy does not alleviate symptoms or mucosal changes develop further. However, despite these efforts, while surgical and medical treatment of the symptoms is generally available and almost free of charge, we could not demonstrate any significant improvement, neither in the survival of esophageal cancer in general or in the subgroup of AC.

Identification of genetic changes within families with increasing cancer incidence is important, and diagnostic methods should focus on the development of strategies for early detection of neoplastic malformations, using specific markers emerging in precancerous stages. This could give us information about the disease advancement and thus help us to determine an appropriate local treatment.

Since we do not yet have any specific molecular markers characteristic for esophageal carcinoma, we should focus on the serine proteases which seemed to play an important role in the progress of esophageal malignancies. The detection of both FAP- α and uPA in precancerous stages of AC (Paper III) confirmed previous results describing the involvement of serine proteases in neoplastic progression from its very beginning and their proteolytic activity already in full progress in metaplastic and dysplastic cells. Interestingly, we also detected expression of seprase, DPPIV and uPA in stromal tissue adjacent to dysplastic sites. The staining intensity in stroma increased in parallel with the changes in dysplastic cells, with strongest stromal staining in grade III dysplasia (Paper II). This finding corroborates a coherence between stromal and dysplastic cells in cancer evolution. Here, the question is rising: is it possible to keep precancerous stages under surveillance by monitoring the molecular marker levels in biopsies? An affirmative answer should be formulated carefully, as much more research is needed to define to which degree the proteases are involved in the evolution of precancerous stages.

8. CONCLUDING REMARKS AND PERSPECTIVES

The low survival rate of esophageal cancer has not been improved during the last decade.

ACs have replaced SCCs as the most common type of esophageal cancer being treated with curative intention. This has not improved the low long-time survival rate from esophageal cancer, neither in our reference hospital nor in the national population. Improved neo-adjuvant regimens seem to be able to improve the survival rate significantly, but not to a large extent and with a considerably increase in cost and morbidity. Without a change towards early diagnosis, most patients will be candidates for palliative treatment. Our clinical results demonstrate that, as long as dysphagia and dysphagia-related symptoms are the diagnostic starting points, this will more or less continue to be the case.

We firmly believe that early diagnosis, combined with local treatment regimens, is the best strategy for improvement. The rapid increase in ACs in the Western world population is closely connected to symptomatic reflux and a slow development of precancerous lesions. Thus, aggressive treatment of reflux, either by means of anti-reflux surgery or efficient drug treatment, is mandatory. Dysplastic lesions are able to continue their development towards cancer, even after efficient anti-reflux therapy. Strict surveillance of such lesions by morphological and biological parameters is essential and should lead to local treatment of areas demonstrating development towards cancer or severe dysplasia. An efficient combination of biological markers is still elusive. The membrane proteins studied in this thesis have given us new knowledge of esophageal cancer development, but alone, they do not have the ability to detect lesions developing towards cancer and would have to be combined with other markers. The endeavour towards the most efficient combination of new and already characterized markers will have to continue.

9. REFERENCE LIST

1. Abbott CA, Yu DM, Woollatt E, Sutherland GR, McCaughan GW, Gorrell MD. Cloning, expression and chromosomal localization of a novel human dipeptidyl peptidase (DPP) IV homolog, DPP8. *Eur J Biochem.* 2000;267:6140-50.
2. Adams R, Hoover WB. Benign tumors of the esophagus. *J Thorac Surg.* 1945;14:279-86.
3. Aloia TA, Harpole DH Jr, Reed CE, Allegra C, Moore MB, Herndon JE 2nd, D'Amico TA. Tumor marker expression is predictive of survival in patients with esophageal cancer. *Ann Thorac Surg.* 2001;72:859-66.
4. al-Sarraf M, Martz K, Herskovic A, Leichman L, Brindle JS, Vaitkevicius VK, Cooper J, Byhardt R, Davis L, Emami B. Progress report of combined chemoradiotherapy versus radiotherapy alone in patients with esophageal cancer: an intergroup study. *J Clin Oncol.* 1997;15:277-84. Erratum in: *J Clin Oncol* 1997;15:866.
5. American Cancer Society. *Cancer Facts and Figures.* Atlanta. 2008.
6. American Joint Committee on Cancer. *AJCC Cancer Staging Manual.* 6th ed. Springer, New York, USA, 2002; pp 91-5.
7. Aoyama A, Chen WT. A 170-kDa membrane-bound protease is associated with expression of invasiveness by human malignant melanoma cells. *Proc Natl Acad Sci USA.* 1990;87:8296-300.
8. Ariga N, Sato E, Ohuchi N, Nagura H, Ohtani H. Stromal expression of fibroblast activation protein/seprase, a cell membrane serine proteinase and gelatinase, is associated with longer survival in patients with invasive ductal carcinoma of breast. *Int. J. Cancer.* 2001;95:67-92.
9. Artym VV, Kindzelski AL, Chen WT, Petty HR. Molecular proximity of seprase and the urokinase-type plasminogen activator receptor on malignant melanoma cell membranes: dependence on beta 1 integrins and cytoskeleton. *Carcinogenesis.* 2002;23:1592-1601.
10. Barr H. Photodynamic therapy in Barrett's oesophagus and early oesophageal cancer. *Acta Endoscopica.* 2003;33:4.
11. Barrett NR. Chronic peptic ulcer of the oesophagus and 'oesophagitis'. *Br J Surg.* 1950;150:175-82.
12. Barrett NR. The lower esophagus lined by columnar epithelium. *Surgery.* 1957;41:881-94.
13. Bauvois B. Transmembrane proteases in focus: diversity and redundancy? *J Leukoc Biol.* 2001;70:11-7.
14. Bedenne L, Michel P, Bouche O, Milan C, Mariette C, Conroy T, Pezet D, Rouillet B, Seitz JF, Herr JP, Paillet B, Arveux P, Bonnetain F, Binquet C. Chemoradiation followed by surgery compared with chemoradiation alone in squamous cancer of the esophagus: FFCD 9102. *J Clin Oncol.* 2007;25:1160-68.
15. Bermpohl F, Löster K, Reutter W, Baum O. Rat dipeptidyl peptidase IV (DPP IV) exhibits endopeptidase activity with specificity for denatured fibrillar collagens. *FEBS Lett.* 1998;428:152-6.
16. Birkedal-Hansen H. Proteolytic remodeling of extracellular matrix. *Curr Opin Cell Biol.* 1995;7:728-35.

17. Blant SA, Ballini JP, Caron CT, Fontollet C, Monnier P, Laurini NR. Evolution of DNA ploidy during squamous cell carcinogenesis in the esophagus. *Dis Esophagus*. 2001;14:178-184.
18. Blot WJ, McLaughlin JK. The changing epidemiology of esophageal cancer. *Semin Oncol*. 1999;5:2-8.
19. Bollschweiler E. Demographic variations in the rising incidence of esophageal adenocarcinoma in white males. *Cancer*. 2001;92:549-55.
20. Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med*. 1997;3:730-7.
21. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem*. 1976;72:248-54.
22. Bremner CG, Lynch VP, Ellis FH Jr. Barrett's esophagus: congenital or acquired? An experimental study of esophageal mucosal regeneration in the dog. *Surgery*. 1970;68:209-16.
23. Brown LM, Devesa SS, Chow WH. Incidence of adenocarcinoma of the esophagus among white Americans by sex, stage, and age. *J Natl Cancer Inst*. 2008;100:1184-7.
24. Brunner G, Preissner KT. Pericellular enzymatic hydrolysis: implications for the regulation of cell proliferation in the vessel wall and the bone marrow. *Blood Coagul Fibrinolysis*. 1994;5:625-39.
25. Bumm R, Wong RJ. More or less surgery for esophageal cancer: extent of lymphadenectomy in esophagectomy for squamous cell esophageal carcinoma. How much is necessary. *Dis Esoph*. 1994;7:151-5.
26. Burke AP, Sobin LH, Shekitka KM, Helwig EB. Dysplasia of the stomach and Barrett esophagus: a follow-up study. *Mod Pathol*. 1991;4:336-41.
27. Burmeister BH, Smithers BM, Gebski V, Fitzgerald L, Simes RJ, Devitt P, Ackland S, Gotlay DC, Joseph D, Millar J, North J, Walpole ET, Denham JW, TTRO Group, AGIT Group. Surgery alone versus chemoradiotherapy followed by surgery for resectable cancer of the oesophagus: a randomised controlled phase III trial. *Lancet Oncol*. 2005;6:659-68.
28. Buskens CJ, Westerterp M, Lagarde SM, Bergmann JJ, ten Kate FJ, van Lanschot JJ. Prediction of appropriateness of local endoscopic treatment for high-grade dysplasia and early adenocarcinoma by EUS and histopathologic features. *Gastrointest Endosc*. 2004;60:703-10.
29. Buttar NS, Wang KK, Lutzke LS, Krishnadath KK, Anderson MA. Combined endoscopic mucosal resection and photodynamic therapy for esophageal neoplasia within Barrett's esophagus. *Gastrointest Endosc*. 2001;54:682-8.
30. Cameron AJ. The history of Barrett esophagus. *Mayo Clin Proc*. 2001;76:94-6.
31. Chaves P, Cardoso P, de Almeida JC, Pereira AD, Leitao CN, Soares J. Non-goblet cell population of Barrett's esophagus: an immunohistochemical demonstration of intestinal differentiation. *Hum Pathol*. 1999;30:1291-5.
32. Chen D, Kennedy A, Wang JY, Zeng W, Zhao Q, Pearl M, Zhang M, Suo Z, Nesland JM, Qiao Y, Ng AK, Hirashima N, Yamane T, Mori Y, Mitsumata M, Ghersi G, Chen WT. Activation of EDTA-resistant gelatinases in malignant human tumors. *Cancer Res*. 2006;66:9977-85.
33. Chen WT, Kelly T. Sepsis complexes in cellular invasiveness. *Cancer Met Rev*. 2003;22:259-69.

34. Chen WT. DPPIV and seprase in cancer invasion and angiogenesis. In: Dipeptidyl Aminopeptidases in Health and Disease. Springer, USA, 2003; pp 197-203.
35. Cheng HC, Abdel-Ghany M, Elble RC, Pauli BU. Lung endothelial dipeptidyl peptidase IV promotes adhesion and metastasis of rat breast cancer cells via tumor cell surface-associated fibronectin. *J Biol Chem.* 1998;273:24207-15.
36. Chiravuri M, Schmitz T, Yardley K, Underwood R, Dayal Y, Huber BT. A novel apoptotic pathway in quiescent lymphocytes identified by inhibition of a post-proline cleaving aminodipeptidase: a candidate target protease, quiescent cell proline dipeptidase. *J Immunol.* 1999;163:3092-9.
37. Cohen BR, Wolf BS, Som M, Janowitz HD. Correlation of manometric, oesophagosopic, and radiological findings in the columnar-lined gullet (Barrett Syndrome). *Gut.* 1963;4:406-12.
38. Collard JM, Otte JB, Fiasse R, Laterre PF, De Kock M, Longueville J, Glineur D, Romagnoli R, Reynaert M, Kestens PJ. Skeletonizing en bloc esophagectomy for cancer. *Ann Surg.* 2001;234:25-32.
39. Corley DA, Buffler PA. Oesophageal and gastric cardia adenocarcinomas: analysis of regional variation using the Cancer Incidence in Five Continents database. *Int J Epidemiol.* 2001;30:1415-25.
40. Cote R, Suster S, Weiss L, Weidner N. *Modern Surgical Pathology (2 Volume Set)*. Ed. Weidner N, London, UK, 2003; chapt. 21.
41. Crissman JD, Gnepp DR, Goodman ML, Hellquist H, Johns ME. Pre-invasive lesions of the upper aerodigestive tract. Histologic definitions and clinical implications (a symposium). *Pathol Annu.* 1987;22:311-53.
42. Crissman JD, Zarbo RJ. Dysplasia, in situ carcinoma, and progression to invasive squamous cell carcinoma of the upper aerodigestive tract. *Am J Surg Pathol.* 1989;13:5-16.
43. Cunningham D, Allum WH, Stenning SP, Thompson JN, Van de Velde CJ, Nicolson M, Scarffe JH, Lofts FJ, Falk SJ, Iveson TJ, Smith DB, Langlely RE, Verma M, Weeden S, Chua YJ, MAGIC Trial Participants. Perioperative chemotherapy versus surgery alone for resectable gastroesophageal cancer. *N Engl J Med.* 2006;355:11-20.
44. Daly JM, Fry WA, Little AG, Winchester DP, McKee RF, Stewart AK, Fremgen AM. Esophageal cancer: results of an American College of Surgeons Patient Care Evaluation Study. *J Am Coll Surg.* 2000;190:562-72.
45. de Dombal FT, Price AB, Thompson H, Williams GT, Morgan AG, Softley A, Clamp SE, Unwin BJ. The British Society of Gastroenterology early gastric cancer/dysplasia survey: an interim report. *Gut.* 1990;31:115-20.
46. de Jonge PJ, Steyerberg EW, Kuipers EJ, Honkoop P, Wolters LM, Kerkhof M, van Dekken H, Siersema PD. Risk factors for the development of esophageal adenocarcinoma in Barrett's esophagus. *Am J Gastroenterol.* 2006;101:1421-9.
47. Devesa SS, Blot WJ, Fraumeni JF Jr. Changing patterns in the incidence of esophageal and gastric carcinoma in the United States. *Cancer.* 1998;83:2049-53.
48. Dong Z, Tang P, Li L, Wang G. The strategy for esophageal cancer control in high-risk areas of China. *Jpn J Clin Oncol.* 2002;32:10-2.

49. Duffy MJ. Do proteases play a role in cancer invasion and metastasis? *Eur J Cancer Clin Oncol.* 1987;23:583-9.
50. Ell C, May A, Gossner L, Pech O, Gunter E, Mayer G, Henrich R, Vieth M, Müller H, Seitz G, Stolte M. Endoscopic mucosal resection of early cancer and high grade dysplasia in Barrett's esophagus. *Gastroenterology.* 2000;118:670-7.
51. El-Rifai W, Harper JC, Cummings OC, Hyytinen ER, Frierson HF, Knuutila S, Powell SM. Consistent genetic alterations in xenografts of proximal stomach and gastro-esophageal junction adenocarcinomas. *Cancer Res.* 1998;58:34-7.
52. El-Serag HB, Mason AC, Petersen N, Key CR. Epidemiological differences between adenocarcinoma of the oesophagus and adenocarcinoma of the gastric cardia in the USA. *Gut.* 2002;50:368-72.
53. Enzinger PC, Mayer RJ. Esophageal cancer. *N Engl J Med.* 2003;349:2241-52.
54. Fenoglio-Preiser CM, Noffsinger AE, Stemmermann GN, Lantz PE, Isaacson PG. *Gastrointestinal Pathology: An Atlas and Text Gastrointestinal Pathology.* Ed. Fenoglio-Preiser CM, Raven Press, New York, USA, 1989; pp 96-100.
55. Fidler IJ, Balch CM. The biology of cancer metastasis and implications for therapy. *Curr Probl Surg.* 1987;24:129-209.
56. Fidler IJ, Ellis LM. The implications of angiogenesis for the biology and therapy of cancer metastasis. *Cell.* 1994;79:185-8.
57. Fidler IJ, Gersten DM, Hart IR. The biology of cancer invasion and metastasis. *Adv Cancer Res.* 1978;28:149-250.
58. Fidler IJ, Hart IR. Biological diversity in metastatic neoplasms: origins and implications. *Science.* 1982;217:998-1003.
59. Fidler IJ. Critical factors in the biology of human cancer metastasis: twenty-eighth G.H.A. Clowes memorial award lecture. *Cancer Res.* 1990;50:6130-8.
60. Fidler IJ. Metastasis: quantitative analysis of distribution and fate of tumor emboli labeled with 125 I-5-iodo-2'-deoxyuridine. *J Natl Cancer Inst.* 1970;45:773-82.
61. Fidler IJ. Understanding bone metastases: the key to the effective treatment of prostate cancer. *Clin Adv Hematol Oncol.* 2003;1:278-9.
62. Fléjou J. Barrett's oesophagus: from metaplasia to dysplasia and cancer. *Gut.* 2005;54:6-12.
63. Folkman J. How is blood vessels growth regulated in normal and neoplastic tissue? G.H.A Clowes memorial award lecture. *Cancer Res.* 1986;46:467-743.
64. Fukai Y, Fukuchi M, Masuda N, Osawa H, Kato H, Nakajima T, Kuwano H. Reduced expression of transforming growth factor-beta receptors is an unfavorable prognostic factor in human esophageal squamous cell carcinoma. *Int J Cancer.* 2003;104:161-6.
65. Fumagalli U. Panel of experts. Resective surgery for cancer of the thoracic esophagus. Results of a Consensus Conference at the VIth World Congress of the International Society for Diseases of the Esophagus. *Dis Esoph.* 1996; suppl. Pp. 30-38.

66. Galipeau PC, Cowan DS, Sanchez CA, Barrett MT, Emond MJ, Levine DS, Rabinovitch PS, Reid BJ. 17p (p53) allelic losses, 4N (G2/tetraploid) populations, and progression to aneuploidy in Barrett's esophagus. *Proc Natl Acad Sci USA*. 1996;93:7081-84.
67. Gao H, Wang LD, Zhou Q, Hong JY, Huang TY, Yang CS. p53 tumor suppressor gene mutation in early esophageal precancerous lesions and carcinoma among high-risk populations in Henan, China. *Cancer Res*. 1994;54:4342-6.
68. Garin-Chesa P, Old LJ, Rettig WJ. Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers. *Proc Natl Acad Sci USA*. 1990;87:7235-9.
69. Ghadirian P. An epidemiologic study of esophageal cancer with particular reference to northern Iran. PhD Thesis. Institute of Cancer Research, Division of Epidemiology, Sutton, Surrey, London 1982.
70. Ghersi G, Dong H, Goldstein LA, Yeh Y, Hakkinen L, Larjava HS, Chen WT. Regulation of fibroblast migration on collagenous matrix by a cell surface peptidase complex. *J Biol Chem*. 2002;277:29231-41.
71. Ghersi G, Zhao Q, Salamone M, Yeh Y, Zucker S, Chen WT. The protease complex consisting of dipeptidyl peptidase IV and seprase plays a role in the migration and invasion of human endothelial cells in collagenous matrices. *Cancer Res*. 2006;66:4652-61.
72. Gillen P, Keeling P, Byrne PJ, West AB, Hennessy TP. Experimental columnar metaplasia in the canine oesophagus. *Br J Surg*. 1988;75:113-5.
73. Goldblum JR. Barrett's esophagus and Barrett's-related dysplasia. *Mod Pathol*. 2003;16:316-24.
74. Goldstein LA, Chen WT. Identification of an alternatively spliced seprase mRNA that encodes a novel intracellular isoform. *J Biol Chem*. 2000;275:2554-9.
75. Gottlinger HG, Funke I, Johnson JP, Gokel JM, Riethmuller G. The epithelial cell surface antigen 17-1A, a target for antibody-mediated tumor therapy: its biochemical nature, tissue distribution and recognition by different monoclonal antibodies. *Int J Cancer*. 1986;38:47-53.
76. Green PH, Fleischauer AT, Bhagat G, Goyal R, Jabri B, Neugut AI. Risk of malignancy in patients with celiac disease. *Am J Med*. 2003;115:191-5.
77. Guo JH, Teng GJ, Zhu GY, He SC, Deng G, He J. Self-expandable stent loaded with 125I seeds: Feasibility and safety in a rabbit model. *Eur J Radiol*. 2007;61:356-61.
78. Guo JH, Teng GJ, Zhu GY, He SC, Fang W, Deng G, Li GZ. Self-expandable esophageal stent loaded with 125I seeds: Initial experience in patients with advanced esophageal cancer. *Radiology*. 2008;247:574-81.
79. Guo YJ. Amplification of int-2 gene in primary esophageal carcinoma and fetal esophageal carcinoma induced by N-methyl-N-benzyl nitrosamine (NMBZA). *Zhonghua Zhong Liu Za Zhi*. 1993;15:91-3.
80. Hajian K. Minimal annual incidence rate of esophageal carcinoma: adult population in Mazandaran province (1992-98), North of Iran. *J Res Med Sci*. 2002;1:18-22.
81. Han B, Liu J, Ma MJ, Zhao L. Clinicopathological significance of heparanase and basic fibroblast growth factor expression in human esophageal cancer. *World J Gastroenterol*. 2005;11:2188-92.
82. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100:57-70.

83. Hansen AP, Petros AM, Meadows RP, Nettesheim DG, Mazar AP, Olejniczak ET, Xu RX, Pederson TM, Henkin J, Fesik SW. Solution structure of the amino terminal fragment of urokinase-type plasminogen activator. *Biochemistry*. 1994;33:4847-64.
84. Hashibe M, Boffetta P, Janout V, Zaridze D, Shangina O, Mates D, Szeszenia-Dabrowska N, Bencko V, Brennan P. Esophageal cancer in Central and Eastern Europe: tobacco and alcohol. *Int J Cancer*. 2007;120:1518-22.
85. He YT, Hou J, Chen ZF, Qiao CY, Song GH, Meng FS, Jin HX, Chen C. Trends in incidence of esophageal and gastric cardia cancer in high-risk areas in China. *Eur J Cancer Prev*. 2008;17:71-76.
86. Heath EI, Limburg PJ, Hawk ET, Forastiere AA. Adenocarcinoma of the esophagus: Risk factors and prevention. *Oncology*. 2000;4:507-14.
87. Herskovic A, Martz K, Al-Sarraf M, Leichman L, Brindle J, Vaitkevicius V, Cooper J, Byhardt R, Davis L, Emami B. Combined chemotherapy and radiotherapy compared with radiotherapy alone in patients with cancer of the esophagus. *N Engl J Med*. 1992;326:1593-8.
88. Hewin DF, Savage PB, Alderson D, Vipond MN. Plasminogen activators in oesophageal carcinoma. *Br J Surg*. 1996;83:1152-5.
89. Hewin DF. Randomized comparison of polyglycolic acid and polyglyconate sutures for abdominal fascial closure after laparotomy in patients with suspected impaired wound healing. *Br J Surg*. 1995;82:1698-9.
90. Hewitt R, Danø K. Stromal cell expression of components of matrix-degrading protease systems in human cancer. *Enzyme Protein*. 1996;49:163-73.
91. Hirai K, Kotani T, Aratake Y, Ohtaki S, Kuma K. Dipeptidyl peptidase IV (DPP IV/CD26) staining predicts distant metastasis of 'benign' thyroid tumor. *Pathol Int*. 1999;49:264-5.
92. Hirota WK, Loughney TM, Lazas DJ, Maydonovitch CL, Rholi V, Wong RK. Specialized intestinal metaplasia, dysplasia, and cancer of the esophagus and esophagogastric junction: prevalence and clinical data. *Gastroenterology*. 1999;116:277-85.
93. Hiyama T, Yokozaki H, Kitadai Y, Haruma K, Yasui W, Kajiyama G, Tahara E. Overexpression of human telomerase RNA is an early event in oesophageal carcinogenesis. *Virchows Arch*. 1999;434:483-7.
94. Hollstein MC, Peri L, Mandard AM, Welsh JA, Montesano R, Metcalf RA, Bak M, Harris CC. Genetic analysis of human esophageal tumors from two high incidence geographic areas: frequent p53 base substitutions and absence of ras mutations. *Cancer Res*. 1991;51:4102-6.
95. Hollstein MC, Smits AM, Galiana C, Yamasaki H, Bos JL, Mandard A, Partensky C, Montesano R. Amplification of epidermal growth factor receptor gene but no evidence of ras mutations in primary human esophageal cancers. *Cancer Res*. 1988;48:5119-23.
96. Holmes RS, Vaughan TL. Epidemiology and pathogenesis of esophageal cancer. *Semin Radiat Oncol*. 2007;17:2-9.
97. Hölscher AH, Bollschweiler E, Schneider PM, Siewert JR. Early adenocarcinoma in Barrett's oesophagus. *Br J Surg*. 1997;84:1470-3.

98. Hölscher AH, Bollschweiler E, Schröder W, Gutschow C, Siewert J. Prognostic differences between early squamous-cell and adenocarcinoma of the esophagus. *Dis Esophagus*. 1997;10:179-84.
99. Hopsu-Havu VK, Glenner GG. A new dipeptide naphthylamidase hydrolyzing glycyl-prolyl-beta-naphthylamide. *Histochemie*. 1966;7:197-201.
100. Hosch SB, Stoecklein NH, Izbicki JR. Molecular markers and staging of early esophageal cancer. *Langenbecks Arch Surg*. 2003;388:77-82.
101. Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. *Cell*. 1992;69:11-25.
102. Iacone C, DeMeester TR, Little AG, Skinner DB. Barrett's esophagus. Functional assessment, proposed pathogenesis, and surgical therapy. *Arch Surg*. 1983;118:543-9.
103. Ikeguchi M, Maeta M, Kaibara N. Bax expression as a prognostic marker of postoperative chemoradiotherapy for patients with esophageal cancer. *Int J Mol Med*. 2001;7:413-17.
104. Inoue H. Endoscopic mucosal resection for esophageal and gastric mucosal cancers. *Can J Gastroenterol*. 1998;12:355-9.
105. Inoue K, Ozeki Y, Suganuma T, Sugiura Y, Tanaka S. Vascular endothelial growth factor expression in primary esophageal squamous cell carcinoma. Association with angiogenesis and tumor progression. *Cancer* 1997; 79: 206-13.
106. Ishibashi Y, Matsumoto T, Niwa M, Suzuki Y, Omura N, Hanyu N, Nakada K, Yanaga K, Yamada K, Ohkawa K, Kawakami M, Urashima M. CD147 and matrix metalloproteinase-2 protein expression as significant prognostic factors in esophageal squamous cell carcinoma. *Cancer*. 2004;101:1994-2000.
107. Iwasa S, Jin X, Okada K, Mitsumata M, Ooi A. Increased expression of seprase, a membrane-type serine protease, is associated with lymph node metastasis in human colorectal cancer. *Cancer Letters*. 2003;199:91-98.
108. Jiang W, Zhang YJ, Kahn SM, Hollstein MC, Santella RM, Lu SH, Harris CC, Montesano R, Weinstein IB. Altered expression of the cyclin D1 and retinoblastoma genes in human esophageal cancer. *Proc Natl Acad Sci USA*. 1993;90:9026-30.
109. Johnson RC, Zhu D, Augustin-Voss HG, Pauli BU. Lung endothelial dipeptidyl peptidase IV is an adhesion molecule for lung-metastatic rat breast and prostate carcinoma cells. *J Cell Biol*. 1993;121:1423-32.
110. Kajiyama H, Kikkawa F, Suzuki T, Shibata K, Ino K, Mizutani S. Prolonged survival and decreased invasive activity attributable to dipeptidyl peptidase IV overexpression in ovarian carcinoma. *Cancer Res*. 2002;62:2753-7.
111. Kelly T, Mueller SC, Yeh Y, Chen WT. Invadopodia promote proteolysis of a wide variety of extracellular matrix proteins. *J Cell Physiol*. 1994;158:299-308.
112. Kelly T. Fibroblast activation protein-alpha and dipeptidyl peptidase IV (CD26): cell-surface proteases that activate cell signaling and are potential targets for cancer therapy. *Drug Resist Updat*. 2005;8:51-8.
113. Keppler D, Abrahamson M, Sordat B. Secretion of cathepsin B and tumour invasion. *Biochem Soc Trans*. 1994;22:43-9.

114. Kerbel RS. Growth dominance of the metastatic cancer cell: cellular and molecular aspects. *Adv Cancer Res.* 1990;55:87-132.
115. Kikkawa F, Kajiyama H, Shibata K, Ino K, Nomura S, Mizutani S. Dipeptidyl peptidase IV in tumor progression. *Biochim Biophys Acta.* 2005;1751:45-51.
116. Killinger WA, Rice TW, Adelstein DJ, Medendorp SV, Zuccaro G, Kirby TJ, Goldblum JR. Stage II esophageal carcinoma. The significance of T and N. *J Thorac Cardiovasc Surg.* 1996;111:935-40.
117. Kimura Y, Shiozaki H, Doki Y, Yamamoto M, Utsunomiya T, Kawanishi K, Fukuchi N, Inoue M, Tsujinaka T, Monden M. Cytoplasmic beta-catenin in esophageal cancers. *Int J Cancer.* 1999;84:174-8.
118. Kleespies A, Guba M, Jauch KW, Bruns CJ. Vascular endothelial growth factor in esophageal cancer. *J Surg Oncol* 2004;87:95–104.
119. Klein CA, Blankenstein TJ, Schmidt-Kittler O, Petronio M, Polzer B, Stoecklein NH, Riethmuller G. Genetic heterogeneity of single disseminated tumour cells in minimal residual cancer. *Lancet.* 2002;360:683-89.
120. Kleinberg LR, Forastiere AA, Heitmiller RF. Cancer of the Esophagus. In: *Clinical Oncology*. Eds. Abloff MD, Armitage JO, Lichter AS, Niederhuber JE, Kastan MB, McKenna WG, Elsevier, Philadelphia, USA, 2004; pp 1787-1818.
121. Kodama M, Kakegawa T. Treatment of superficial cancer of the esophagus: a summary of responses to a questionnaire on superficial cancer of the esophagus in Japan. *Surgery.* 1998;123:432-9.
122. Koppert L, Wijnhoven B, van Dekken H, Tilanus H, Dinjens W. The molecular biology of esophageal adenocarcinoma. *J Surg Oncol.* 2005;92: 169-90.
123. Koshy M, Esiashvilli N, Landry JC, Thomas CR Jr, Matthews RH. Multiple management modalities in esophageal cancer. *Oncologist.* 2004;9:137-146 and 147-159.
124. Kuo KT, Chow KC, Wu YC. Clinicopathologic significance of cyclooxygenase-2 overexpression in esophageal squamous cell carcinoma. *Ann Thorac Surg.* 2003;76:909–14.
125. Kuwano H, Kato H, Miyazaki T, Fukuchi M, Masuda N, Nakajima M, Fukai Y, Sohda M, Kimura H, Faried A. Genetic Alterations in Esophageal Cancer. *Surg Today.* 2005;35:7-18.
126. Kuwano H, Nagamatsu M, Ohno S, Matsuda H, Mori M, Sugimachi K. Coexistence of intraepithelial carcinoma and glandular differentiation in esophageal squamous cell carcinoma. *Cancer.* 1988;62:1568-72.
127. Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature.* 1970;227:680-5.
128. Lagergren J, Bergström R, Lindgren A, Nyrén O. Symptomatic gastroesophageal reflux as a risk factor for esophageal adenocarcinoma. *N Engl J Med.* 1999;340:825-31.
129. Lam AK. Molecular biology of esophageal squamous cell carcinoma. *Crit Rev Oncol Hematol.* 2000;33:71-90.
130. Lambert R. Endoscopic detection and treatment of early esophageal cancer: a critical analysis. *Endoscopy.* 1995;27:58-9.

131. Layke JC, Lopez PP. Esophageal cancer: a review and update. *Am Fam Physician*. 2006;73:2187-94.
132. Lerut T, Coosemans W, Decker G, De Leyn P, Moons J, Nafteux P, Van Raemdonck D. Extended surgery of cancer of the esophagus and gastroesophageal junction. *J Surg Res*. 2004;117:58-63.
133. Levine DS, Haggitt RC, Blount PL, Rabinovitch PS, Rusch VW, Reid BJ. An endoscopic biopsy protocol can differentiate high-grade dysplasia from early adenocarcinoma in Barrett's esophagus. *Gastroenterology*. 1993;105:40-50.
134. Levy MT, McCaughan GW, Abbott CA, Park JE, Cunningham AM, Müller E, Rettig WJ, Gorrell MD. Fibroblast activation protein: a cell surface dipeptidyl peptidase and gelatinase expressed by stellate cells at the tissue remodelling interface in human cirrhosis. *Hepatology*. 1999;29:1768-78.
135. Lewin KJ, Appelman HD. Barrett's esophagus, columnar dysplasia and adenocarcinoma of the esophagus. In: *Tumors of the esophagus and stomach*. Eds. Lewin KJ, Appelman HD, Armed Forces Institute of Pathology, Washington DC, USA, 1996; pp 99-137.
136. Lewin KJ, Appelman HD. Carcinoma of the stomach. In: *Tumors of the esophagus and stomach*. Eds. Lewin KJ, Appelman HD, Armed Forces Institute of Pathology, Washington DC, USA, 1996; pp 245-321.
137. Lewin KJ. Nomenclature problems of gastrointestinal epithelial neoplasia. *Am J Surg Pathol*. 1998;22:1043-7.
138. Liotta LA, Kohn EC. The microenvironment of the tumour-host interface. *Nature*. 2001;411:375-9.
139. Liotta LA. Tumor invasion and metastases-role of the extracellular matrix. Rhoads Memorial Award lecture. *Cancer Res*. 1986;46:1-7.
140. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods*. 2001;25:402-8.
141. López-Otín C, Matrisian LM. Emerging roles of proteases in tumour suppression. *Nat Rev Cancer*. 2007;7:800-8.
142. Lu JB, Yang WX, Dong WZ, Sang JY. A prospective study of esophageal cytological atypia in Linxian county. *Int J Cancer*. 1988;41:805-8.
143. Lu SH, Hsieh LL, Luo FC, Weinstein IB. Amplification of the EGF receptor and c-myc genes in human esophageal cancers. *Int J Cancer*. 1988;42:502-5.
144. Lu SH. Alterations of oncogenes and tumor suppressor genes in esophageal cancer in China. *Mutat Res*. 2000;462:343-53.
145. MacDougall JR, Matrisian LM. Contributions of tumor and stromal matrix metalloproteinases to tumor progression, invasion and metastasis. *Cancer Met Rev*. 1995;14:351-62.
146. Makuuchi H, Machimura T, Mizutani K, Shimada H, Kanno K, Tajima T, Mitomi T. Controversy in the treatment of superficial esophageal carcinoma--indications and problems of the procedures. *Nippon Geka Gakkai Zasshi*. 1992;93:1059-62.
147. Makuuchi H. Esophageal endoscopic mucosal resection (EEMR) tube. *Surg Laparosc Endosc*. 1996;6:160-1.

148. Mandard AM, Hainaut P, Hollstein M. Genetic steps in the development of squamous cell carcinoma of the esophagus. *Mutat Res.* 2000;462:335-42.
149. Mandard AM, Marnay J, Gignoux M, Segol P, Blanc L, Ollivier JM. Cancer of the esophagus and associated lesions: detailed pathologic study of 100 esophagectomy specimens. *Hum Pathol.* 1984;15:660-9.
150. Mathew S, Scanlan MJ, Mohan Raj BK, Murty VV, Garin-Chesa P, Old LJ, Rettig WJ, Chaganti RS. The gene for fibroblast activation protein alpha (FAP), a putative cell surface-bound serine protease expressed in cancer stroma and wound healing, maps to chromosome band 2q23. *Genomics.* 1995;25:335-7.
151. May A, Gossner L, Pech O, Fritz A, Gunter E, Mayer G, Muller H, Seitz G, Vieth M, Stolte M, Ell C. Local endoscopic therapy for intraepithelial high-grade neoplasia and early adenocarcinoma in Barrett's oesophagus: acute-phase and intermediate results of a new treatment approach. *Eur J Gastroenterol Hepatol.* 2002;14:1085-91.
152. May A, Gossner L, Pech O, Müller H, Vieth M, Stolte M, Ell C. Intraepithelial high-grade neoplasia and early adenocarcinoma in short-segment Barrett's esophagus (SSBE): curative treatment using local endoscopic treatment techniques. *Endoscopy.* 2002;34:604-10.
153. Mehlen P, Puisieux A. Metastasis: a question of life or death. *Nat Rev Cancer.* 2006;6:449-58.
154. Mendelsohn J, Baselga J. Epidermal growth factor receptor targeting in cancer. *Semin Oncol.* 2006;4:369-85.
155. Mendelsohn J, Baselga J. The EGF receptor family as targets for cancer therapy. *Oncogene.* 2000;19:6550-65.
156. Merry AH, Schouten LJ, Goldbohm RA, van den Brandt PA. Body Mass Index, height and risk of adenocarcinoma of the oesophagus and gastric cardia: a prospective cohort study. *Gut.* 2007;56:1503-11.
157. Mills SE, Gaffey MJ, Frierson HF Jr. Conventional squamous cell carcinoma. In: Tumors of the upper aerodigestive tract and ear. Atlas of tumor pathology. Eds. Rosai J, Sobin LW, Armed Forces Institute of Pathology, Washington DC, USA, 2000; pp 45-70.
158. Miyazono F, Metzger R, Warnecke-Eberz U, Baldus SE, Brabender J, Bollschweiler E, Doerfler W, Mueller RP, Dienes HP, Aikou T, Hoelscher AH, Schneider PM. Quantitative c-erbB-2 but not c-erbB-1 mRNA expression is a promising marker to predict minor histopathologic response to neoadjuvant radiochemotherapy in oesophageal cancer. *Br J Cancer.* 2004;91:666-72.
159. Monsky WL, Lin CY, Aoyama A, Kelly T, Akiyama SK, Mueller SC, Chen WT. A potential marker protease of invasiveness, seprase, is localized on invadopodia of human malignant melanoma cells. *Cancer Res.* 1994;54:5702-10.
160. Mori Y, Kono K, Matsumoto Y, Fujii H, Yamane T, Mitsumata M, Chen WT. The expression of type II transmembrane serine protease (seprase) in human gastric carcinoma. *Oncology.* 2004;67:411-419.
161. Morimoto C, Lord CI, Zhang C, Duke-Cohan JS, Letvin NL, Schlossman SF. Role of CD26/dipeptidyl peptidase IV in human immunodeficiency virus type 1 infection and apoptosis. *Proc Natl Acad Sci USA.* 1994;91:9960-4.
162. Morimoto C, Schlossman SF. CD 26: A key costimulatory molecule on CD4 memory T cells. *The Immunologist* 1994;2:4-7.

163. Morino N, Mimura T, Hamasaki K, Tobe K, Ueki K, Kikuchi K, Takehara K, Kadowaki T, Yazaki Y, Nojima Y. Matrix/integrin interaction activates the mitogen-activated protein kinase, p44erk-1 and p42erk-2. *J Biol Chem.* 1995;270:269-73.
164. Moskaluk CA, Hu J, Perlman EJ. Comparative genomic hybridization of esophageal and gastroesophageal adenocarcinomas shows consensus areas of DNA gain and loss. *Genes Chromosomes Cancer.* 1998;22:305-11.
165. Mueller SC, Ghersi G, Akiyama SK, Sang QX, Howard L, Pineiro-Sanchez M, Nakahara H, Yeh Y, Chen WT. A novel protease-docking function of integrin at invadopodia. *J Biol Chem.* 1999;274:24947-52.
166. Mukada T, Sasano N, Sato E. Evaluation of esophageal dysplasia by cytofluorometric analysis. *Cancer.* 1978;41:1399-404.
167. Munoz N, Crespi M, Grassi A, Qing WG, Qiong S, Cai LZ. Precursor lesions of oesophageal cancer in high-risk populations in Iran and China. *Lancet.* 1982;1:876-9.
168. Naef AP, Savary M. Conservative operations for peptic esophagitis with stenosis in columnar-lined lower esophagus. *Ann Thorac Surg.* 1972;13:543-51.
169. Nakagawa S, Nishimaki T, Kanda T, Suzuki T, Hatakeyama K. Composite tumor with papillary adenocarcinoma and squamous cell carcinoma of the esophagus: report of a case. *Surg Today.* 2000;30:364-7.
170. Nakajima S, Hattori T. Oesophageal adenocarcinoma or gastric cancer with or without eradication of *Helicobacter pylori* infection in chronic atrophic gastritis patients: a hypothetical opinion from a systematic review. *Aliment Pharmacol Ther.* 2004;20:54-61.
171. Nasjonalt folkehelseinstitutt (FHI). Kroppsmasseindeks og vurdering av vekt (Body mass index and weight assessment). www.fhi.no, Oslo, Norway, 2004/2007.
172. Natali PG, Nicotra MR, Botti C, Mottolese M, Bigotti A, Segatto O. Changes in expression of alpha 6/beta 4 integrin heterodimer in primary and metastatic breast cancer. *Br J Cancer.* 1992;66:318-22.
173. National Cancer Institute. Incidence and mortality rate trends. A snapshot of esophageal cancer 2006.
174. Nekarda H, Schlegel P, Schmitt M, Stark M, Mueller DJ, Fink U, Siewert JR. Strong prognostic impact of tumor-associated urokinase-type plasminogen activator in completely resected adenocarcinoma of the esophagus. *Clin Cancer Res.* 1998;4:1755-63.
175. Nicolson GL. Cancer metastasis: tumor cell and host organ properties important in metastasis to specific secondary sites. *Biochim Biophys Acta.* 1988;948:175-224.
176. Nigro JJ, Hagen JA, DeMeester TR, DeMeester SR, Theisen J, Peters JH, Kiyabu M. Occult esophageal adenocarcinoma: extent of disease and implications for effective therapy. *Ann Surg.* 1999;230:433-8.
177. Nishino N, Aoki K, Tokura Y, Sakaguchi S, Takada Y, Takada A. The urokinase type of plasminogen activator in cancer of digestive tract. *Thromb Res.* 1988;50:527-35.
178. Norton JA. Surgery: Basic Science and Clinical Evidence. Eds. Norton JA, Barie PS, Bollinger RR, Chang AE, Lowry SF, Mulvihill SJ, Pass HI, Thompson RW. Springer Verlag, Berlin, Germany, 2000; p 456.

179. Oberg S, Wenner J, Johansson J, Walther B, Willen R. Barrett esophagus: Risk factors for progression to dysplasia and adenocarcinoma. *Ann Surg.* 2005;242:49-54.
180. Oehlert W, Keller P, Henke M, Strauch M. Gastric mucosal dysplasia: what is its clinical significance? *Front Gastrointest Res.* 1979;4:173-82.
181. Ogata Y, Fujita H, Yamana H, Sueyoshi S, Shirouzu K. Expression of vascular endothelial growth factor as a prognostic factor in node-positive squamous cell carcinoma in the thoracic esophagus: long-term follow-up study. *World J Surg.* 2003;27:584-9.
182. Okada K, Chen WT, Iwasa S, Jin X, Yamane T, Ooi A, Mitsumata M. Seprase, a membrane-type serine protease, has different expression patterns in intestinal- and diffuse-type gastric cancer. *Oncology.* 2003;65:363-70.
183. Orringer MB. Technical aids in performing transhiatal esophagectomy without thoracotomy. *Ann Thorac Surg.* 1984;38:128-32.
184. Ożóg J, Jarzab M, Pawlaczek A, Oczko-Wojciechowska M, Włoch J, Roskosz J, Gubała E. Expression of DPP4 gene in papillary thyroid carcinoma. *Endokrynol Pol.* 2006;57:12-17.
185. Paget S. The distribution of secondary growths in cancer of the breast. *Lancet.* 1889;1:571-3.
186. Park JE, Lenter MC, Zimmermann RN, Garin-Chesa P, Old LJ, Rettig WJ. Fibroblast activation protein, a dual specificity serine protease expressed in reactive human tumor stromal fibroblasts. *J Biol Chem.* 1999;274:36505-12.
187. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin.* 2005;55:74-108.
188. Pera M, Cameron AJ, Trastek VF, Carpenter HA, Zinsmeister AR. Increasing incidence of adenocarcinoma of the esophagus and esophagogastric junction. *Gastroenterology.* 1993;104:510-3.
189. Pera M, Manterola C, Vidal O, Grande L. Epidemiology of esophageal adenocarcinoma. *J Surg Oncol.* 2005;92:151-9.
190. Peters JH, Clark GW, Ireland AP, Chandrasoma P, Smyrk TC, DeMester TR. Outcome of adenocarcinoma arising in Barrett's esophagus in endoscopically surveyed and nonsurveyed patients. *J Thorac Cardiovasc Surg.* 1994;108:813-22.
191. Pethiyagoda CL, Welch DR, Fleming TP. Dipeptidyl peptidase IV (DPPIV) inhibits cellular invasion of melanoma cells. *Clin Exp Metastasis.* 2000;18:391-400.
192. Petterson EJ. Benign neoplasm of the esophagus: rapport of a case of myxofibroma. *Ann Otol Rhino Laryngol.* 1932;41:942-50.
193. Piazza GA, Callanan HM, Mowery J, Hixson DC. Evidence for a role of dipeptidyl peptidase IV in fibronectin-mediated interactions of hepatocytes with extracellular matrix. *Biochem J.* 1989;262:327-34.
194. Piñeiro-Sánchez ML, Goldstein LA, Dodt J, Howard L, Yeh Y, Tran H, Argraves WS, Chen WT. Identification of the 170-kDa melanoma membrane-bound gelatinase (seprase) as a serine integral membrane protease. *J Biol Chem.* 1997;272:7595-601.
195. Polednak AP. Trends in survival for both histologic types of esophageal cancer in US surveillance, epidemiology and end results areas. *Int J Cancer.* 2003;105:98-100.

196. Posner MC, Forastiere AA, Minsky B. Cancer of the Esophagus. In: *Cancer: Principles and Practice of Oncology*. Eds. DeVita VT, Hellman S, Rosenberg SA, Lippincott-Williams & Wilkins, Philadelphia, USA, 2005; pp 861-908.
197. Raida M, Sarbia M, Clement JH, Adam S, Gabbert HE, Höffken K. Expression, regulation and clinical significance of bone morphogenetic protein 6 in esophageal squamous-cell carcinoma. *Int J Cancer*. 1999;83:38-44.
198. Reid BJ, Blount PL, Rubin CE, Levine DS, Haggitt RC, Rabinovitch PS. Flow-cytometric and histological progression to malignancy in Barrett's esophagus: prospective endoscopic surveillance of a cohort. *Gastroenterology*. 1992;102:1212-19.
199. Reid BJ, Levine DS, Longton G, Blount PL, Rabinovitch PS. Predictors of progression to cancer in Barrett's esophagus: baseline histology and flow cytometry identify low- and high-risk patient subsets. *Am J Gastroenterol*. 2000;95:1669-76.
200. Reid BJ, Weinstein WM, Lewin KJ, Haggitt RC, VanDeventer G, Den Besten L, Rubin CE. Endoscopic biopsies diagnose high-grade dysplasia or early operable adenocarcinoma in Barrett's esophagus without grossly recognizable neoplastic lesions. *Gastroenterology*. 1988;94:81-90.
201. Rettig WJ, Garin-Chesa P, Beresford HR, Oettgen HF, Melamed MR, Old LJ. Cell-surface glycoproteins of human sarcomas: differential expression in normal and malignant tissues and cultured cells. *Proc Natl Acad Sci USA*. 1988;85:3110-4.
202. Rettig WJ, Garin-Chesa P, Healey JH, Su SL, Ozer HL, Schwab M, Albino AP, Old LJ. Regulation and heteromeric structure of the fibroblast activation protein in normal and transformed cells of mesenchymal and neuroectodermal origin. *Cancer Res*. 1993;53:3327-3335.
203. Rettig WJ, Su SL, Fortunato SR, Scanlan MJ, Mohan Raj BK, Garin-Chesa P, Healey JH, Old LJ. Fibroblast activation protein: purification, epitope mapping and induction by growth factors. *Int J Cancer*. 1994;58:385-92.
204. Rice TW, Blackstone EH, Goldblum JR, DeCamp MM, Murthy SC, Falk GW, Ormsby AH, Rybicki LA, Richter JE, Adelstein DJ. Superficial adenocarcinoma of the esophagus. *J Thorac Cardiovasc Surg*. 2001;122:1077-90.
205. Riddell RH, Goldman H, Ransohoff DF, Appelman HD, Fenoglio CM, Haggitt RC, Ahren C, Correa P, Hamilton SR, Morson BC, et al. Dysplasia in inflammatory bowel disease: standardized classification with provisional clinical applications. *Hum Pathol*. 1983;14:931-68.
206. Robaszekiewicz M, Hardy E, Volant A, Noursbaum JB, Cauvin JM, Calament G, Robert FX, Saleun JP, Gouerou H. Flow cytometric analysis of cellular DNA content in Barrett's esophagus. A study of 66 cases. *Gastroenterol Clin Biol*. 1991;15:703-10.
207. Roncalli M, Bosari S, Marchetti A, Buttitta F, Bossi P, Graziani D, Peracchia A, Bonavina L, Viale G, Coggi G. Cell cycle-related gene abnormalities and product expression in esophageal carcinoma. *Lab Invest*. 1998;78:1049-57.
208. Rubio CA, Liu FS, Zhao HZ. Histological classification of intraepithelial neoplasias and microinvasive squamous carcinoma of the esophagus. *Am J Surg Pathol*. 1989;13:685-90.
209. Rucklidge GJ, Edvardsen K, Bock E. Cell-adhesion molecules and metalloproteinases: a linked role in tumour cell invasiveness. *Biochem Soc Trans*. 1994;22:63-8.
210. Saidi F. The historical basis for the "Esophageal cancer belt of south central Asia". *Archives of Iranian Medicine*. 1999;1:26-7.

211. Sammon AM. Protease inhibitors and carcinoma of the esophagus. *Cancer*. 1998;83:405-8.
212. Sato H, Takino T, Okada Y, Cao J, Shinagawa A, Yamamoto E, Seiki M. A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature*. 1994;370:61-5.
213. Sawhney N, Garrahan N, Douglas-Jones AG, Williams ED. Epithelial-stromal interaction in tumors. A morphologic study of fibroepithelial tumors of the breast. *Cancer*. 1992;7:2115-20.
214. Scanlan MJ, Raj BK, Calvo B, Garin-Chesa P, Sanz-Moncasi MP, Healey JH, Old LJ, Rettig WJ. Molecular cloning of fibroblast activation protein alpha, a member of the serine protease family selectively expressed in stromal fibroblasts of epithelial cancers. *Proc Natl Acad Sci USA*. 1994;91:5657-61.
215. Schlemper RJ, Itabashi M, Kato Y, Lewin KJ, Riddell RH, Shimoda T, Sipponen P, Stolte M, Watanabe H, Takahashi H, Fujita R. Differences in diagnostic criteria for gastric carcinoma between Japanese and western pathologists. *Lancet*. 1997;349:1725-9.
216. Schmidt HG, Riddell RH, Walther B, Skinner DB, Riemann JF. Dysplasia in Barrett's esophagus. *J Cancer Res Clin Oncol*. 1985;110:145-52.
217. Schnell TG, Sontag SJ, Chejfec G. Adenocarcinomas arising in tongues or short segments of Barrett's esophagus. *Dig Dis Sci*. 1992;37:137-43.
218. Sedo A, Malik R, Drbal K, Lisa V, Vlasticova K, Mares V. Dipeptidyl peptidase IV in two human glioma cell lines. *Eur J Histochem*. 2001;45:57-63.
219. Sedo A, Mandys V, Krepela E. Cell membrane-bound proteases: not "only" proteolysis. *Physiol Res*. 1996;45:169-76.
220. Seitz JF, Dahan L, Jacob J, Artru P, Maingon P, Bedenne L, Triboulet JP. French guideline, esophagus cancer. *Gastroenterol Clin Biol*. 2006;30:2S5-2S15.
221. Selaru FM, Zou T, Xu Y, Shustova V, Yin J, Mori Y, Sato F, Wang S, Oлару A, Shibata D, Greenwald BD, Krasna MJ, Abraham JM, Meltzer SJ. Global gene expression profiling in Barrett's esophagus and esophageal cancer: a comparative analysis using cDNA microarrays. *Oncogene*. 2002;21:475-78.
222. Shaheen N, Ransohoff DF. Gastroesophageal reflux, Barrett esophagus, and esophageal cancer: scientific review. *JAMA*. 2002;287:1972-81.
223. Sharma R, Chattopadhyay TK, Mathur M, Ralhan R. Prognostic significance of stromelysin-3 and tissue inhibitor of matrix metalloproteinase-2 in esophageal cancer. *Oncology*. 2004;67:300-9.
224. Shih CH, Ozawa S, Ando N, Ueda M, Kitajima M. Vascular endothelial growth factor expression predicts outcome and lymph node metastasis in squamous cell carcinoma of the esophagus. *Clin Cancer Res*. 2000;6:1161-8.
225. Shimada H, Takeda A, Nabeya Y, Okazumi SI, Matsubara H, Funami Y, Hayashi H, Gunji Y, Kobayashi S, Suzuki T, Ochiai T. Clinical significance of serum vascular endothelial growth factor in esophageal squamous cell carcinoma. *Cancer*. 2001;92:663-9.
226. Shimada Y, Watanabe G, Yamasaki S, Maeda M, Kawabe A, Kaganoi JI, Itami A, Fukumoto M, Kanda Y, Imamura M. Histological response of cisplatin predicts patients' survival in oesophageal cancer and p53 protein accumulation in pretreatment biopsy is associated with cisplatin sensitivity. *Eur J Cancer*. 2000;36:987-93.

227. Sibille A, Lambert R, Souquet JC, Sabben G, Descos F. Long-term survival after photodynamic therapy for esophageal cancer. *Gastroenterology*. 1995;108:337-44.
228. Sier CF, Verspaget HW, Griffioen G, Verheijen JH, Quax PH, Dooijewaard G, De Bruin PA, Lamers CB. Imbalance of plasminogen activators and their inhibitors in human colorectal neoplasia. Implications of urokinase in colorectal carcinogenesis. *Gastroenterology*. 1991;101:1522-8.
229. Siewert JR, Burkhard HA, von Rahden H, Stein HJ. Current status of esophageal cancer - West versus East: the European point of view. *Esophagus*. 2004;1:147-59.
230. Siewert JR, Dittler HJ. Esophageal carcinoma: impact of staging on treatment. *Endoscopy*. 1993;25:28-32.
231. Simone NL, Bonner RF, Gillespie JW, Emmert-Buck MR, Liotta LA. Laser-capture microdissection: opening the microscopic frontier to molecular analysis. *Trends Genet*. 1998;14:272-6.
232. Skinner DB. En bloc resection of neoplasms of the esophagus and cardia. *J Thorac Cardiovasc Surg*. 1983;85:59-71.
233. Sloane BF, Herman CJ, Padarathsingh M. Molecular mechanisms of progression and metastasis of human tumors: a pathology B study section workshop. Working Report from the Division of Research Grants, NIH. *Cancer Res*. 1994;54:5241-5.
234. Smith RRL, Hamilton SR, Boitnott JK, Rogers EL. The spectrum of carcinoma arising in Barrett's esophagus: a clinicopathologic study of 26 patients. *Am J Surg Pathol*. 1984;8:562-73.
235. Soler M, Bosetti C, Franceschi S, Negri E, Zambon P, Talamini R, Conti E, La Vecchia C. Fiber intake and the risk of oral, pharyngeal and esophageal cancer. *Int J Cancer*. 2001;91:283-7.
236. Souza RF, Spechler SJ. Concepts in the prevention of adenocarcinoma of the distal esophagus and proximal stomach. *CA Cancer J Clin*. 2005;55:334-51.
237. Spechler SJ, Goyal RK. Barrett's esophagus. *N Engl J Med*. 1986;315:362-71.
238. Spechler SJ, Goyal RK. The columnar-lined esophagus, intestinal metaplasia and Norman Barrett. *Gastroenterology*. 1996;110:614-21.
239. Spechler SJ. Barrett's esophagus. *Semin Oncol*. 1994;21:431-7.
240. Spechler SJ. Managing Barrett's oesophagus. *BMJ*. 2003;326:892-4.
241. Spencer GM, Thorpe SM, Blackman GM, Solano J, Tobias JS, Lovat LB, Bown SG. Laser augmented by brachytherapy versus laser alone in the palliation of adenocarcinoma of the oesophagus and cardia: a randomised study. *Gut*. 2002;50:224-7
242. Spin FP. Adenomas of the esophagus: a case report and review of the literature. *Gastrointest Endosc*. 1973;1:26-7.
243. Stahl M, Stuschke M, Lehmann N, Meyer HJ, Walz MK, Seeber S, Klump B, Budach W, Teichmann R, Schmitt M, Schmitt G, Franke C, Wilke H. Chemoradiation with and without surgery in patients with locally advanced squamous cell carcinoma of the esophagus. *J Clin Oncol*. 2005;23:2310-17.

244. Statistisk sentralbyrå (SSB). Stadig færre røyker daglig. Røyking i Norge (Continuously decreasing number of smokers. Smoking in Norway). Statistics Norway, www.ssb.no, Oslo, Norway, 2008.
245. Stein HJ, Feith M, Bruecher BL, Naehrig J, Sarbia M, Siewert JR. Early esophageal cancer pattern of lymphatic spread and prognostic factors for long-term survival after surgical resection. *Ann Surg.* 2005;242:566-75.
246. Stein HJ, Feith M, Müller J, Werner M, Siewert JR. Limited resection for early Barrett's cancer. *Ann Surg.* 2000;232:733-42.
247. Stein HJ, Feith M, von Rahden BH, Siewert JR. Approach to early Barrett's cancer. *World J Surg.* 2003;27:1040-6.
248. Stein HJ, Siewert J. Barrett's esophagus: pathogenesis, epidemiology, functional abnormalities, malignant degeneration, and surgical management. *Dysphagia.* 1993;8:276-88.
249. Stewart BW, Kleihues P. World cancer report. IARC. Lyon, 2003.
250. Stoner GD, Gupta A. Etiology and chemoprevention of esophageal squamous cell carcinoma. *Carcinogenesis.* 2001;22:1737-46.
251. Stoner GD, Wang LS, Chen T. Chemoprevention of esophageal squamous cell carcinoma. *Toxicol Appl Pharmacol.* 2007;224:337-49.
252. Streitz JM Jr, Ellis FH Jr, Gibb SP, Balogh K, Watkins E Jr. Adenocarcinoma in Barrett's esophagus. A clinicopathologic study of 65 cases. *Ann Surg.* 1991;213:122-5.
253. Sturm I, Petrowsky H, Volz R, Lorenz M, Radetzki S, Hillebrand T, Wolff G, Hauptmann S, Dörken B, Daniel PT. Analysis of p53/BAX/p16(ink4a/CDKN2) in esophageal squamous cell carcinoma: high BAX and p16(ink4a/CDKN2) identifies patients with good prognosis. *J Clin Oncol.* 2001;19:2272-81.
254. Sugarbaker EV, Thornthwaite J, Ketcham AS. Progress in cancer research and therapy. Eds. Day SB, Myers WPL, Stansly P, Garattini S, Lewis MG, Raven Press, New York, USA, 1977; pp 227-40.
255. Sugarbaker EV. Cancer metastasis: a product of tumor-host interactions. *Curr Probl Cancer.* 1979;3:1-59.
256. Sun S, Albright CF, Fish BH, George HJ, Selling BH, Hollis GF, Wynn R. Expression, purification, and kinetic characterization of full-length human fibroblast activation protein. *Protein Expr Purif.* 2002;24:274-81.
257. Swisher SG, Ajani JA, Komaki RK, Ferguson MK. Neoplasms of the Esophagus. In: Kufe DW, Pollock RE, Weichselbaum RR, Bast RC, Gansler TS, Holland JF, Frei E, eds. *Cancer Medicine.* 6th ed. Hamilton, 2003;1499-1514.
258. Syrjänen KJ. HPV infections and oesophageal cancer. *J Clin Pathol.* 2002;55:721-8.
259. Tajima Y, Nakanishi Y, Ochiai A, Tachimori Y, Kato H, Watanabe H, Yamaguchi H, Yoshimura K, Kusano M, Shimoda T. Histopathologic findings predicting lymph node metastasis and prognosis of patients with superficial esophageal carcinoma: analysis of 240 surgically resected tumors. *Cancer.* 2000;88:1285-93.

260. Takeichi M. Cadherin cell adhesion receptors as a morphogenetic regulator. *Science*. 1991;251:1451-5.
261. Takeichi M. Morphogenetic roles of classic cadherins. *Curr Opin Cell Biol*. 1995;7:619-27.
262. Takino T. Roles of membrane-type 1 matrix metalloproteinase in tumor invasion and progression. *J Oral Bios*. 2007;49:120-7.
263. Tanaka H, Kijima H, Tokunaga T, Tajima T, Himeno S, Kenmochi T, Oshiba G, Kise Y, Nishi T, Chino O, Shimada H, Machimura T, Tanaka M, Makuuch H. Frequent expression of inducible nitric oxide synthase in esophageal squamous cell carcinomas. *Int J Oncol*. 1999;14:1069-73.
264. Tanaka T, Duke-Cohan JS, Kameoka J, Yaron A, Lee I, Schlossman SF, Morimoto C. Enhancement of antigen-induced T-cell proliferation by soluble CD26/dipeptidyl peptidase IV. *Proc Natl Acad Sci USA*. 1994;91:3082-6.
265. Tang W, Gunn TM, McLaughlin DF, Barsh GS, Schlossman SF, Duke-Cohan JS. Secreted and membrane attractin result from alternative splicing of the human ATRN gene. *Proc Natl Acad Sci USA*. 2000;97:6025-30.
266. Tanioka Y, Yoshida T, Yagawa T, Saiki Y, Takeo S, Harada T, Okazawa T, Yanai H, Okita K. Matrix metalloproteinase-7 and matrix metalloproteinase-9 are associated with unfavourable prognosis in superficial oesophageal cancer. *Br J Cancer*. 2003;89:2116-21.
267. Teodori L, Gohde W, Persiani M, Ferrario F, Tirindelli Danesi D, Scarpignato C, Di Tondo U, Alo P, Capurso L. DNA/protein flow cytometry as a predictive marker of malignancy in dysplasia-free Barrett's esophagus: thirteen-year follow-up study on a cohort of patients. *Cytometry*. 1998;34:257-63.
268. Thompson JJ, Zinsser KR, Enterline HT. Barrett's metaplasia and adenocarcinoma of the esophagus and gastroesophageal junction. *Hum Pathol*. 1983;1:42-61.
269. Thurberg BL, Duray PH, Odze RD. Polypoid dysplasia in Barrett's esophagus: a clinicopathologic, immunohistochemical, and molecular study of five cases. *Hum Pathol*. 1999;30:745-52.
270. Torzewski M, Sarbia M, Verreet P, Dutkowski P, Heep H, Willers R, Gabbert HE. Prognostic significance of urokinase-type plasminogen activator expression in squamous cell carcinoma of esophagus. *Clin Cancer Res*. 1997;3:2263-2268.
271. Tosi P, Baak JP, Luzi P, Miracco C, Lio R, Barbini P. Morphometric distinction of low- and high-grade dysplasias in gastric biopsies. *Hum Pathol*. 1989;20:839-44.
272. Tosi P, Luzi P, Baak JP, Miracco C, Vindigni C, Lio R, Barbini P. Gastric dysplasia: a stereological and morphometrical assessment. *J Pathol*. 1987;152:83-94.
273. Traweek ST, Liu J, Battifora H. Keratin gene expression in non-epithelial tissues. Detection with polymerase chain reaction. *Am J Pathol*. 1993;142:1111-18.
274. Trier JS. Morphology of the epithelium of the distal esophagus in patients with midesophageal peptic strictures. *Gastroenterology*. 1970;58:444-61.
275. UICC. TNM Atlas, 5th Edition: An illustrated guide to the TNM/pTNM classification of malignant tumors. Heidelberg, Germany, Springer-Verlag 2005.
276. Vallböhmer D, Lenz HJ. Predictive and prognostic molecular markers in outcome of esophageal cancer. *Dis Esophagus*. 2006;19:425-32.

277. van Dekken H, Geelen E, Dinjens WNM, Wijnhoven BPL, Tilanus H, Tanke HJ, Rosenberg C. Comparative genomic hybridization of cancer of the gastroesophageal junction: Deletion of 14q31-32.1 discriminates between esophageal (Barrett's) and gastric cardia adenocarcinomas. *Cancer Res.* 1999;59:748-52.
278. van Sandick JW, van Lanschot JJ, Kuiken BW, Tytgat GN, Offerhaus GJ, Obertop H. Impact of endoscopic biopsy surveillance of Barrett's oesophagus on pathological stage and clinical outcome of Barrett's carcinoma. *Gut.* 1998;43:216-22.
279. Veuillez V, Rougier P, Seitz JF. The multidisciplinary management of gastrointestinal cancer. Multimodal treatment of oesophageal cancer. *Best Pract Res Clin Gastroenterol.* 2007;21:947-63.
280. Vivier I, Marguet D, Naquet P, Bonicel J, Black D, Li CX, Bernard AM, Gorvel JP, Pierres M. Evidence that thymocyte-activating molecule is mouse CD26 (dipeptidyl peptidase IV). *J Immunol.* 1991;147:447-54.
281. Wang LD, Yang HH, Fan ZM, Lü XD, Wang JK, Liu XL, Sun Z, Jiang YN, He X, Zhou Q. Cytological screening and 15 years' follow-up (1986-2001) for early esophageal squamous cell carcinoma and precancerous lesions in a high-risk population in Anyang County, Henan Province, Northern China. *Acta Cytol.* 2002;46:445-52.
282. Weiss L. Metastasis of cancer: a conceptual history from antiquity to the 1990s. *Cancer Met Rev.* 2000;19:193-383.
283. Weiss L. Metastatic inefficiency: causes and consequences. *Cancer Met Rev.* 1986;3:1-24.
284. Wesley UV, Albino AP, Tiwari S, Houghton AN. A role for dipeptidyl peptidase IV in suppressing the malignant phenotype of melanocytic cells. *J Exp Med.* 1999;190:311-22.
285. WHO Classification of tumours. Pathology and genetics of tumours of the digestive system. In: Chapter I, Tumours of the esophagus. Hamilton SR, Aaltonem LA, eds. IARC Press, Lyon 2000;9-11.
286. WHO. World Health Organization International Histological Classification of Tumors: Histological Typing of Oesophageal and Gastric Tumors. Eds. Watanabe H, Jass JR, Sobin LH, Springer-Verlag, Berlin, Germany, 1990.
287. Wilson MJ, Haller R, Li SY, Slaton JW, Sinha AA, Wasserman NF. Elevation of dipeptidylpeptidase iv activities in the prostate peripheral zone and prostatic secretions of men with prostate cancer: possible prostate cancer disease marker. *J Urol.* 2005;174:1124-8.
288. Xing EP, Nie Y, Wang LD, Yang GY, Yang CS. Aberrant methylation of p16INK4a and deletion of p15INK4b are frequent events in human esophageal cancer in Linxian, China. *Carcinogenesis.* 1999;20:77-84.
289. Xu Y, Selaru FM, Yin J, Zou TT, Shustova V, Mori Y, Sato F, Liu TC, Olaru A, Wang S, Kimos MC, Perry K, Desai K, Greenwald BD, Krasna MJ, Shibata D, Abraham JM, Meltzer SJ. Artificial neural networks and gene filtering distinguish between global gene expression profiles of Barrett's esophagus and esophageal cancer. *Cancer Res.* 2002;62:3493-97.
290. Yang H, Berner A, Mei Q, Giercksky KE, Warloe T, Yang G, Cui J, Suo Z, Zhang S, Nesland JM. Cytologic screening for esophageal cancer in a high-risk population in Anyang County, China. *Acta Cytol.* 2002;46:445-52.
291. Yaron A, Naider F. Proline-dependent structural and biological properties of peptides and proteins. *Crit Rev Biochem Mol Biol.* 1993;28:31-81.

292. Ye W, Held M, Lagergren J, Engstrand L, Blot WJ, McLaughlin JK, Nyren O. Helicobacter pylori infection and gastric atrophy: risk of adenocarcinoma and squamous-cell carcinoma of the esophagus and adenocarcinoma of the gastric cardia. *J Natl Cancer Inst.* 2004;96:388-96.
293. Yee YK, Cheung TK, Chan AO, Yuen MF, Wong BC. Decreasing trend of esophageal adenocarcinoma in Hong Kong. *Cancer Epidemiol Biomarkers Prev.* 2007;16:2637-40.
294. Yokoyama A, Ohmori T, Makuuchi H, Maruyama K, Okuyama K, Takahashi H, Yokoyama T, Yoshino K, Hayashida M, Ishii H. Successful screening for early esophageal cancer in alcoholics using endoscopy and mucosa iodine staining. *Cancer.* 1995;76:928-34.
295. Younes M, Henson DE, Ertan A, Miller CC. Incidence and survival trends of esophageal carcinoma in the United States: racial and gender differences by histological type. *Scand J Gastroenterol.* 2002;37:1359-65.
296. Yu HP, Xu SQ, Liu L, Shi LY, Cai XK, Lu WH, Lu B, Su YH, Li YY. Cyclooxygenase-2 expression in squamous dysplasia and squamous cell carcinoma of the esophagus. *Cancer Lett.* 2003;198:193-201.
297. Zhang G, Kim H, Cai X, López-Guisa JM, Alpers CE, Liu Y, Carmeliet P, Eddy AA. Urokinase receptor deficiency accelerates renal fibrosis in obstructive nephropathy. *J Am Soc Nephrol.* 2003;14:1254-71.
298. Zhang G, Kim H, Cai X, Lopez-Guisa JM, Carmeliet P, Eddy AA. Urokinase receptor modulates cellular and angiogenic responses in obstructive nephropathy. *J Am Soc Nephrol.* 2003;14:1234-53.
299. Zhang X, Chaudhry A, Chintala SK. Inhibition of plasminogen activation protects against ganglion cell loss in a mouse model of retinal damage. *Mol Vis.* 2003;9:238-48.
300. Zimmermann KC, Sarbia M, Weber AA, Borchard F, Gabbert HE, Schrör K. Cyclooxygenase-2 expression in human esophageal carcinoma. *Cancer Res.* 1999;59:198-204.
301. Zucker S. A critical appraisal of the role of proteolytic enzymes in cancer invasion: emphasis on tumor surface proteinases. *Cancer Invest.* 1988;6:219-31.